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REVIEW

Skin Biopsy in the Context of Systemic Disease[☆]

A Fernandez-Flores^{a,b,c}



^a Servicio de Anatomía Patológica, Hospital El Bierzo, Ponferrada, León, Spain

^b Instituto de Investigación Biomédica de A Coruña, Grupo de Investigación CellCOM-SB, A Coruña, Spain

^c Servicio de Anatomía Patológica, Hospital de la Reina, Ponferrada, León, Spain

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Kikuchi-Fujimoto disease

Abstract The skin is the largest and most exposed organ in the human body and the ideal place to look for signs that aid in the early diagnosis of systemic diseases with cutaneous effects. As the concepts that underpin our understanding of many of these diseases have evolved or expanded in recent years, there have also been changes in the criteria we use for early diagnosis, including our approaches to skin biopsy and dermatopathologic evaluation. This review focuses on some of the systemic processes with skin manifestations for which our basic understanding has changed most in recent decades.

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PALABRAS CLAVE

Enfermedad autoinflamatoria;
Lupus eritematoso;
Granuloma de sílice;
Nevus de Wiesner;
BAPoma;
Síndrome de Muir-Torre;
Enfermedad de Kikuchi-Fujimoto

La biopsia cutánea en el contexto de la enfermedad sistémica

Resumen La piel es el órgano más extenso y más expuesto del cuerpo humano. Esto implica un magnífico terreno para el diagnóstico precoz de las enfermedades sistémicas que cursan con afectación sistémica y para las cuales, la piel se vuelve un marcador diagnóstico. Las bases conceptuales así como los criterios diagnósticos de muchas de estas entidades se han visto modificados o ampliados en los últimos años, con lo que la aproximación a la biopsia cutánea y la evaluación de los signos dermatopatológicos útiles en el diagnóstico precoz, han variado también. En esta revisión intentamos hacer un enfoque de algunos de los procesos sistémicos con repercusión cutánea que más han variado conceptualmente en las últimas décadas.

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E-mail address: dermatopathonline@gmail.com

Introduction

The skin is the largest and most exposed organ of the body. As a result, it is also the first to manifest many systemic conditions.

A review of all the systemic diseases that have skin manifestations would be impossible even in a book chapter. In the present article, we will focus on some recent or novel aspects of certain multiorgan diseases with skin involvement, as well as management of the best characterized of these in recent years, with a new conceptual framework for many of them.

Pathological Focus of Autoinflammatory Diseases

Autoinflammatory diseases are chronic, heterogeneous disorders arising from an abnormal phenotype in innate immunity.¹ This results in a defect in the inflammasomes (multiprotein oligomers of myeloid cells), in turn causing an excessive inflammatory response, regulated by interleukin 1, to a diverse group of triggers.

Theoretically, these disorders are not associated with infectious, allergic, autoimmune, or immunodeficient processes. However, according to current thinking, most autoimmune and autoinflammatory disorders are actually mixed entities on a continuous spectrum.

Originally, 3 main types of autoinflammatory diseases were identified: Sweet syndrome (aseptic febrile neutrophilic dermatosis), pyoderma gangrenosum, and hidradenitis suppurativa. Over the last few decades, several entities associated with vascular diseases, certain genetic mutations, vasculitis, myelodysplasia, hereditary disorders, and solid tumors have been added to the list. Diseases now considered autoinflammatory diseases include erythema elevatum diutinum, dermatosis/arthritis syndromes associated with intestinal diseases, erysipeloid conditions associated with familial Mediterranean fever, urticarial rash, and periodic syndromes associated with cryopyrin or tumor necrosis factor receptor.

