



ACTAS Dermo-Sifiliográficas

Full English text available at
www.actasdermo.org



CONSENSUS DOCUMENT

Protocol for the Histologic Diagnosis of Cutaneous Melanoma: Consensus Statement of the Spanish Society of Pathology and the Spanish Academy of Dermatology and Venereology (AEDV) for the National Cutaneous Melanoma Registry[☆]



A. Tejera-Vaquerizo,^{a,*} M.T. Fernández-Figueras,^b Á. Santos-Briz,^c J.J. Ríos-Martín,^d C. Monteagudo,^e Á. Fernández-Flores,^f C. Requena,^g V. Traves,^h M.A. Descalzo-Gallego,ⁱ J.L. Rodríguez-Peralto^j

^a Servicio de Dermatología, Instituto Dermatológico GlobalDerm, Palma del Río, Córdoba, España; Unidad de Oncología Cutánea, Hospital San Juan de Dios, Córdoba, Spain

^b Servicio de Anatomía Patológica, Hospital Universitari General de Catalunya, Grupo Quironsalud, Sant Cugat del Vallès, Barcelona, Spain

^c Servicio de Anatomía Patológica, Complejo Asistencial Universitario de Salamanca, Salamanca, Spain

^d Servicio de Anatomía Patológica, Hospital Universitario Virgen Macarena, Sevilla, Spain

^e Servicio de Anatomía Patológica, Hospital Clínico Universitario de Valencia, Universidad de Valencia, Valencia, Spain

^f Servicio de Anatomía Patológica, Hospital del Bierzo, Ponferrada, León, España; Servicio de Anatomía Patológica, Hospital de la Reina, Ponferrada, León, Spain

^g Servicio de Dermatología, Instituto Valenciano de Oncología, Valencia, Spain

^h Servicio de Anatomía Patológica, Instituto Valenciano de Oncología, Valencia, Spain

ⁱ Unidad de Investigación, Fundación Academia Española de Dermatología y Venereología, Madrid, Spain

^j Servicio de Anatomía Patológica, Hospital Universitario 12 de Octubre, Madrid, Spain

Received 23 July 2020; accepted 19 September 2020

KEYWORDS

Melanoma;
Consensus;
Histology;
Registries;

Abstract This article describes a proposed protocol for the histologic diagnosis of cutaneous melanoma developed for the National Cutaneous Melanoma Registry managed by the Spanish Academy of Dermatology and Venereology (AEDV). Following a review of the literature, 36 variables relating to primary tumors, sentinel lymph nodes, and lymph node dissection were evaluated using the modified Delphi method by a panel of 8 specialists (including 7 pathologists).

[☆] Please cite this article as: Tejera-Vaquerizo A, Fernández-Figueras MT, Santos-Briz Á, Ríos-Martín JJ, Monteagudo C, Fernández-Flores Á et al. Protocolo de diagnóstico histológico para muestras de pacientes con melanoma cutáneo. Documento de consenso de la SEAP y la AEDV para el Registro Nacional de Melanoma. Serie de casos. Actas Dermosifiliogr. 2021;112:32–43.

* Corresponding author.

E-mail address: antoniotejera@aedv.es (A. Tejera-Vaquerizo).

Delphi technique;
Prognosis

PALABRAS CLAVE

Melanoma;
Consenso;
Histología;
Registros;
Método Delphi;
Prognóstico

Consensus was reached on the 30 variables that should be included in all pathology reports for cutaneous melanoma and submitted to the Melanoma Registry. This list can also serve as a model to guide routine reporting in pathology departments.

© 2020 AEDV. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Protocolo de diagnóstico histológico para muestras de pacientes con melanoma cutáneo. Documento de consenso de la SEAP y la AEDV para el Registro Nacional de Melanoma

Resumen El presente texto es una propuesta de protocolo de diagnóstico histológico para el melanoma cutáneo, realizado a instancias del Registro Nacional de Melanoma de la Academia Española de Dermatología y Venereología. Tras una búsqueda bibliográfica, un grupo de 8 panelistas (7 patólogos) decidieron entre 36 variables del tumor primario, el ganglio centinela y la linfadenectomía incluir un total de 30 variables mediante el método de Delphi modificado. Se han consensado las variables que deberían contener un informe histológico de melanoma cutáneo para que puedan ser utilizadas en el Registro de Melanoma o servir de modelo para los distintos Servicios de Anatomía Patológica a la hora de elaborar sus propios informes de forma rutinaria.

© 2020 AEDV. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

The annual incidence of cutaneous melanoma in Spain is 8.7 cases per 100,000 population.¹ This corresponds to approximately 4000 new cases every year. Access to incidence and mortality data and a system containing standardized information would facilitate melanoma research in Spain and provide valuable epidemiologic data. This information is available in provinces with cancer registries. The Spanish National Cutaneous Melanoma Registry was created in 1997 under the auspices of the Spanish Academy of Dermatology and Venereology (AEDV) and has gone through 2 organizational phases over the years.²

One of the main drawbacks of the registry is the lack of a standardized protocol to guide researchers wishing to enter data. Facilitating this task by standardizing the information to be collected, combined with the use of flexible data capture tools,³ could increase participation in the registry. In addition, a uniform reporting protocol could serve as a model to guide routine reporting of cutaneous melanoma by pathology laboratories.

Consensus is lacking on which histologic variables should be included in pathology reports for cutaneous melanoma, although certain associations, such as the College of American Pathologists, issue recommendations in the form of checklists that are updated annually.⁴ Previous work done in Spain includes a proposed protocol for reporting histologic data for primary cutaneous melanomas published by a group of researchers from the Community of Valencia.⁵

The aim of this study was to achieve consensus on which histologic variables should be recorded in pathology reports for cutaneous melanoma in Spain to facilitate their subsequent inclusion in the National Cutaneous Melanoma Registry.

Material and Methods

This consensus statement is an initiative of the Spanish National Cutaneous Melanoma Registry's coordinator (ATV) and the process was managed by the AEDV's Healthy Skin Foundation and the Spanish Pathology Society (SEAP). The different stages in the process are summarized in Appendix B Supplementary Fig. 1.

The Spanish Dermopathology Research Group, which is part of the AEDV, was asked to propose participants with a special interest in cutaneous melanoma. Several melanoma experts were also consulted and asked to propose additional candidates. The expert panel formed to create this consensus statement consisted of 8 experts (7 pathologists and 1 dermatologist), all with extensive experience and numerous publications on the subject.

The first phase of the process consisted of searching similar documents (mainly the most widely used guidelines in this field^{5–9}) to identify commonly used histologic variables. Additional variables were identified by a literature search for potential prognostic factors in PubMed using the terms («Melanoma/pathology»[MAJR]) AND «Prognosis»[MeSH]) AND «Skin Neoplasms/pathology»[MAJR]).

