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ORIGINAL ARTICLE

Dermoscopy as a Tool for Estimating Breslow Thickness in Melanoma[☆]



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Dermoscopy;
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Abstract

Background and objective: The incidence of melanoma has increased significantly, and early diagnosis is the most effective way to reduce associated deaths. Dermoscopy increases diagnostic accuracy in melanoma and analysis of dermoscopic structures can help in the estimation of tumor thickness. The aim of this study was to analyze the influence of Breslow thickness on the dermoscopic characteristics of melanoma.

Material and methods: Observational, cross-sectional study of patients with histologically confirmed melanoma and dermoscopic images of the tumor. The patients were divided into 3 groups: melanoma in situ, thin melanoma (≥ 1 mm Breslow thickness), and thick melanoma (≥ 1 mm Breslow thickness). Age, sex, tumor location, and histologic and dermoscopic characteristics were analyzed in all cases.

Results: We studied 215 patients: 88 with melanoma in situ, 73 with thin melanoma, and 54 with thick melanoma. The frequency of the following dermoscopic features increased with increasing Breslow thickness: the blue-white veil ($P < .001$), white shiny structures ($P < .001$), and milky-red areas ($P < .003$). Angulated lines, by contrast, became less common with increasing thickness ($P < .002$).

Conclusions: Dermoscopy not only improves diagnostic accuracy for pigmented lesions but also helps in the preoperative assessment of Breslow thickness in melanoma.

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PALABRAS CLAVE

Dermatoscopia;
Melanoma;
Breslow;
Invasión

La dermatoscopia como herramienta para inferir el Breslow del melanoma**Resumen**

Antecedentes y objetivo: La incidencia del melanoma se ha incrementado significativamente y la forma más efectiva para disminuir su mortalidad es el diagnóstico precoz. La dermatoscopia aumenta la sensibilidad en el diagnóstico del melanoma, y por medio del análisis de las estructuras dermatoscópicas es posible estimar su grosor. Nuestro objetivo fue analizar la influencia del Breslow en las características dermatoscópicas del melanoma.

Materiales y métodos: Estudio observacional de corte transversal. Se incluyeron pacientes con melanoma confirmado histológicamente y una imagen dermatoscópica del mismo. Se dividieron en tres grupos, melanoma in situ, melanoma fino (< 1 mm de Breslow) y melanoma grueso (\geq 1 mm de Breslow), y se analizaron el sexo, la edad, la localización, las características histológicas y las características dermatoscópicas.

Resultados: Se analizaron 215 pacientes, 88 con melanoma in situ, 73 con melanoma fino y 54 con melanoma grueso. Las estructuras dermatoscópicas que incrementaron su frecuencia a medida que aumentó el Breslow del melanoma fueron el velo azul blanquecino ($p < 0,001$), las estructuras blanco brillantes ($p < 0,001$) y las áreas rojo lechosas ($p < 0,003$). Por otro lado, las líneas anguladas disminuyeron su frecuencia a medida que se incrementó el Breslow ($p < 0,002$).

Conclusiones: La evaluación dermatoscópica tiene un importante rol, no solo en la precisión diagnóstica de las lesiones pigmentadas, sino también en ayudarnos a estimar el grosor preoperatorio del melanoma.

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Introduction

The incidence of melanoma has risen dramatically in recent years, particularly in men, in whom it is now the fastest-increasing cancer. In women, the increase in incidence is exceeded only by that of lung cancer.¹ Despite the advances that have been made in the treatment of invasive melanoma, early diagnosis of thin tumors continues to be the most effective way of reducing mortality.²

Dermoscopy is a cost-effective, noninvasive tool that provides better visualization of the colors and structures of skin lesions, allowing for better analysis.³ Several meta-analyses have demonstrated that dermoscopy performed by an experienced examiner improves diagnostic accuracy in pigmented lesions.⁴ It is 10% to 27% more sensitive than naked eye examination.⁵

Studies have shown certain dermoscopic features to be associated with Breslow thickness in melanomas on glabrous skin, indicating that dermoscopy might be a useful tool for estimating tumor thickness.⁵ Using dermoscopic findings to estimate Breslow thickness would improve the multidisciplinary management of melanoma, as it would facilitate surgical planning and decisions regarding imaging or laboratory tests, which could be ordered whilst awaiting definitive results from the pathology laboratory.