Two entities that have been reclassified as autoinflammatory diseases are interstitial granulomatous dermatitis and palisaded neutrophilic granulomatous dermatitis, which are now considered to lie at either end of a continuous morphologic spectrum.²

To summarize, autoinflammatory diseases are distributed over a broad spectrum. At one end of the spectrum are rare pure monogenic autoinflammatory diseases (DITRA, DIRA, Blau syndrome, FMF, TRAPS, HIDS, PAPA...), while at the other are rare monogenic autoimmune diseases (ALPS, IPEX, APCED...). In between there is a whole range of diseases with a combination of autoimmune and autoinflammatory components but a predominance of one or the other. Predominantly autoimmune diseases include classic polygenic autoimmune diseases (rheumatoid arthritis, pernicious anemia, autoimmune thyroiditis, Addison disease, myasthenia gravis, ANCA vasculitis, systemic lupus erythematosus, Sjögren syndrome, type 1 diabetes mellitus, gluten intolerance). Autoinflammatory diseases include classic polygenic diseases such as Crohn disease, ulcerative colitis, gout, reactive arthritis unrelated to major

histocompatibility complex, vasculitis without antibodies, and idiopathic uveitis. Diseases in the mixed group with both autoimmune and autoinflammatory components in equal measure include reactive arthritis, ankylosing spondyloarthritis, psoriatic arthritis, Behcet syndrome, and HLA-B27-associated uveitis.

Several new entities have been included in the group of monogenic autoinflammatory diseases, such as pyogenic arthritis, pyoderma gangrenosum, and acne (PAPA), pyogenic arthritis, pyoderma gangrenosum, acne, and suppurative hidradenitis (PAPASH),³ pyoderma gangrenosum, acne, suppurative hidradenitis (PASH),⁴ and psoriatic arthritis, pyoderma gangrenosum, acne, and suppurative hidradenitis (PsPASH).

Skin manifestations in autoinflammatory diseases are frequent, and in many cases an essential part of diagnosis. These manifestations include urticarial and maculopapular rash, pustules, cold sores, pyoderma gangrenosum, granulomatous lesions, and erysipelas-like lesions. Although none of these are pathognomonic of a specific syndrome, neutrophilic infiltrate is common to all of them.

Innate immunity is governed by neutrophils, basophils, and eosinophils, and so it is not unexpected that polymorphonuclear neutrophils, with or without vasculitis, are one of the main pathologic markers of autoimmune diseases. Although the presence of such markers is often obvious in a routine preparation with hematoxylin-eosin, special techniques, such as myeloperoxidase, may at times be required for diagnosis.⁵ This is because the neutrophil component is fragile and neutrophils may appear flattened, fragmented, or masked by more resistant cells such as lymphocytes, and so may resemble chronic infiltrate instead of an acute one.

Some studies suggested that neutrophilic infiltrates in autoinflammatory diseases are clonal or pseudoclonal,⁶ although this line of investigation has been subject of controversy in subsequent years.

Neutrophilic infiltrates can be found in different layers of the skin, depending on the entity. Thus, they are preferentially epidermal in pustular psoriasis, generalized exanthematous pustulosis (Fig. 1), keratoderma blennorrhagicum, subcorneal pustular dermatosis (Sneddon-Wilkinson), IgA pemphigus, amicrobial pustulosis of the folds, infantile acropustulosis, and transient neonatal pustulosis. By contrast, the neutrophilic infiltrate is essentially dermal in entities such as Sweet syndrome, pyoderma gangrenosum, Behcet disease, arthritis-dermatosis associated with inflammatory bowel disease, neutrophilic eccrine hidradenitis, rheumatoid neutrophilic dermatitis, neutrophilic urticaria, Still disease, erythema marginatum, and hereditary periodic fever syndrome. Even within this last group, both the density and the depth of the dermal infiltrate may vary, being more superficial in Sweet syndrome—see for example Fig. 1B,c—while presentation may be deeper in Behcet disease or infiltrate the entire dermis and penetrate the hypodermis in pyoderma gangrenosum.