The candidate variables were classified into 3 groups: primary tumor variables, sentinel lymph node (SLN) variables, and lymph node dissection (LND) variables.

These variables were then evaluated by the expert panel using a modified 2-round Delphi approach¹⁰ designed to achieve consensus on which variables to include in the protocol for the histologic diagnosis of cutaneous melanoma. The panelists were asked to score each variable according to 1) relevance—potential impact on decision-making or potential prognostic value—and 2) feasibility—ease with which the variable can be measured or evaluated by his-

tology. Each variable was scored on a scale of 1 (not at all relevant/not at all feasible) to 9 (very relevant/very feasible). In the first round of the Delphi process, the panelists were able to suggest other potentially relevant or feasible variables considered not to be adequately covered by the other options. In the second round, they were given the opportunity to revise their scores from the first round. To do this, they were able to consult these scores together with the group scores presented as means, medians, modes, maximums, and minimums. At the end of the 2 rounds, the variables were classified as having achieved sufficient consensus for inclusion (median and mode scores > 7), sufficient consensus for exclusion (median and mode scores < 3), or insufficient consensus for either inclusion or exclusion (median and mode scores > 3 and < 6). Variables with median and mode scores outside the above ranges were considered not to have achieved consensus.

Variables with insufficient consensus for either inclusion or exclusion were discussed by the experts at an online meeting to decide whether they should be included in the final document or not. Variables without consensus or with sufficient consensus for exclusion were discarded. Finally, the panel discussed a number of additional points following an external review.

Results

Thirty-six variables (21 primary tumor, 10 SLN, and 5 LND variables) were selected for evaluation following the review of the literature and the main protocols. Consensus for inclusion was reached for 19 variables (8 primary tumor, 6 SLN, and 5 LND variables) in the first round of the modified Delphi process (Appendix B Supplementary Table 1). An additional variable (angiotropism, relating to the primary tumor) proposed by the experts was added during this round. Consensus was reached for an additional 5 variables (all primary tumor variables) in the second round, bringing the total to 25 (Appendix B, Supplementary Table 2). In the final online meeting, consensus was reached for the inclusion of 30 variables—18 primary tumor, 7 SLN, and 5 LND variables (Appendix B supplementary table 3)—in the final version of the proposed protocol for the histologic diagnosis of cutaneous melanoma (Table 1).

Discussion

The aim of this study was to propose a set of primary tumor, SLN, and LND histologic variables that should systematically be included in pathology reports for cutaneous melanoma.

Although the main aim of creating a protocol for the histologic evaluation of cutaneous melanoma was to standardize the reporting of melanoma data in the Spanish National Cutaneous Melanoma Registry, there is no doubt that the content of the protocol can serve as a model for melanoma reports issued by pathology or dermatopathology laboratories.

It is important to note that apart from the variables required for correct melanoma staging by the American Joint Committee on Cancer (AJCC),¹¹ the protocol contains other variables that the panel considered to be potentially relevant in terms of their prognostic value or usefulness in

decision-making. Most of these variables are also used in existing protocols.^{7,8} Not all the variables selected for evaluation were included in the final protocol as the experts did not reach consensus on their relevance or feasibility. One example is ulceration width (part of the College of American Pathologists' protocol⁷), which was considered to be particularly prone to subjective interpretation. The presence of nevus cells in SLNs (featured in the European Organisation for Research and Treatment of Cancer [EORTC] protocol for the evaluation of SLNs in melanoma⁸) was also excluded, as it was considered that discrimination of these cells is relatively straightforward.

Other variables were excluded because it was considered that they did not add any new information not already covered (e.g., the SLN S-classification¹²) or because they involved particular difficulties and were not supported by sufficient evidence (e.g., SLN metastasis mitotic rate).¹³

Although tumor-infiltrating lymphocyte (TIL) density is also prone to subjective interpretation, it achieved sufficient consensus for inclusion, mainly because of its potential relevance in the new era of immunotherapy for melanoma.¹⁴

The variables included in the final protocol for the histologic diagnosis of melanoma are defined and briefly discussed below.

Primary Tumor Variables

Location

Pathologists should be informed of the location of the melanoma and in turn specify this in the pathology report, as patients may have more than 1 tumor. In addition, prognosis varies according to location. Melanomas on the upper and lower limbs (not counting the hands and feet) are associated with a better prognosis.^{15,16}

Type of Biopsy

An excisional biopsy is generally recommended.¹⁷ Although biopsy type (excisional, incisional, shave, or punch) has not been associated with differences in survival, it has been linked to differences in Breslow thickness and percentage of positive margins (especially in the case of shave biopsies).¹⁸

Macroscopic Tumor Diameter

Although macroscopic tumor diameter is considered a prognostic factor in squamous cell carcinoma, this is not currently the case for melanoma. Nonetheless, several studies have reported a link between larger diameter and worse prognosis.^{19,20} Tumor diameter has also been used to calculate tumor volume as a prognostic factor in melanoma,²¹ and some authors have started to use it to calculate tumor growth rate.²²

Melanoma In Situ Versus Invasive Melanoma

Melanoma in situ is confined to the epidermis. As such, there is no basement membrane involvement. Although melanoma in situ is considered to be virtually curable, mortality has been described in some series.²³

Maximum Tumor (Breslow) Thickness

Maximum tumor thickness must be measured using a calibrated ocular micrometer. It is measured from the upper

Table 1 Proposed Pathology Report for Cutaneous Melanoma.

Cutaneous Melanoma Protocol	
I. Primary tumor biopsy	
1. Location	—
2. Type of biopsy	Excisional Incisional Shave Punch Other:
3. Macroscopic tumor diameter (mm)	—
4. Melanoma	In situ Invasive
5. Primary tumor (Breslow) thickness (mm)	— ____ mm At least: ____ mm
6. Ulceration	No Yes
7. Clark level	I II III IV M At least Cannot be determined
8. Mitotic rate	— ____ mitoses/mm ²
9. Histologic subtype	—Superficial spreading melanoma —Melanoma maligna lentigo —Desmoplastic melanoma —Spitzoid melanoma —Acral lentiginous melanoma —Mucosal melanoma —Melanoma arising in congenital nevus —Melanoma arising in blue nevus —Uveal melanoma —Nodular melanoma
Tumor-infiltrating lymphocytes	Density Brisk Nonbrisk Absent Location Intratumoral Peritumoral Both Neither
Neurotropism	—Absent —Present
Lymphovascular invasion	—Absent —Present
Angiotropism	—Absent —Present
Microsatellitosis	—Absent —Present
Regression	—Absent —Present — < 75% — > 75%