Objective

The aim of this study was to analyze the influence of Breslow thickness on the dermoscopic characteristics of melanoma.

Materials and Methods

We performed an observational cross-sectional study of all patients with histologically confirmed melanoma on glabrous skin treated at Hospital Italiano de Buenos Aires in Argentina between January 1, 2011 and December 31, 2016. Only patients for whom dermoscopic images were available were included. These images had been obtained using the DermaGraphix Mirror body mapping system (Canfield). Multiple images were taken of lesions that were too large to fit into a single photographic field to ensure that all areas of the tumor were covered. The images were analyzed by 2 dermatologists with over 7 years' experience in the capture and analysis of dermoscopic images; they worked separately using a set of unified criteria and definitions (Table 1). Any discrepancies regarding the presence or absence of a given feature were resolved by joint analysis of the images in question until agreement was reached. Sex, age, tumor location, histologic characteristics (histologic subtype, Breslow thickness, ulceration, mitotic rate, and regression), and dermoscopic characteristics were analyzed in all cases. The patients were then divided into 3 groups: melanoma in situ, thin melanoma (Breslow thickness < 1 mm), and thick melanoma (Breslow thickness \geq 1 mm).

Histologic characteristics were analyzed only for patients with thin and thick melanomas, as detailed histologic findings are not described for melanomas in situ.

Quantitative variables are expressed as medians and interquartile range (IQR), and categorical variables as absolute and relative frequencies. Between-group differences were analyzed using the Kruskal-Wallis test for quantitative variables and the χ^2 or Fisher test for categorical variables. We also performed multiple comparisons, with Bonferroni correction. Statistical significance was set

Table 1 Criteria for Analyzing Dermoscopic Features and Definitions.

Dermoscopic Feature	Definition	Type of Variable	Value
Asymmetry	In 0, 1, or 2 perpendicular axes; assess contour, colors, and structures	Qualitative dichotomous	YES/NO
Atypical pigment network	Thick brown or black interconnected lines forming a meshed pattern, surrounding irregular unpigmented holes	Qualitative dichotomous	YES/NO
Reverse network	Hypopigmented interconnected lines forming a meshed pattern, surrounding pigmented globules	Qualitative dichotomous	YES/NO
Angulated lines	Pigmented lines forming a rhomboid or zig-zag pattern	Qualitative dichotomous	YES/NO
Pseudopods	Bulbous projections at the edge of a lesion, connected to the body of the tumor or the pigment network. They will never be seen distributed regularly or symmetrically around the lesion. When they are connected directly to the body of the tumor, they must form an acute angle with the tumor edge or emerge from linear or curvilinear extensions. When they are connected to the pigment network, the end portion of the bulb must be wider than any part of the surrounding network and at least twice as wide as the part that is directly connected to the network.	Qualitative dichotomous	YES/NO
Radial streaming	Brown or black linear structures of variable thickness that are not clearly connected to the lines of the network; they are more evident at the edge of the lesion	Qualitative dichotomous	YES/NO
Irregular dots and/or globules	Round or oval black or brown structures of varying sizes irregularly distributed within the lesion	Qualitative dichotomous	YES/NO
Bologna sign	Thick or hyperpigmented area of the pigment network at the edge of the lesion	Qualitative dichotomous	YES/NO
Irregularly distributed hyperpigmented areas	Structureless black, brown, and/or gray areas symmetrically or asymmetrically distributed within the lesion	Qualitative dichotomous	YES/NO
Blue-white veil	Irregular structureless area of confluent blue pigment with an overlying white "ground glass" film. The pigmentation will not be present in all lesion and generally corresponds to a clinically elevated part.	Qualitative dichotomous	YES/NO
Regression	White scar-like depigmentation with or without blue-gray dots on a flat part of the lesion	Qualitative dichotomous	YES/NO
White shiny structures	Thick white structures dispersed in an orthogonal or stellate pattern	Qualitative dichotomous	YES/NO
Milky-red areas	Fuzzy or unfocused milky-red globules and/or larger areas	Qualitative dichotomous	YES/NO
Atypical vessels	Dot-like, glomerular, linear, irregular, serpentine, polymorphous, or corkscrew vessels	Qualitative dichotomous	YES/NO



Figure 1 Thick melanoma with a blue-white veil, defined as an irregular, structureless area of confluent blue pigmentation with an overlying white ‘‘ground glass’’ film.

at less than 5%. Statistical analyses were performed in R software.