Atypical Skin Manifestations of Still Disease

Still disease is a systemic inflammatory disorder of unknown etiology that presents clinically with peaks of

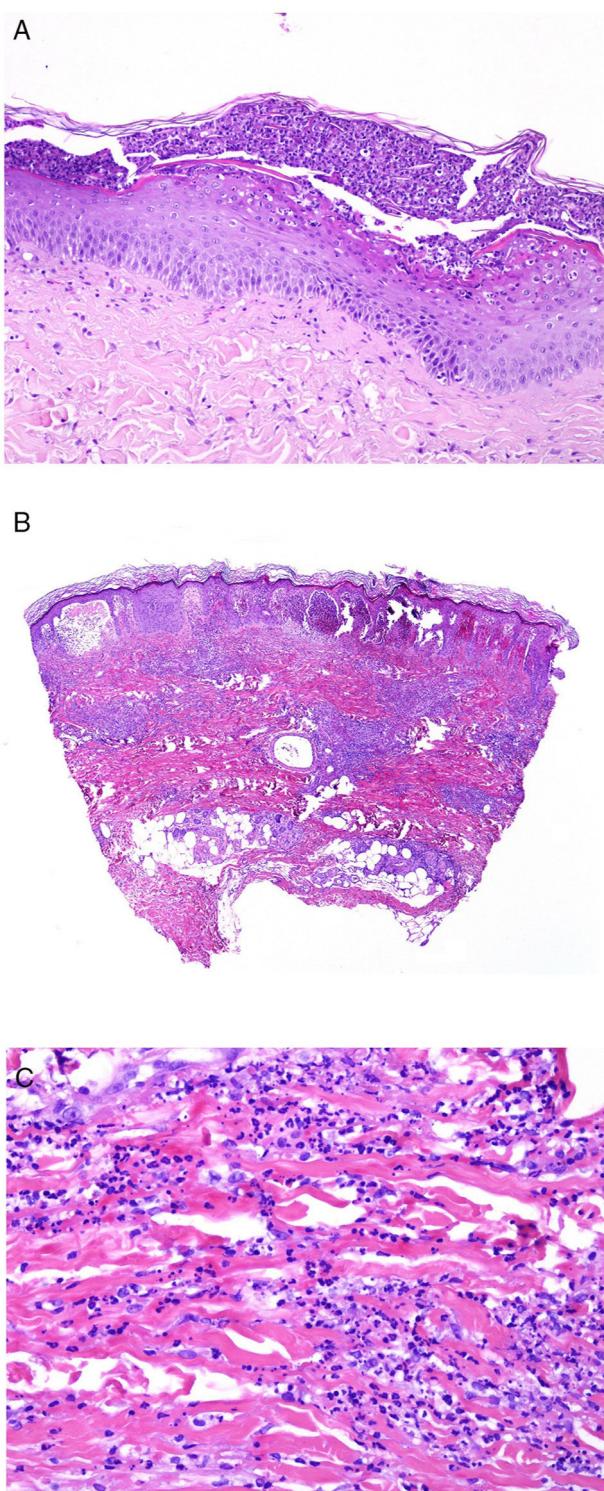


Fig. 1 A, Preferentially epidermal neutrophilic infiltrate in generalized exanthematous pustulosis (hematoxylin and eosin [H&E] $\times 100$). B, Diffuse dermal infiltrate rich in mature polymorphonuclear neutrophils with intense subdermal edema and areas of bleeding in the papillary dermis (H&E $\times 20$). C, At this magnification, polymorphonuclear neutrophils, as well as karyorrhexis, can be seen in greater detail (H&E $\times 200$).

intermittent fever, arthralgia, generalized swollen lymph nodes, sore throat, splenomegaly, and altered liver function.⁷ At times, the condition can be accompanied by serositis, myopericarditis, interstitial lung disease, and neurological involvement.⁷

The main skin manifestation of Still syndrome is evanescent rash comprising a salmon-pink maculopapular rash.⁸ However, atypical forms have been reported, with or without characteristic rash.⁹ Systemic symptoms may be concomitant with rash, precede rash by weeks or even years, or present after rash.