Table 1 (Continued)

Cutaneous Melanoma Protocol	
Association with nevus:	<ul style="list-style-type: none"> —No —Congenital —Blue —Acquired —Dysplastic —Other:.....
Margin involvement	<ul style="list-style-type: none"> —No —Lateral margin: distance_____ mm At least_____ mm Not assessable _____ —Deep margin: distance_____ mm At least_____ mm Not assessable:.....
Pathologic stage	<ul style="list-style-type: none"> ___ pTX: Primary tumor thickness cannot be assessed (e.g., diagnosis by curettage) ___ pT0: No evidence of primary tumor (e.g., completely regressed melanoma) ___ pTis: Melanoma in situ ___ pT1: Melanoma 1.0 mm or less in thickness, ulceration status unknown or unspecified ___ pT1: Melanoma < 0.8 mm in thickness, no ulceration ___ pT1b: Melanoma < 0.8 mm in thickness with ulceration, or melanoma 0.8–1.0 mm in thickness with or without ulceration ___ pT2: Melanoma > 1.0–2.0 mm in thickness, ulceration status unknown or unspecified ___ pT2a: Melanoma > 1.0–2.0 mm in thickness, no ulceration ___ pT2b: Melanoma > 1.0–2.0 mm in thickness, with ulceration ___ pT3: Melanoma > 2.0–4.0 mm in thickness, ulceration status unknown or unspecified ___ pT3a: Melanoma > 2.0–4.0 mm in thickness, no ulceration ___ pT3b: Melanoma > 2.0–4.0 mm in thickness, with ulceration ___ pT4: Melanoma > 4.0 mm in thickness, ulceration unknown or not specified ___ pT4a: Melanoma > 4.0 mm in thickness, no ulceration ___ pT4b: Melanoma > 4.0 mm in thickness, with ulceration
Sentinel lymph node biopsy	
Number of lymph nodes sent or found	— nodes
Number of positive lymph nodes	— nodes
Size of largest metastatic deposit	— mm
Location of metastasis in sentinel lymph node	<ul style="list-style-type: none"> —Subcapsular —Parenchymal —Combined —Multifocal —Extensive —Cannot be determined
Extranodal extension	<ul style="list-style-type: none"> —Present —Absent —Cannot be determined

(Continued)

Cutaneous Melanoma Protocol	
Number of metastatic deposits	<ul style="list-style-type: none"> –1 –2–5 –6–10 –11–20 –>20 –Cannot be determined
Matted nodes	<ul style="list-style-type: none"> –Absent –Present
Lymphadenectomy	
Number of lymph nodes submitted or found	–nodes
Number of lymph nodes with metastatic deposits:	–nodes
Lymph node ratio (number of positive nodes/total nodes examined):	–
Matted nodes	<ul style="list-style-type: none"> –Absent –Present
5. Pathologic stage	<ul style="list-style-type: none"> –pN0: No regional lymph node metastasis detected –pN1: One tumor-involved node or microsattellites and/or satellites or in-transit metastases with no tumor-involved nodes <ul style="list-style-type: none"> –pN1a: One tumor-involved node (e.g., detected by sentinel lymph node biopsy) with no microsattellites and/or satellites or in-transit metastases –pN1b: One clinically detected tumor-involved node without satellitosis or in-transit metastases –pN1c: Presence of microsattellites and/or satellites or in-transit metastases with no tumor-involved nodes –pN2: Metastasis in 2 or 3 lymph nodes or microsattellites and/or satellites or in-transit metastases with just 1 tumor-involved node <ul style="list-style-type: none"> –pN2a: Two or 3 clinically occult tumor-involved nodes (e.g., detected by sentinel lymph node biopsy) with no microsattellites and/or satellites or in-transit metastases –pN2b: Two or 3 tumor-involved nodes at least 1 of which was clinically detected, with no microsattellites and/or satellites or in-transit metastases –pN2c: One clinically occult or clinically apparent tumor-involved node with microsattellites and/or satellites or in-transit metastases –pN3: Metastasis in 4 or more regional lymph nodes or microsattellites and/or satellites or in-transit metastases with 2 or more involved regional lymph nodes or any number of matted nodes <ul style="list-style-type: none"> –pN3a: Four or more clinically occult tumor-involved nodes (e.g., detected by sentinel node biopsy) with no microsattellites and/or satellites or in-transit metastases –pN3b: Metastasis in 4 or more lymph nodes, at least 1 of which was clinically detected, with no microsattellites and/or satellites or in-transit metastases –pN3c: Metastasis in 2 or more clinically occult or clinically detected nodes with microsattellite, satellite and/or in-transit metastases or any number of matted nodes with microsattellites and/or satellites or in-transit metastases

edge of the granular layer (or stratum spinosum in the absence of this layer) to the deepest point of the tumor. If the tumor is ulcerated, the starting point for the measurement is the base of the tumor.

Breslow thickness is the most powerful prognostic factor in melanoma and is included in all AJCC staging systems.¹¹ The measurement must be rounded to the nearest tenth of a millimeter (e.g., 0.1 mm) and not hundredth of a millimeter (e.g., 0.01 mm), as recommended in previous classifications. Accordingly, a Breslow thickness of between 0.75 and 0.84 mm must be rounded to 0.8 mm and reported as T1b, while one of between 1.01 and 1.04 mm must be reported as 1.0 mm.²⁴

It may be difficult to measure Breslow thickness in tumors arising in a previous nevus or in variants such as nevoid melanoma with maturing nevus cells in the dermis. Periadnexal extensions can also make measurement more difficult and should not be counted in Breslow thickness measurements.²⁵

If the deepest point is invaded by the tumor, Breslow thickness should be reported as "... at least ____ mm".

Breslow thickness in polypoid melanomas should be measured using the same points as above (top of the granular layer to the deepest point of the tumor). Clark level is not a valid measure in polypoid tumors and should not be reported.

Breslow thickness is not a sum of measurements. In other words, the thickness of a melanoma observed on re-excision for positive margins cannot be added to the thickness calculated in the initial biopsy specimen.