Results

We analyzed 215 patients: 88 (40.9%) with melanoma in situ, 73 (34%) with thin melanoma, and 54 (25.1%) with thick melanoma. Their median age was 67 years (IQR, 51–75 years) and 51% were female. The most common location was the trunk (52.1%, $n=112$) followed by the lower limbs (25.6%, $n=55$), the upper limbs (12.1%, $n=26$), and the head and neck (10.2%, $n=22$). The most common histologic subtype in the 127 patients with invasive melanoma was superficial spreading melanoma (74%, $n=94$), followed by nodular melanoma (22%, $n=28$), acral lentiginous melanoma (2.4%, $n=3$), and lentigo maligna melanoma (1.6%, $n=2$).

Ulceration and a high mitotic rate were observed in a higher proportion of patients with thick melanoma ($P < .001$). Superficial spreading melanoma was more common in patients with thin melanoma ($P < .001$), while nodular melanoma were more common in those with thick melanoma ($P < .001$). The absolute and relative frequencies of all the histologic features identified in thin and thick melanomas are summarized in Table 2. As mentioned, melanomas in situ were excluded from this analysis due to the lack of detailed information. The following dermoscopic features were more common in thick melanomas: the blue-white veil ($P < .001$) (Fig. 1), white shiny structures ($P < .001$) (Fig. 2), and milky-red areas ($P < .003$) (Fig. 3). Angulated lines (Fig. 4), by contrast, were more common ($P < .002$). The absolute and relative frequencies of the dermoscopic features identified in melanomas in situ, thin melanomas, and thick melanomas are shown in Table 3.

Discussion

Our study supports previous findings showing that the blue-white veil, white shiny structures, and milky-red areas are more common in thick melanomas than in thin melanomas,^{5–11} while angulated lines in the pigment network are more common in thin melanomas.¹²

Dermoscopy is a cost-effective tool for diagnosing pigmented skin lesions and it is 10% to 27% more sensitive for the diagnosis of melanoma than naked eye examination. The findings of a number of studies that have analyzed the influ-

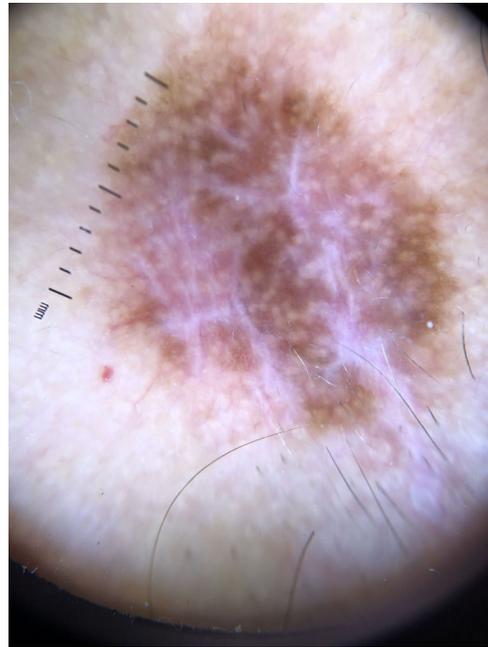


Figure 2 Thick melanoma with white shiny structures, defined as thick, linear, white structures dispersed in an orthogonal or stellate pattern.

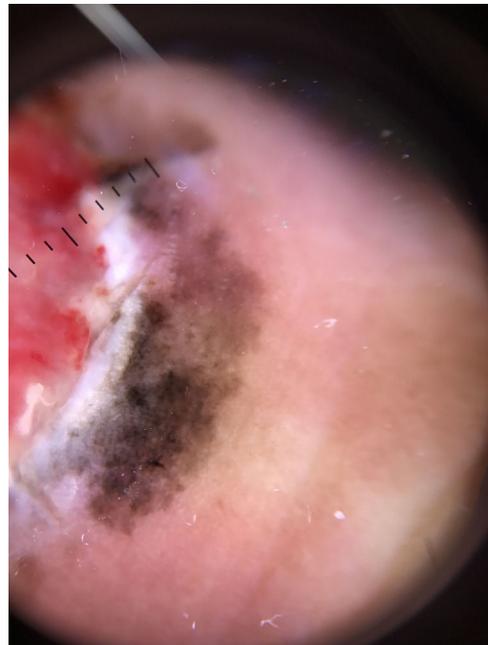


Figure 3 Thick melanoma with milky-red areas, defined as fuzzy or unfocused milky-red globules and/or larger areas.

ence of Breslow thickness on dermoscopic features^{5–8} point to a possible association in melanomas on glabrous skin.