Within the atypical manifestations, the most frequent are persistent papules or plaques,¹⁰⁻¹² (at times with linear or reticular distribution),¹³ urticarial reactions,^{14,15} prurigo pigmentosa-like reactions,¹⁶ generalized persistent nonpruritic erythema, or vesiculopustular eruption.⁹

The pathologic presentation of classic evanescent rash in Still disease is nonspecific, with a superficial infiltrate that includes neutrophils, above which the epidermis appears preserved (Fig. 2A,B).

In contrast, in persistent lesions, necrotic keratinocytes are observed, either in isolation or grouped in the upper parts of the epidermis, along with a superficial perivascular lymphohistiocytic infiltrate (at times also interstitial), which may be accompanied by neutrophils and eosinophils (Fig. 2C,D).¹⁰ This pathologic pattern is also the one observed when persistent plaques present a reticulated clinical pattern.¹³

Necrotic keratinocytes have been described in the literature as a source of diagnostic error, confused with erythema multiforme.¹⁷ However, erythema multiforme is an interface dermatitis in which at least some vacuolization of the basal layer is expected and in which necrotic keratinocytes are preferentially seen in the basal layer, although some may be present in higher layers. On the other hand, these are located in the upper layers of the epidermis in Still syndrome. Nevertheless, we should remember that on rare occasions, Still disease can show a pathologic pattern of vacuolar degeneration of the basal layer with subcorneal or intracorneal pustules.¹⁸

In some cases with clinical presentation of persistent brownish-red papules and plaques, biopsies have shown mucin deposits in the reticular dermis,¹⁹ a finding that has also been reported in cases of linear maculopapular rash.²⁰

Urticariform clinical manifestations show typical pathologic characteristics of urticaria, with dermal edema and perivascular infiltrate (which is sometimes also interstitial), rich in neutrophils, without vasculitis.¹⁴

Another histopathologic pattern seen in atypical cases (mainly those that present with persistent diffuse pruritic erythema) is edema of the mid- and upper dermis with perivascular infiltrate of mononuclear predominance in which some neutrophils are observed.

The clinical description in the literature of prurigo pigmentosa-like lesions corresponds in biopsy to areas of parakeratosis and eosinophilic spongiosis in an acanthotic epidermis, isolated dyskeratotic cells in the epidermis, and a superficial perivascular dermal infiltrate comprised of neutrophils, eosinophils, and lymphocytes.¹⁶

Some atypical cases can present spongiosis with dermal edema accompanied by perivascular dermal infiltrate. These cases usually present clinically as vesiculopustular rash.