It may also be difficult to measure Breslow thickness in melanomas located around hair follicles. There are 3 possible situations:

- A A melanoma extending down from a hair follicle and then invading the perifollicular dermis
- B A folliculotropic melanoma invading the follicle from the dermis
- C A primary follicular melanoma extending to the dermis

In the second case (folliculotropic melanoma invading the follicle from the dermis), Breslow thickness would be measured as usual, that is, from the upper edge of the granular layer to the deepest point of the melanoma. In the other 2 cases, however, it would be more correct to measure from the innermost layer of the outer root sheath, perpendicular to the main axis of the follicle, to the furthest point of the melanoma.⁹

To avoid these problems, 3 thickness measurements are usually made for perifollicular melanoma:

- A Breslow thickness: measured from the granular layer of the epidermis to the deepest point of the nonperifollicular melanoma.
- B Follicular Breslow thickness: measured from the granular layer of the epidermis to the deepest point of the perifollicular melanoma.
- C Follicular thickness: measured from the innermost layer of the outer root sheath, perpendicular to the main axis of the follicle, to the furthest point of the melanoma.

Breslow thickness may also be difficult to measure in acral skin when there is extensive epidermal hyperplasia. In such cases, the pathologist should add a note specifying that much of the thickness reported is due to this hyperplasia. Where possible, the thickness of the epidermal hyperplasia should also be measured and specified.

An additional challenge in the case of verrucous (papillated) melanoma is that Breslow thickness varies enormously from the base to the apex of the papillae. The recommended strategy in such cases is to measure the thickness from a point halfway between the base and apex to the deepest point of the melanoma.²⁶

Ulceration

Ulceration, that is the complete disappearance of the overlying epithelium, is associated with a worse prognosis in melanoma. It is included in the AJCC staging system.¹¹ As ulceration is an adverse prognostic factor, its presence will result in an upstaging from "a" to "b" for any thickness (T). True ulceration is characterized by the presence of a tissue reaction to loss of epidermis with fibrin and acute inflammation.²⁷ Although ulceration width may have prognostic implications,²⁸ it did not achieve sufficient consensus for inclusion in this protocol.

Clark Level (Depth)

Clark level reflects the depth of a melanoma from the epidermis to the subcutaneous tissue. It is measured on a scale of I to V. It was used as a staging criterion for thin melanomas in earlier AJCC classifications.^{29,30} The panelists considered that it may influence decision-making in certain cases, especially thin melanomas.

Dermal Mitotic Rate

Although dermal mitotic rate is not part of the AJCC staging system, it has been demonstrated to have prognostic value.^{31,32} The correct way to measure it is to identify the dermal hot spot (area of the dermis with the greatest mitotic activity) and then count the number of figures in an area corresponding to 1 mm². The result should be reported as a full number. If no mitotic figures are observed, the pathologist should report the mitotic rate as 0 mitoses/mm² and not as < 1 mitosis/mm² or "not identified".

According to the National Comprehensive Cancer Network (NCCN), a high dermal mitotic rate is an indication for SLN biopsy in patients with thin melanoma.¹⁷ A mitotic rate of more than 2 mitoses/mm² has been linked to a high risk of SLN positivity in thin melanoma.³³

Histologic Subtype

Histologic subtype is based on the 2018 World Health Organization classification of melanomas, which takes into account UV radiation exposure, cellular origin, and genetic characteristics or evolutionary pathways.³⁴

Tumor-Infiltrating Lymphocytes

TILs are regarded as a host response to the tumor. Their density has been linked to prognosis.³⁵

TILs should be classified as *absent* (not identified or identified but not in contact with the tumor), *brisk* (infiltration of the entire base of the tumor [Fig. 1]) or diffuse infiltra-

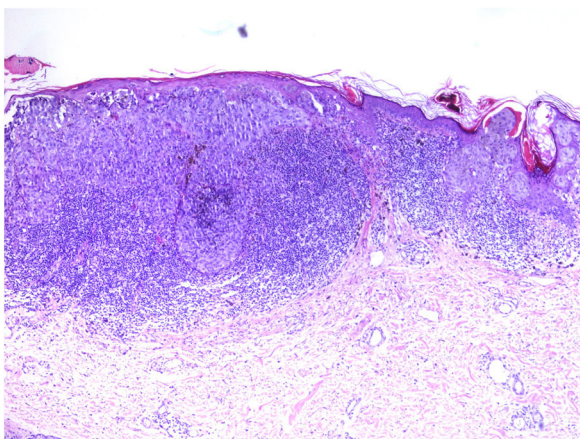


Figure 1 Intense lymphocytic infiltrate across the entire base of a cutaneous melanoma. Hematoxylin-eosin, original magnification $\times 40$.

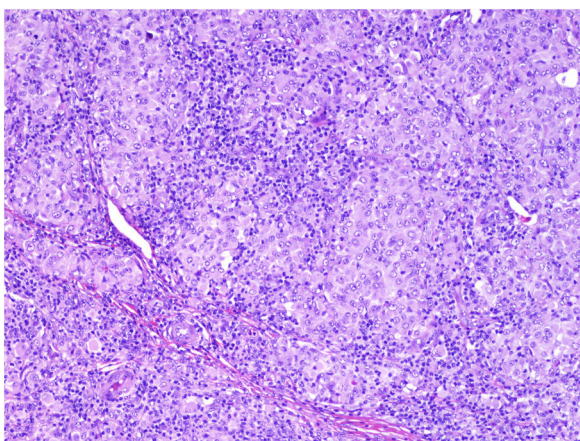


Figure 2 Detail of intense intratumoral lymphocytic infiltrate. Hematoxylin-eosin, original magnification $\times 100$.

tion of the tumor) (Fig. 2), or *nonbrisk* (focal infiltration or infiltration of part of the base of the tumor).

The pathology report should also specify whether the infiltrate is intratumoral, peritumoral, or both.

Neurotropism

Neurotropism is defined as the presence of melanoma cells either adjacent to nerve sheaths, usually circumferentially (perineural invasion) (Fig. 3), or within a nerve (intra-neural invasion) (Fig. 4). It is more commonly seen at the periphery of the tumor. Nerve entrapment due to an expanding tumor should not be regarded as neurotropism. Neurotropism is often observed in desmoplastic melanomas. It can sometimes extend beyond the primary tumor and is therefore associated with a higher risk of local recurrence.³⁶ Neural differentiation in melanomas, generally desmoplastic, is also considered to be a form of neurotropism (Fig. 5).¹⁰

Lymphovascular Invasion

Lymphovascular invasion is defined as the unequivocal presence of endothelium-attached tumor cells within the lumina of lymphatic or blood vessels. Immunohistochemical staining with D2-40 and CD31 together with melanocytic cell mark-

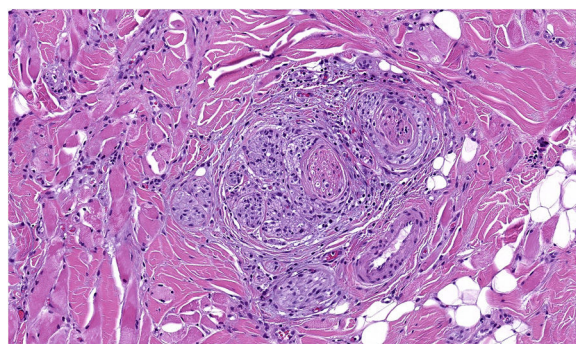


Figure 3 Perineural invasion by melanoma cells (neurotropism). Hematoxylin-eosin, original magnification $\times 200$.

