Localized Primary Cutaneous Nodular Amyloidosis in a Patient With Paraproteinemia *

Amiloidosis nodular primaria cutánea localizada en un paciente con paraproteinemia

To the Editor:

The term primary cutaneous amyloidosis (PCA) refers to a group of diseases caused by the extracellular deposition of amyloid in the skin without the involvement of other organs. PCA has been divided into the following types: macular, lichen and nodular. The first 2 types are characterized by the deposition in the papillary dermis of amyloid derived from the degeneration of keratin filaments. Primary localized cutaneous nodular amyloidosis (PLCNA) is the rarest form of PCA and the only one in which the amyloid deposits are of the amyloid light-chain (AL) type, as in the primary and myeloma-associated systemic forms of amyloidosis. AL amyloidosis is due to monoclonal immunoglobulin (Ig) lightchain deposition.

We report the case of an 83-year-old man with a history of systemic hypertension, type 2 diabetes mellitus, and hyperuricemia, who was seen for a 2-year history of asymptomatic but progressive lesions that had appeared on the left lower limb. One year before consultation, he had been diagnosed with monoclonal gammopathy of undetermined significance (MGUS), after detecting an $IgG(\kappa)$ paraprotein in the serum with no evidence of multiple myeloma in the bone marrow study. Physical examination revealed a plaque with an area of ecchymotic appearance and several hard, reddish-orange nodules with an ulcerated surface in the pretibial region of the left leg (Fig. 1).

Histopathological examination showed deposits of an amorphous eosinophilic material diffusely distributed



Figure 1 Well-defined, shiny, reddish-orange plaque with an area of ecchymotic appearance, and 2 firm nodules covered by reddish-orange skin except for one area of ulceration, located in the left lower limb.

through the papillary and reticular dermis, extending into the subcutaneous tissue, accompanied by a dense infiltrate of plasma cells (Fig. 2). When stained with thioflavin, the deposits fluoresced under ultraviolet light. Immunohistochemical staining demonstrated λ light-chain restriction in the majority of the plasma cells. These findings suggested a possible diagnosis of PLCNA, although systemic amyloidosis could not be ruled out.

Laboratory tests including complete blood count, biochemistry tests, and liver and kidney profiles were normal. Immunoglobulin concentrations were normal, although the IgG value was at the upper limit of normal: 1590 mg/dL (normal range, 700–1600 mg/dL). Immunoelectrophoresis showed a monoclonal IgG(κ) band with oligoclonal λ bands in the serum, but was normal in the urine. No amyloid deposits were observed in a biopsy of the abdominal fat. After 18 months of follow-up, the patient has not shown evidence of progression to myeloma or systemic amyloidosis.

PLCNA is a disease that occurs predominantly in women, with a mean age at diagnosis of 60 years.¹ It has sometimes been described in association with systemic diseases such as Sjögren syndrome,² diabetes mellitus, or CREST syndrome.³ It manifests as waxy nodules that are usually solitary or localized or, less commonly, disseminated.⁴ It tends to affect acral areas, the most common site being the lower limbs, followed by the face and trunk. Histopathological findings are indistinguishable from those of primary systemic amyloidosis or myeloma-associated amyloidosis.⁵ Immunohistochemical staining may show the presence of immunoglobulin light chains in the amyloid or in the cytoplasm of plasma cells.

Based on the demonstration of clonality of the plasma cell infiltrate in the skin using gene rearrangement techniques and on the absence of clonal rearrangement in bone marrow, some authors have suggested that PLCNA should be considered an extramedullary plasmacytoma that produces localized amyloid deposition.⁶ Extramedullary plasmacytomas are plasma cell neoplasms that arise in any organ except bone marrow; they are able to produce an M-component in the serum in up to 20% of cases.⁷ Extramedullary plasmacytomas that initially do not produce the M-component in serum progress to multiple myeloma in 20% of cases.⁸