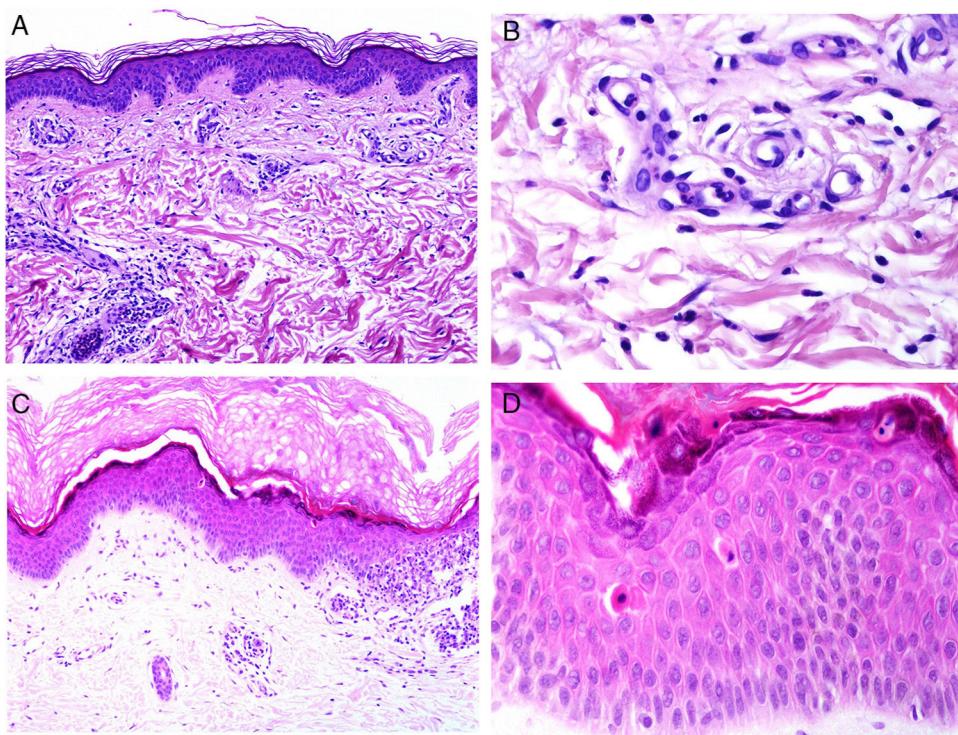


Fig. 2 A, Biopsy of an evanescent rash in a patient with Sjögren disease, showing a superficial perivascular lymphohistiocytic inflammatory infiltrate (hematoxylin and eosin [H&E] $\times 100$) with some neutrophils (B, H&E $\times 400$). C, In biopsies of patients with chronic Still disease, a superficial perivascular inflammatory infiltrate is still evident with some neutrophils (H&E $\times 100$), but necrotic keratinocytes are also observed in the upper parts of the epidermis (D, H&E $\times 400$).

Finally, in the literature, some cases have been reported with oral involvement, with brownish macules on the internal oral mucosa of the lips and on the tongue.²¹ In these cases, biopsy has shown chorionic edema with a perivascular infiltrate rich in neutrophils.

Concept of IgG4-Related Disease

IgG4-related disease is an entity described only relatively recently that presents with skin and systemic manifestations.²² In fact, it can affect almost any organ, although the most frequently involved are the pancreas and salivary and lacrimal glands.

The pathologic marker for IgG-related disease is a lymphoplasmacytic infiltrate rich in IgG4+ cells along with fibrosis of storiform pattern (Fig. 3A,B).²³ Specifically, the ratio of IgG4-IgG should be greater than 40% and IgG4+ plasma cell count should be higher than 10 per high-magnification field.²⁴ Fibrosis, in contrast, does not usually appear until the disease is advanced and, therefore absence does not exclude diagnosis. The other frequent pathologic characteristic in this entity is obliterative phlebitis.

However, this pathologic marker (richness of IgG4+ plasma cells) is controversial on its own outside a clinical context. This is because there are many entities that may present with IgG4+ cell enrichment. It has been suggested that skin diseases rich in plasma cells could actually

be forms of IgG4+ related disease, as is the case for Rosai-Dorfman disease or angiolympoid hyperplasia with eosinophilia.²³ But not all the infiltrates rich in IgG4 are IgG4-related: abundant IgG4 plasma cells can be seen in chronic inflammation of the oral mucosa or in rheumatoid arthritis.²⁵

There is evidence that IgG4 molecules are not inflammatory. Therefore, their presence seems more a secondary response to a primary process (which could be fibrosis) in individuals very likely with genetic predisposition.²⁶⁻²⁹ However, it could also be a phenomenon secondary to some immune inflammatory diseases or to chronic inflammatory processes that can present with infiltrates rich in IgG4+ plasma cells, without needing to meet the criteria for IgG4-related disease.³⁰

From the clinical point of view, the disease presents with thickening and localized or diffuse masses or tumors in multiple organs.³¹ In the skin, the presentation is varied, in the form of macules, papules, plaques, nodules, purpura, blisters, or rash, with the most frequent site being the head and neck.³² Dermatologists should be alert to this diagnosis when these manifestations are associated with idiopathic pancreatitis, retroperitoneal fibrosis, aortitis, or previously diagnosed systemic IgG4-related disease.³²

One of the most complicated differential diagnoses is systemic plasmacytosis, an entity most often occurring in Japan, which affects several organs including the skin, in the