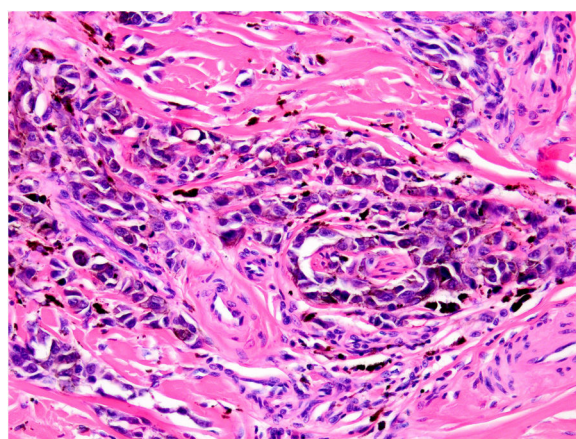


Figure 4 Intraneural invasion by melanoma cells (neurotropism). Hematoxylin-eosin, original magnification $\times 200$.

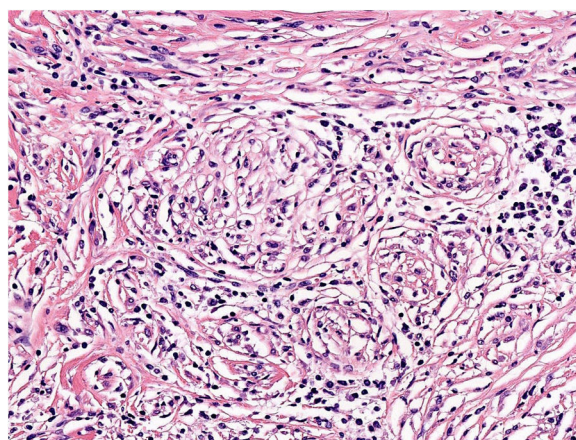


Figure 5 Neural transformation of a melanoma, also known as neurotropism. Hematoxylin-eosin, original magnification $\times 200$.

ers is sometimes used to aid visualization. Lymphovascular invasion is associated with a worse prognosis.^{37,38}

Angiotropism

Angiotropism is defined as the presence of melanoma cells in perivascular spaces, similar to perineural invasion; the cells are considered to act in a pericytic manner (pericytic mimicry) without intravasation (Fig. 6).

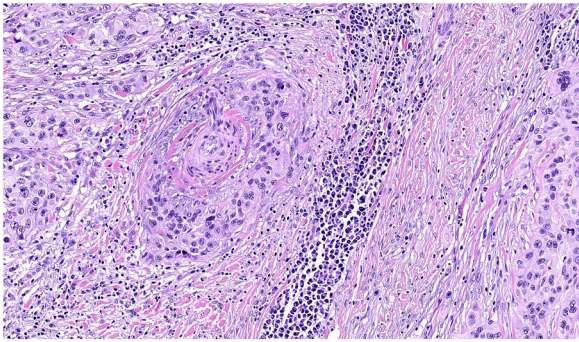


Figure 6 Melanoma cells in perivascular space (angiotropism). Hematoxylin-eosin, original magnification $\times 400$.

Angiotropism has been associated with an increased risk of metastasis.³⁹

Microsatellitosis

Microsatellitosis is defined as the presence of micrometastases adjacent or deep to the primary tumor. It is visualized as a discontinuous nest of metastatic cells separated from the primary tumor by normal skin (Fig. 7A-C). The minimum size and distance requirements specified in the 7th edition of the AJCC Cancer Staging Manual no longer apply. It is advisable to check other tissue sections to ensure that the cells truly correspond to microsatellitosis and are not a continuation of the tumor or an extension of eccrine sweat glands.²⁴

Tumor Regression

Regression in melanoma is regarded as a host response to the presence of the tumor. Characteristic features include replacement of tumor cells by lymphocytic inflammation, attenuation of the epidermis, and nonlaminated dermal fibrosis with inflammatory cells, melanophagocytosis, and increased microvascular density (Fig. 8). The panelists recommend calculating the percentage of regression on the horizontal surface of the tumor and specifying this as $> 75\%$ or $< 75\%$ in the pathology report.^{40,41}

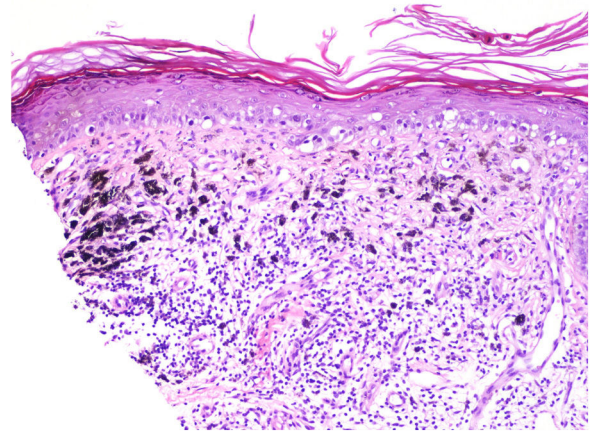


Figure 8 Replacement of tumor cells by lymphocytic inflammation, attenuation of epidermis, melanophagocytosis, and numerous telangiectatic vessels in the regression area of a melanoma. Hematoxylin-eosin, original magnification $\times 100$.

Excision Margins and Distance to Lateral and Deep Margins

It is essential to know margin status and distance from the melanoma to the deep and lateral margins, as based on current recommendations, the surgical margins used (0.5–2 cm) should be proportional to the thickness of the tumor.¹⁷ Clinical and histologic margins are not closely correlated in melanoma. It has also been demonstrated that tumors with a larger diameter will need wider margins to achieve clearance.^{17,42,43}

Regional Lymph Node Metastasis Variables

Number of SLNs Analyzed

Number of SLNs removed has been associated with prognosis and false-negative rate.⁴⁴

Number of Positive SLNs

Number of positive SLNs is a prognostic factor in the 8th version of the AJCC staging manual.¹¹

Size of Largest Metastatic Deposit in SLN

While not currently considered a staging criterion by the AJCC, size of the largest metastatic deposit in SLNs should

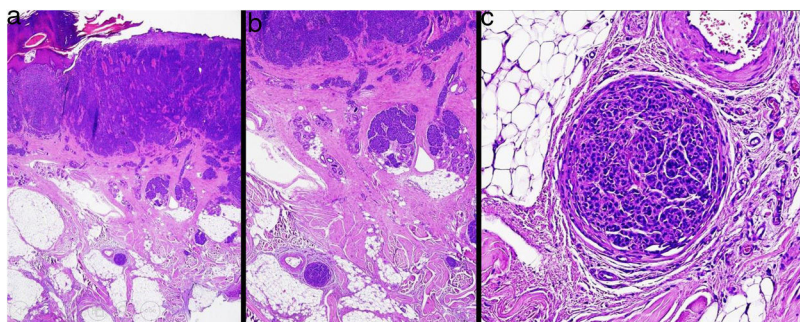


Figure 7 Presence of microscopic metastases (microsatellitosis) deep to a primary melanoma. Hematoxylin-eosin, original magnification $\times 40$ (A), $\times 100$ (B), $\times 200$ (C).