Martins da Silva et al.⁵ found that invasive melanomas analyzed by dermoscopy tend to have 3 or more colors, milky-red areas, a blue-white veil, and an atypical pigment network. Gallegos et al.,⁷ in turn, found that the most common dermoscopic features in this setting were asymmetry in 2 axes and 2 or more colors. González et al.,⁸ on analyzing associations between dermoscopic features and sentinel

Table 2 Absolute and Relative Frequencies of Histologic Features in Thin and Thick Melanomas.

Histologic Feature, No. (%) (n = 127)	Thin melanomas, No. (%) (n = 73)	Thick melanomas, No. (%) (n = 54)	<i>P</i> Value
Ulceration 15 (12)	2 (3)	13 (24)	<.001
High mitotic rate 33 (26)	3 (4)	30 (56)	<.001
Regression 27 (21)	18 (25)	9 (17)	<.001
Histologic subtype, No. (%) (n = 127)	Thin melanomas, No. (%) (n = 73)	Thick melanomas, No. (%) (n = 54)	<i>P</i> Value
Superficial spreading 93 (73)	70 (96)	23 (43)	<.001
Nodular 29 (23)	1 (1.4)	28 (52)	<.001
Acral lentiginous 3 (2.5)	0	3 (5)	.18
Lentigo maligna melanoma 2 (1.6)	2 (2.6)	0	.74

Table 3 Absolute and Relative Frequencies of Dermoscopic Features According to Degree of Invasion (Melanoma in situ, Thin Melanoma, Thick melanoma).

Dermoscopic Feature (n = 215)	Melanomas in situ (%) (n = 88)	Thin Melanomas, No. (%) (n = 73)	Thick Melanomas, No. (%) (n = 54)	P Value
Asymmetry	85 (96.6)	71 (97.3)	53 (98.1)	.861
Atypical pigment network	80 (90.9)	65 (89)	43 (79.6)	.127
Reverse network	15 (17)	14 (19.2)	8 (14.8)	.812
Angulated lines	19 (21.6)	7 (9.6)	1 (1.9)	.002
Pseudopods	12 (13.6)	14 (19.2)	10 (18.5)	.594
Radial streaming	22 (25)	26 (35.6)	17 (31.5)	.335
Irregular dots and/or globules	32 (36.4)	37 (50.7)	23 (42.6)	.188
Bologna sign	7 (8)	9 (12.3)	2 (3.7)	.218
Irregularly distributed hyperpigmented areas	43 (48.9)	33 (45.2)	25 (46.3)	.892
Blue-white veil	20 (22.7)	31 (42.5)	37 (68.5)	<.001
Regression	29 (33)	35 (47.9)	24 (44.4)	.130
White shiny structures	15 (17)	23 (31.5)	31 (57.4)	<.001
Milky-red areas	10 (11.4)	16 (21.9)	19 (35.2)	.003
Atypical vessels	13 (14.8)	20 (27.4)	14 (25.9)	.110



Figure 4 Melanoma in situ with angulated lines, defined as pigmented lines forming a rhomboid or zig-zag pattern.

lymph node (SLN) positivity in melanoma, found that ulceration, blotches (homogeneous hyperpigmented areas), and absence of a pigment network were correlated with a positive SLN biopsy.

Other authors have linked blue-gray areas, radial streaming, and an atypical vascular pattern to a Breslow thickness of greater than 0.75 mm.^{9–11}

Breslow thickness is the histologic feature with the strongest prognostic value in melanoma; it determines excision margins, the selection of patients for SLN biopsy, and the need for preoperative imaging studies for staging.¹ Histology is necessary for establishing a diagnosis of melanoma, and Breslow thickness and ulceration help determine an appropriate course of action. Using dermoscopic features to estimate Breslow thickness would improve multidisciplinary management by facilitating surgical planning and enabling the performance of appropriate imaging or laboratory tests while the biopsy specimen is being processed.