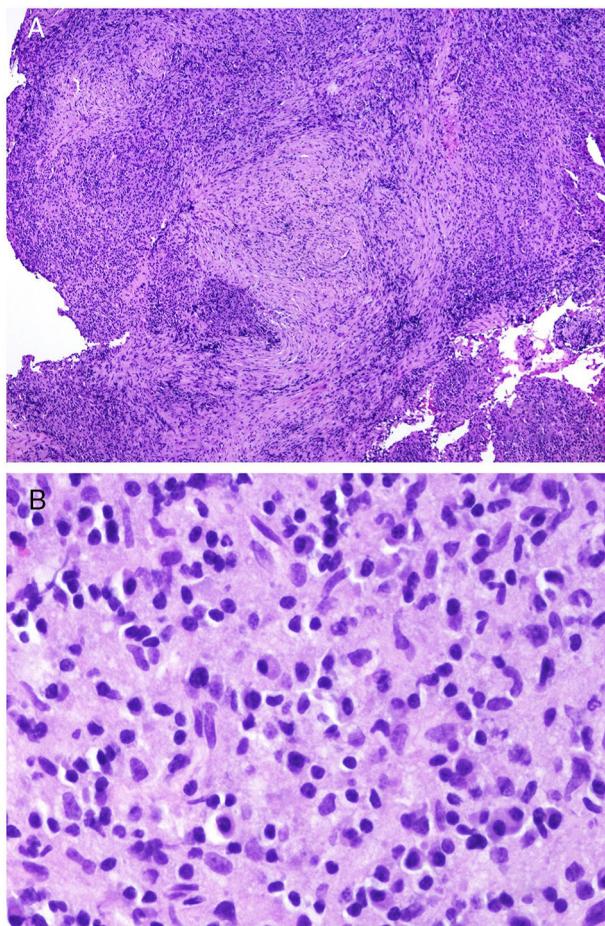


Fig. 3 A, At low magnification, IgG4-related disease should be suspected on finding an infiltrate rich in plasmacytoid dendritic cells coexisting with areas of concentric fibrosis (hematoxylin and eosin [H&E] $\times 40$). B, Infiltrate rich in plasmacytoid dendritic cells in the case of IgG4-related disease (H&E $\times 400$).

form of an eruption of reddish-brown papules preferentially distributed on the trunk.³³ In biopsy, systemic plasmacytosis is characterized by a superficial and profound perivascular and periadnexal infiltrate of mature plasma cells with polytypical expression of immunoglobulins. Given that at times, the IgG4 subpopulation is prominent in this infiltrate, it has been suggested that at least in some cases, these could be forms of IgG4-related disease.^{34–36}

Finally, we point out that IgG4-related disease is a recently described entity and so its causes and pathogenesis are not completely understood. This is probably a heterogeneous group of conditions that will likely soon be characterized.³⁷

New Advances in Biopsy of Lupus

From the clinical point of view, lupus erythematosus with skin involvement can be divided into 2 main groups: one group with mainly skin involvement (which basically includes discoid lupus erythematosus and the subacute form) and systemic lupus (for example, the acute form), which also affects internal organs.

A lengthy debate has considered whether there are histopathologic signs suggestive of systemic lupus erythematosus, and also whether there are forms of lupus confined exclusively to the skin, without any systemic impact. Progression to systemic forms in patients with chronic discoid lupus erythematosus indicates that there is no clear separation between the 2 groups. The distinction is, however, maintained because of the low risk of systemic progression of many of the cutaneous forms of lupus.

One of the forms of lupus that was considered as a primary cutaneous form without any systemic impact is lupus erythematosus tumidus, which is considered a separate entity.³⁸ However, recently, a case of lupus erythematosus tumidus was reported in which conversion occurred from discoid lupus and which supports the interchangeability of the different variants of lupus.³⁹ In contrast, other authors suggest that lupus should be considered a systemic disease with variable impact on different organs, including the skin.⁴⁰

From the pathologic point of view, there are changes that are interpreted as signs of chronic or acute disease. Thus, while systemic lupus usually shows little or no epidermal atrophy (Fig. 4A), very little vacuolar degeneration of the basal layer (Fig. 4B), absence of thickening of the basal layer, minimal horny plugs, minimal dermal edema, and minimal periadnexal infiltrate (Fig. 4C); all these features are prominent in discoid lupus erythematosus (Fig. 4D).⁴¹

Moreover, improved understanding in recent years has led to very different pathogenic scenarios of cutaneous lesions for systemic and localized forms. These 2 scenarios are, however, dynamic and may change over the course of a patient's lifetime according to external and internal factors.