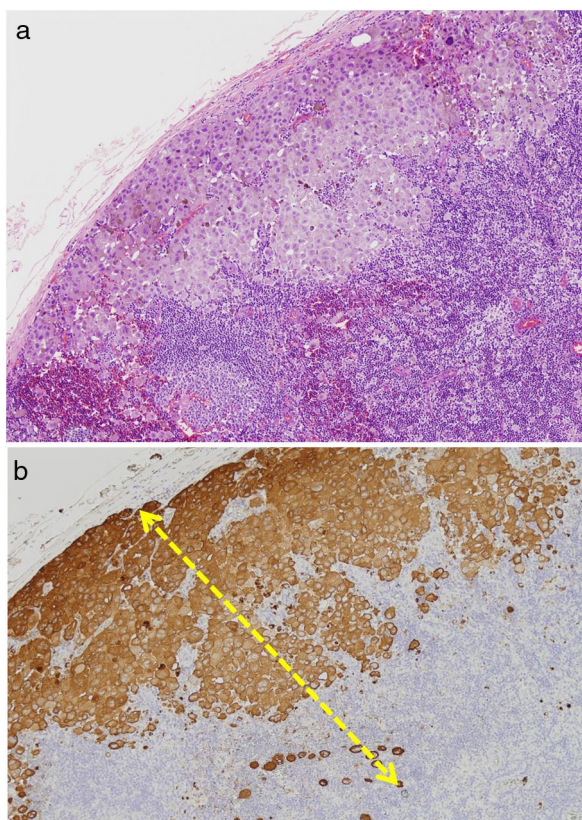


Figure 9 A, Combined metastasis (subcapsular and parenchymal). B, Melanoma cocktail (Melan-A, HMB-45, and tyrosinase) and measurement of maximum diameter.

be specified in pathology reports as it has been correlated with survival (Fig. 9).^{24,45}

Location of SLN Metastases

The Dewar criteria should be used to report the location of micrometastases (subcapsular, parenchymal, combined, multifocal, or diffuse).^{46,47}

Extranodal Extension

Although extranodal extension is rare in melanoma, its presence in SLNs is predictive of a worse prognosis.⁴⁸

Number of Metastatic Deposits

Although the number of metastatic deposits identified in the SLN may vary according to the tissue section analyzed, as recommended by the EORTC this figure should be reported as 1, 2–5, 6–10, 11–20, or > 20.⁸

Lymph Node Ratio After LND

Lymph node ratio, defined as the number of positive lymph nodes out of the total number of lymph nodes excised after LND, has been proposed as a prognostic factor in melanoma.⁴⁹

Conclusions

The strength of this study lies in its design, as the Delphi technique is a standardized, validated method for preparing guidelines and quality indicators⁵⁰ in medicine.

It should be noted, however, that the outcomes of our study may have been biased to some degree by the choice of participants or the lack of evidence on some of the variables excluded from the final protocol. There was significant disagreement among the experts in some cases, meaning that certain variables were not included because of insufficient consensus. They could be reassessed in future updates. One example is the recommendation to report Breslow thickness to a precision of 2 decimal places. Although the AJCC recommends reporting thickness to a single decimal place, there are cases when 2 decimal points are recommended if this is practical or feasible.⁵¹ Other discrepancies were related to the presence or absence of solar elastosis as a sign of more or less sun exposure, which forms the basis of the latest World Health Organization classification of cutaneous tumors,⁵² and the quantification of metastatic deposits in SLNs due to the inherent difficulties and lack of evidence.

In conclusion, we believe that an improved prognostic classification of cutaneous melanoma will ultimately lead to better patient management. It is therefore essential to standardize the reporting of melanoma data to facilitate analysis and subsequent interpretation. We believe that this proposed protocol for the histologic diagnosis of cutaneous melanoma will facilitate and increase participation in the Spanish National Register of Cutaneous Melanoma.

Funding

This study was promoted and funded by the AEDV's Healthy Skin Foundation. No other entity has funded this project or participated in its design or execution.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.adengl.2020.12.009>.

References

1. Tejera-Vaquero A, Descalzo-Gallego MA, Otero-Rivas MM, et al. Incidencia y mortalidad del cáncer cutáneo en España: revisión sistemática y metaanálisis. *Actas Dermosifiliogr.* 2016;107(4):318–28, <http://dx.doi.org/10.1016/j.ad.2015.12.008>.
2. Ríos L, Nagore E, López JL, et al. Registro nacional de melanoma cutáneo. Características del tumor en el momento del diagnóstico: 15 años de experiencia. *Actas Dermosifiliogr.* 2013;104(9):789–99, <http://dx.doi.org/10.1016/j.ad.2013.02.003>.
3. Harris PA, Taylor R, Minor BL, et al. The REDCap consortium: Building an international community of software platform partners. *J Biomed Inform.* 2019, <http://dx.doi.org/10.1016/j.jbi.2019.103208>.
4. Renshaw AA, Mena-Allauca M, Gould EW, Sirintrapun SJ. Synoptic reporting: Evidence-based review and