This study has some limitations. We analyzed the different histologic subtypes as a whole, for example, and did not distinguish between tumor locations (except for tumors of the scalp, which were not included). This may have influenced our results, as dermoscopic findings may vary according to anatomic site.

Conclusions

We have shown significant associations between certain dermoscopic features and Breslow thickness. Dermoscopy is important not only for diagnosing pigmented lesions, but also for estimating preoperative Breslow thickness in melanoma. Although our results are significant, their reproducibility and validity need to be confirmed in prospective studies including evaluations by different groups of observers.

Conflicts of interest

The authors declare that they have no conflicts of interest.

References

1. National Comprehensive Cancer Network, Cutaneous Melanoma. Available at: <https://www.nccn.org>.
2. Hu S, Parmet Y, Allen G, Parker DF, Ma F, Rouhani P, et al. Disparity in melanoma: a trend analysis of melanoma incidence and stage at diagnosis among whites, Hispanics, and blacks in Florida. *Arch Dermatol*. 2009;145:1369–74, <http://dx.doi.org/10.1001/archdermatol.2009.302>.
3. Thomas L, Puig S. Dermoscopy, digital dermoscopy and other diagnostic tools in the early detection of melanoma and follow-up of high-risk skin cancer patients. *Acta Derm Venereol*. 2017;14–21, <http://dx.doi.org/10.2340/00015555-2719>.
4. Vestergaard ME, Macaskill P, Holt PE, Menzies SW. Dermoscopy compared with naked eye examination for the diagnosis of primary melanoma: a meta-analysis of studies performed in a clinical setting. *Br J Dermatol*. 2008;159:669–76, <http://dx.doi.org/10.1111/j.1365-2133.2008.08713.x>.
5. Martins da Silva VP, Ikino JK, Sens MM, Nunes DH, Di Giunta G. Dermoscopic features of thin melanomas: a comparative study of melanoma in situ and invasive melanomas smaller than or equal to 1mm. *An Bras Dermatol*. 2013;88:712–7, <http://dx.doi.org/10.1590/abd1806-4841.20132017>.
6. Mun J-H, Jo G, Darmawan CC, Park J, Bae JM, Jin H, et al. Association between Breslow thickness and dermoscopic findings in acral melanoma. *J Am Acad Dermatol*. 2018;79:831–5, <http://dx.doi.org/10.1016/j.jaad.2018.06.004>.
7. Gallegos-Hernández JF, Ortiz-Maldonado AL, Minauro-Muñoz GG, Arias-Ceballos H, Hernández-Sanjuan M. Dermatoscopia en melanoma cutáneo. *Cir Cir*. 2015;83:107–11, <http://dx.doi.org/10.1016/j.circir.2015.04.004>.
8. González-Álvarez T, Carrera C, Bennassar A, Vilalta A, Rull R, Alos L, et al. Dermoscopy structures as predictors of sentinel lymph node positivity in cutaneous melanoma. *Br J Dermatol*. 2015;172:1269–77, <http://dx.doi.org/10.1111/bjd.13552>.
9. De Giorgi V, Carli P. Dermoscopy and preoperative evaluation of melanoma thickness. *Clin Dermatol*. 2002;20:305–8, [http://dx.doi.org/10.1016/s0738-081x\(02\)00224-9](http://dx.doi.org/10.1016/s0738-081x(02)00224-9).
10. Stante M, De Giorgi V, Cappugi P, Giannotti B, Carli P. Non-invasive analysis of melanoma thickness by means of dermoscopy: a retrospective study. *Melanoma Res*. 2001;11:147–52, <http://dx.doi.org/10.1097/00008390-200104000-00009>.
11. Argenziano G, Fabbrocini G, Carli P, De Giorgi V, Delfino M. Clinical and dermoscopic criteria for the preoperative evaluation of cutaneous melanoma thickness. *J Am Acad Dermatol*. 1999;40:61–8, [http://dx.doi.org/10.1016/s0190-9622\(99\)70528-1](http://dx.doi.org/10.1016/s0190-9622(99)70528-1).
12. Daelen AV, Vanden Daelen AV, Ferreira I, Marot L, Tromme I. A digital dermoscopy follow-up illustration and a histopathologic correlation for angulated lines in extrafacial lentigo maligna. *JAMA Dermatol*. 2016;152:200–3, <http://dx.doi.org/10.1001/jamadermatol.2015.4132>.