Thus, migration of cytotoxic T cells and natural killer cells from circulation to the skin is characteristic of cutaneous forms but is not seen in systemic ones.⁴¹ Likewise, plasmacytoid dendritic cells, which have turned out to be very useful as a diagnostic marker of lupus, are usually abundant in cutaneous forms of lupus but not present to such an extent in skin biopsies of patients with systemic lupus, in which cells are preferentially confined to circulation. The opposite occurs with regulatory T cells, which appear very much reduced in skin biopsies of systemic lupus, but are not so reduced in lupus with skin involvement only.⁴¹

Plasmacytoid dendritic cells produce an environment with an excess of interferon alfa and beta, and these are not easy to measure in skin in everyday practice. However, myxovirus-resistant protein (MxA) seems to be a good alternative marker of levels of interferon in skin and peripheral blood.⁴²

Some of the pathologic signs of biopsies of lupus erythematosus may be of prognostic use, in addition to supporting diagnosis. For example, there is a close relationship between the number of apoptotic keratinocytes and lupus erythematosus activity (Fig. 5A).⁴¹ There is greater apoptotic activity in subacute lupus and in the systemic form, 2 of the forms in which greatest systemic activity is observed.⁴³ Furthermore, accumulation of apoptosis seems to be crucial as a trigger for skin lesions,⁴¹ contributing to