- future directions. *JCO Clin Cancer Informatics*. 2018, <http://dx.doi.org/10.1200/cci.17.00088>.
5. Nagore E, Monteagudo C, Pinazo MI, et al. Propuesta de protocolo para el informe histológico del tumor primario de los pacientes con un melanoma cutáneo del Grupo de Trabajo para el Melanoma Cutáneo de la Comunidad Valenciana. *Actas Dermosifiliogr*. 2007, [http://dx.doi.org/10.1016/s0001-7310\(07\)70108-4](http://dx.doi.org/10.1016/s0001-7310(07)70108-4).
 6. Scolyer RA, Rawson RV, Gershenwald JE, Ferguson PM, Prieto VG. Melanoma pathology reporting and staging. *Mod Pathol*. 2020, <http://dx.doi.org/10.1038/s41379-019-0402-x>.
 7. Shon W., Frishberg D., Gershenwald J.E., et al. Cancer Protocol Templates | College of American Pathologists. <https://www.cap.org/protocols-and-guidelines/cancer-reporting-tools/cancer-protocol-templates>. Accessed May 26, 2020.
 8. Cook MG, Massi D, Szumera-Ciećkiewicz A, et al. An updated European Organisation for Research and Treatment of Cancer (EORTC) protocol for pathological evaluation of sentinel lymph nodes for melanoma. *Eur J Cancer*. 2019, <http://dx.doi.org/10.1016/j.ejca.2019.03.010>.
 9. Slater D, Walsh M. <https://www.rcpath.org/uploads/assets/fb177728-072d-4b8a-97ae94319eaac5fd/Dataset-for-the-histological-reporting-of-primary-cutaneous-malignant-melanoma-and-regional-lymph-nodes.pdf>, 2019.
 10. Valera Ruiz M, Díaz Bravo L, García Durán R. Descripción y usos del método Delphi en investigaciones del área de la salud. *Investig educ médica*. 2012;1(2):90–5.
 11. Amin M, Edge S, Greene F, et al. *AJCC Cancer Staging Manual*. 8th ed New York: Springer; 2017.
 12. Starz H, Siedlecki K, Balda BR. Sentinel lymphonectomy and s-classification: a successful strategy for better prediction and improvement of outcome of melanoma. *Ann Surg Oncol Off J Soc Surg Oncol*. 2004, <http://dx.doi.org/10.1245/aso.2004.12.920>.
 13. Baum C, Weiss C, Gebhardt C, et al. Sentinel node metastasis mitotic rate (SN-MMR) as a prognostic indicator of rapidly progressing disease in patients with sentinel node-positive melanomas. *Int J Cancer*. 2017, <http://dx.doi.org/10.1002/ijc.30563>.
 14. Lee N, Zakka LR, Mihm MC, Schatton T. Tumour-infiltrating lymphocytes in melanoma prognosis and cancer immunotherapy. *Pathology*. 2016, <http://dx.doi.org/10.1016/j.pathol.2015.12.006>.
 15. Clark WH, Elder DE, Guerry D, et al. Model predicting survival in stage I melanoma based on tumor progression. *J Natl Cancer Inst*. 1989, <http://dx.doi.org/10.1093/jnci/81.24.1893>.
 16. Balch CM, Soong SJ, Gershenwald JE, et al. Prognostic factors analysis of 17,600 melanoma patients: validation of the American Joint Committee on Cancer melanoma staging system. *J Clin Oncol*. 2001;19(16):3622–34 <http://www.ncbi.nlm.nih.gov/pubmed/11504744>
 17. National Comprehensive Cancer Network. Melanoma (Version 1.2019). https://www.nccn.org/professionals/physician_gls/PDF/melanoma.pdf. Accessed January 15, 2019.
 18. Mills JK, White I, Diggs B, Fortino J, Vetto JT. Effect of biopsy type on outcomes in the treatment of primary cutaneous melanoma. *Am J Surg*. 2013, <http://dx.doi.org/10.1016/j.amjsurg.2013.01.023>.
 19. Moreno-Ramírez D, Ojeda-Vila T, Riós-Martín JJ, Nieto-García A, Ferrándiz L. Association between tumor size and Breslow's thickness in malignant melanoma: A cross-sectional, multicenter study. *Melanoma Res*. 2015, <http://dx.doi.org/10.1097/CMR.000000000000184>.
 20. Crocetti E, Fancelli L, Caldarella A, Buzzoni C. Thickness and diameter in melanoma: Is there a relation? *Tumori*. 2016, <http://dx.doi.org/10.5301/tj.5000369>.
 21. Voss B, Wilop S, Jonas S, et al. Tumor volume as a prognostic factor in resectable malignant melanoma. *Dermatology*. 2014, <http://dx.doi.org/10.1159/000356121>.
 22. Tejera-Vaquerizo A, Cañueto J, Toll A, et al. Estimación del efecto en el tamaño y la supervivencia de los tumores cutáneos debido al confinamiento por COVID-19: Modelo basado en un crecimiento exponencial. *Actas Dermosifiliogr*. 2020, <http://dx.doi.org/10.1016/j.ad.2020.05.001>.
 23. Nosrati A, Berliner JG, Goel S, et al. Outcomes of melanoma in situ treated with Mohs micrographic surgery compared with wide local excision. *JAMA Dermatology*. 2017, <http://dx.doi.org/10.1001/jamadermatol.2016.6138>.
 24. Gershenwald JE, Scolyer RA, Hess KR, et al. Melanoma staging: Evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. *CA Cancer J Clin*. 2017, <http://dx.doi.org/10.3322/caac.21409>.
 25. Dodds TJ, Lo S, Jackett L, Nieweg O, Thompson JF, Scolyer RA. Prognostic significance of periadnexal extension in cutaneous melanoma and its implications for pathologic reporting and staging. *Am J Surg Pathol*. 2018, <http://dx.doi.org/10.1097/PAS.0000000000000999>.
 26. Ferrara G. The histopathological diagnosis. In: Argenziano G, Lallas A, Longo C, Moscarella E, Kyrgidis A, Ferrara G, editors. *Cutaneous Melanoma: A Pocket Guide for Diagnosis and Management*. Oxford: Academy Press; 2017. p. 63–80.
 27. In' t Hout FE, Haydu LE, Murali R, Bonenkamp JJ, Thompson JF, et al. Prognostic importance of the extent of ulceration in patients with clinically localized cutaneous melanoma. *Ann Surg*. 2012;255(6):1165–70, <http://dx.doi.org/10.1097/SLA.0b013e31824c4b0b>.
 28. Namikawa K, Aung PP, Gershenwald JE, Milton DR, Prieto VG. Clinical impact of ulceration width, lymphovascular invasion, microscopic satellitosis, perineural invasion, and mitotic rate in patients undergoing sentinel lymph node biopsy for cutaneous melanoma: a retrospective observational study at a comprehensive cancer center. *Cancer Med*. 2018, <http://dx.doi.org/10.1002/cam4.1320>.
 29. Clark WH, From L, Bernardino EA, Mihm MC. The histogenesis and biologic behavior of primary human malignant melanomas of the skin. *Cancer Res*. 1969;29(3):705–27 <http://www.ncbi.nlm.nih.gov/pubmed/5773814>
 30. Balch CM, Buzaid AC, Soong SJ, et al. Final version of the American Joint Committee on Cancer staging system for cutaneous melanoma. *J Clin Oncol*. 2001, <http://dx.doi.org/10.1200/JCO.2001.19.16.3635>.
 31. Azzola MF, Shaw HM, Thompson JF, et al. Tumor mitotic rate is a more powerful prognostic indicator than ulceration in patients with primary cutaneous melanoma: An analysis of 3661 patients from a single center. *Cancer*. 2003;97(6):1488–98, <http://dx.doi.org/10.1002/cncr.11196>.
 32. Francken AB, Shaw HM, Thompson JF, et al. The prognostic importance of tumor mitotic rate confirmed in 1317 patients with primary cutaneous melanoma and long follow-up. *Ann Surg Oncol*. 2004;11(4):426–33, <http://dx.doi.org/10.1245/ASO.2004.07.014>.
 33. Tejera-Vaquerizo A, Ribero S, Puig S, et al. Survival analysis and sentinel lymph node status in thin cutaneous melanoma: A multicenter observational study. *Cancer Med*. 2019, <http://dx.doi.org/10.1002/cam4.2358>.
 34. Elder DE, Bastian BC, Cree IA, Massi D, Scolyer RA. The 2018 World Health Organization classification of cutaneous, mucosal, and uveal melanoma: Detailed analysis of 9 distinct subtypes defined by their evolutionary pathway. *Arch Pathol Lab Med*. 2020, <http://dx.doi.org/10.5858/arpa.2019-0561-ra>.
 35. Azimi F, Scolyer RA, Rumcheva P, et al. Tumor-infiltrating lymphocyte grade is an independent predictor of sentinel lymph node status and survival in patients with