In our patient, there was a monoclonal infiltrate of plasma cells that synthesized λ chains and produced the amyloid deposits in the skin, and a finding of a monoclonal lgG(κ) band and oligoclonal λ bands in the serum. This suggests a biclonal gammopathy (the presence of 2 monoclonal components in serum), which has been observed in approximately 3% of MGUS.⁹

The appearance of paraproteinemia in cases of PLCNA has rarely been described in the literature, though it should always be investigated at the initial assessment. In the cases in which it has been observed, it was generally associated with an advanced stage of the disease, and may indicate progression to systemic amyloidosis.¹⁰

The risk of progression to systemic amyloidosis in PLCNA has been well defined by several authors. Since 1970, the risk has been established at around 50%. However, in 2001, Woollons and Black⁵ observed only 1 case of progression to systemic amyloidosis in a series of 15 patients with PLCNA, in addition to high immunoglobulin concentrations in 40%

^{*} Please cite this article as: Villar M, et al. Amiloidosis nodular primaria cutánea localizada en un pacinete con paraproteinemia. Actas Dermosifiliogr. 2012;103:161–2.

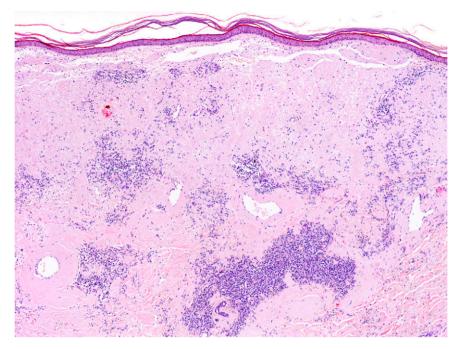


Figure 2 Deposits of large amounts of amyloid in the dermis, mainly around vessels and adnexa, accompanied by a dense plasma cell infiltrate.

of cases. In 2003, Moon et al.¹⁰ obtained similar results after 10 years of follow-up in 16 patients with PLCNA and reported progression in the only patient presenting with paraproteinemia at the time of diagnosis.

Despite its generally benign course, it is important to follow-up these patients closely, especially in cases associated with paraproteinemia, to monitor for possible progression to systemic amyloidosis and to exclude plasma cell dyscrasias.

References

- Schwendiman MN, Beachkofsky TM, Wisco OJ, Owens NM, Hodson DS. Primary cutaneous nodular amyloidosis: case report and review of the literature. Cutis. 2009;84:87–92.
- Chavarría E, González-Carrascosa M, Hernanz JM, Lecona M. Amiloidosis cutánea nodular primaria asociada a síndrome de Sjögren: presentación de un caso. Actas Dermosifiliogr. 2005;96:446-9.
- Shiman M, Ricotti C, Miteva M, Kerdel F, Romanelli P. Primary localized cutaneous nodular amyloidosis associated with CREST (calcinosis, Raynaud's phenomenon, esophageal motility disorders, sclerodactyly, and telangiectasia) syndrome. Int J Dermatol. 2010;49:229–30.
- Feito-Rodríguez M, García-Macarrón J, Pagán-Muñoz B, Mariño-Enríquez A, Vidaurrázaga-Díaz Y, Arcaya C, et al. Amiloidosis cutánea primaria localizada nodular con patrón diseminado. Actas Dermosifiliogr. 2008;99:648–52.
- Woollons A, Black MM. Nodular localized primary cutaneous amyloidosis: a long-term follow-up study. Br J Dermatol. 2001;145:105–9.

- Grünewald K, Sepp N, Weyrer K, Lhotta K, Feichtinger H, Konwalinka G, et al. Gene rearrangement studies in the diagnosis of primary systemic and nodular primary localized cutaneous amyloidosis. J Invest Dermatol. 1991;97:693–6.
- Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, et al. WHO classification of tumors of haematopoietic and lymphoid tissues. 4th ed. Lyon: IARC; 2008.
- Daoud MS, Lust JA, Kyle RA, Pittelkow MR. Monoclonal gammopathies and associated skin disorders. J Am Acad Dermatol. 1999;40:507–35.
- Kyle RA, Therneau TM, Rajkumar SV, Offord JR, Larson DR, Plevak MF, et al. A long-term study of prognosis in monoclonal gammopathy of undetermined significance. N Engl J Med. 2002;21:564–9.
- 10. Moon AO, Calamia KT, Walsh JS. Nodular amyloidosis: review and long term follow-up of 16 cases. Arch Dermatol. 2003;139:1157–9.

M. Villar,^{a,*} M. Burgués,^a J.L. Rodríguez-Peralto,^b R. Rivera,^a F. Vanaclocha^a

^a Servicio de Dermatología, Hospital Universitario 12 de octubre, Madrid, Spain

^b Servicio de Anatomía Patológica, Hospital Universitarion 12 de octubre, Madrid, Spain

* Corresponding author. *E-mail address*: mariavb20@hotmail.com (M. Villar).

Body-Hair Transplant for Cicatricial Alopecia *

Reconstrucción de alopecia cicatricial mediante trasplante de pelo corporal

To the Editor:

Hair transplantation is a continuously evolving field. Since 1959, when Orentreich¹ described the phenomenon of donor dominance in androgenic alopecia, thus paving the way for body-hair transplants, new techniques have not ceased to appear. During the last 15 years, the perception of hair transplantation as an aggressive technique with an artificial result has finally disappeared thanks to the use of follicular units, which have made the technique minimally invasive with mostly acceptable results.²⁻⁴

We present the case of a 30-year-old man with Norwood class III alopecia who underwent hair transplantation at another center in 2008 with the strip method, by means of which he received 3800 follicular units in the frontoparietal region. The follicular units were harvested from the parieto-occipital region.

He consulted 1.5 years after the intervention for assessment of the reconstruction of the donor strip scar $(3 \text{ cm} \times 2 \text{ cm})$ and reinforcement of the frontal area behind the hairline (Fig. 1A and B).

We considered the possibility of using the occipital region as the donor strip. However, the low follicular density (50 units/cm^2) at this site led us to repopulate the frontal area using a combination of follicular units from the occipital region and chest. This approach provided a reasonable number of follicles.

Follicular units were obtained using a 0.8-mm punch after applying local anesthetic with 2% lidocaine with 1:100,000 epinephrine. A total of 335 follicular units were obtained from the chest (Fig. 2A) and 600 from the parieto-occipital region.

Once the recipient site had been anesthetized, the incisions were made in the scar area using a 19G needle, and the body hair follicular units were implanted at a density of 17 units/cm^2 .

Of the 335 follicular units obtained, 100 were implanted in the scar of the parietotemporal area to evaluate the

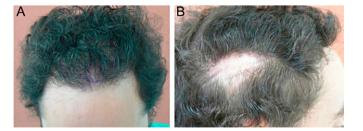


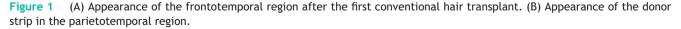
Figure 2 (A) Donor strip on the chest after follicular unit extraction. (B) Implantation of follicular units of body hair in the scar.

response (Fig. 2B), and the remaining body-hair units plus the 600 units from the parieto-occipital region were implanted in the frontal region.

Evaluation at 4 and 6 months after treatment revealed that more than 80% of the units had survived in the scar; the hair was practically indistinguishable from the scalp hair in terms of thickness and length (Fig. 3A and B). The donor area on the chest healed correctly without visible scars or keloids, thus leaving the patient with a very satisfactory cosmetic outcome.

Cicatricial alopecia resulting from burns, surgery, injury, or radiotherapy is an excellent indication for hair transplantation.⁵⁻⁷ Our patient developed an iatrogenic parietal scar after a ''megasession'' of follicle donation, possibly because the donor strip was larger than generally





^{*} Please cite this article as: Meyer-González T, Bisanga C. Reconstrucción de alopecia cicatricial mediante transplante de pelo corporal. Actas Dermosifiliogr. 2012;103:163–4.