- cutaneous melanoma. *J Clin Oncol*. 2012;30(21):2678–83, <http://dx.doi.org/10.1200/JCO.2011.37.8539>.
36. Chen JY, Hruby G, Scolyer RA, et al. Desmoplastic neurotropic melanoma: A clinicopathologic analysis of 128 cases. *Cancer*. 2008, <http://dx.doi.org/10.1002/cncr.23895>.
 37. Nagore E, Oliver V, Botella-Estrada R, Moreno-Picot S, Insa A, Fortea JM. Prognostic factors in localized invasive cutaneous melanoma: High value of mitotic rate, vascular invasion and microscopic satellitosis. *Melanoma Res*. 2005, <http://dx.doi.org/10.1097/00008390-200506000-00005>.
 38. Petersson F, Diwan AH, Ivan D, et al. Immunohistochemical detection of lymphovascular invasion with D2-40 in melanoma correlates with sentinel lymph node status, metastasis and survival. *J Cutan Pathol*. 2009, <http://dx.doi.org/10.1111/j.1600-0560.2008.01242.x>.
 39. Landsberg J, Tüting T, Barnhill RL, Lugassy C. The role of neutrophilic inflammation, angiotropism, and pericytic mimicry in melanoma progression and metastasis. *J Invest Dermatol*. 2016;136(2):372–7, <http://dx.doi.org/10.1016/j.jid.2015.11.013>.
 40. Aung PP, Nagarajan P, Prieto VG. Regression in primary cutaneous melanoma: Etiopathogenesis and clinical significance. *Lab Invest*. 2017, <http://dx.doi.org/10.1038/labinvest.2017.8>.
 41. Requena C, Botella-Estrada R, Traves V, Nagore E, Almenar S, Guillén C. Regresión en el Melanoma: Problemas en su Definición e Implicación Pronóstica. *Actas Dermosifiliogr*. 2009;100(9):759–66 <http://www.ncbi.nlm.nih.gov/pubmed/19889297>
 42. Friedman EB, Dodds TJ, Lo S, et al. Correlation between surgical and histologic margins in melanoma wide excision specimens. *Ann Surg Oncol*. 2019;26(1):25–32, <http://dx.doi.org/10.1245/s10434-018-6858-y>.
 43. Ellison PM, Zitelli JA, Brodland DG. Mohs micrographic surgery for melanoma: A prospective multicenter study. *J Am Acad Dermatol*. 2019, <http://dx.doi.org/10.1016/j.jaad.2019.05.057>.
 44. Puza CJ, Josyula S, Terando AM, et al. Does the number of sentinel lymph nodes removed affect the false negative rate for head and neck melanoma? *J Surg Oncol*. 2018, <http://dx.doi.org/10.1002/jso.25025>.
 45. van der Ploeg AP, van Akkooi AC, Rutkowski P, et al. Prognosis in patients with sentinel node-positive melanoma is accurately defined by the combined Rotterdam tumor load and Dewar topography criteria. *J Clin Oncol*. 2011;29(16):2206–14, <http://dx.doi.org/10.1200/JCO.2010.31.6760>.
 46. van Akkooi ACJ, Spatz A, Eggermont AMM, Mihm M, Cook MG. Expert opinion in melanoma: The sentinel node; EORTC Melanoma Group recommendations on practical methodology of the measurement of the microanatomic location of metastases and metastatic tumour burden. *Eur J Cancer*. 2009, <http://dx.doi.org/10.1016/j.ejca.2009.08.015>.
 47. Dewar DJ, Newell B, Green MA, Topping AP, Powell BWEM, Cook MG. The microanatomic location of metastatic melanoma in sentinel lymph nodes predicts nonsentinel lymph node involvement. *J Clin Oncol*. 2004, <http://dx.doi.org/10.1200/JCO.2004.12.177>.
 48. Crookes TR, Scolyer RA, Lo S, Drummond M, Spillane AJ. Extracodal spread is associated with recurrence and poor survival in Stage III cutaneous melanoma patients. *Ann Surg Oncol*. 2017, <http://dx.doi.org/10.1245/s10434-016-5723-0>.
 49. Sandro P, Andrea M, Nicola M, et al. Lymph-node ratio in patients with cutaneous melanoma: A multi-institution prognostic study. *Ann Surg Oncol*. 2015, <http://dx.doi.org/10.1245/s10434-014-4132-5>.
 50. Poveda-Montoyo I, García-Doval I, Descalzo M, et al. Indicadores de calidad en la atención dermatológica a pacientes con dermatitis atópica. Documento de consenso de la AEDV. *Actas Dermosifiliogr*. 2020, <http://dx.doi.org/10.1016/j.ad.2019.06.007>.
 51. Gershenwald JE, Scolyer RA, Hess KR, et al. Melanoma staging: Evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. *CA Cancer J Clin*. 2017;67(6):472–92, <http://dx.doi.org/10.3322/caac.21409>.
 52. Elder DE, Massi D, Scolyer RWR. WHO Classification of skin tumours, Volume 11. In: *Skin Tumours. Pathology and Genetics*. Lyon (France): IARCPress; 2018.