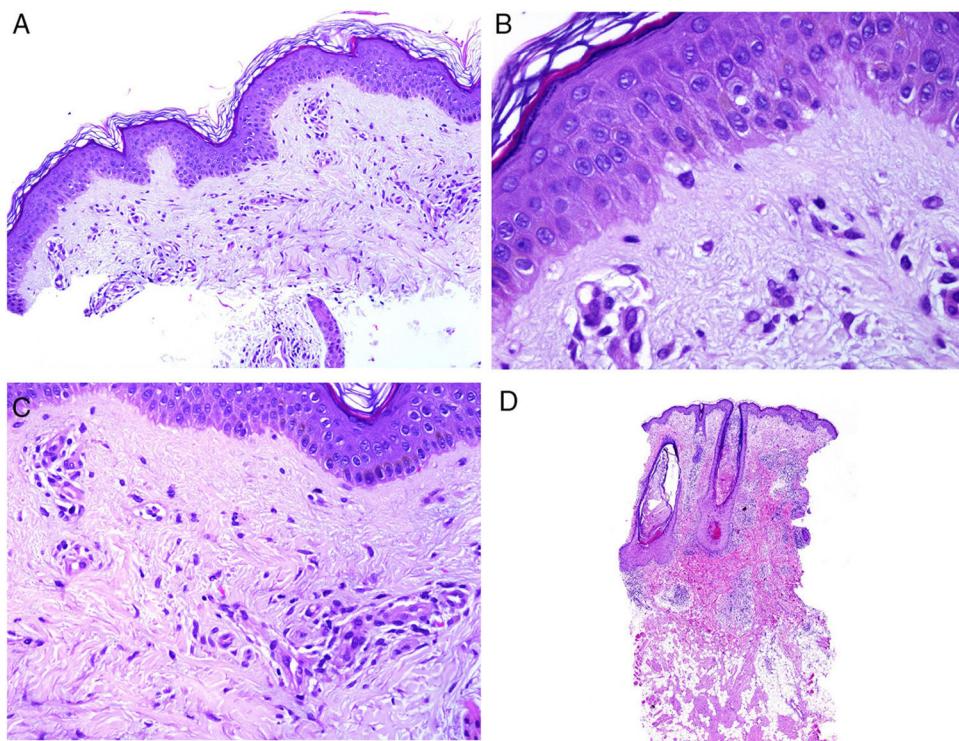


Fig. 4 Skin involvement in a case of lupus with systemic manifestations. Absence of epidermal atrophy (A, hematoxylin and eosin [H&E] $\times 100$). There is very little vacuolar degeneration of the basal layer and no evidence of thickening of the basal membrane (B, H&E $\times 400$). The dermal inflammatory infiltrate is discrete and may contain neutrophils (C, H&E $\times 200$). D, In contrast, discoid lupus erythematosus shows infundibular dilatations, prominent horny plugs, and marked periadnexal infiltrate (H&E $\times 20$).

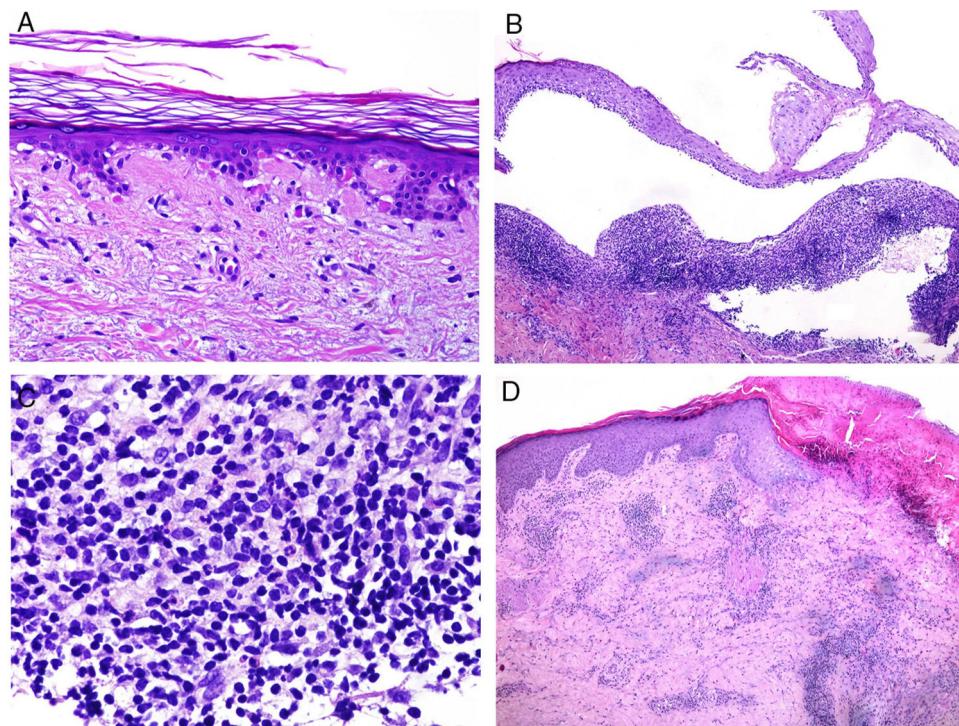


Fig. 5 A, Numerous apoptotic figures in the basal epidermal layer (hematoxylin and eosin [H&E] $\times 200$). The number of apoptotic keratinocytes has been correlated with lupus activity. B, Subepidermal detachment in a case of bullous lupus (H&E $\times 40$). C, Presence of numerous polymorphonuclear cells in chronic infiltrate from a patient with bullous lupus (H&E $\times 400$). D, Perivascular infiltrate in an ulcerated lesion corresponding to skin involvement in Kikuchi disease (H&E $\times 40$).

inflammation and immune processes on release of proinflammatory molecules such as interferon alfa, interleukin 1, and HMGB1.⁴⁴

One of the uncommon variants of lupus is the blistering form, which has recently been studied in a larger series of patients.⁴⁵ The study showed that this should be considered a systemic form, associated with lupus nephritis in half of the cases. From the pathologic point of view, in addition to subepidermal detachment (Fig. 5B), of particular interest is the infiltrate rich in polymorphonuclear neutrophils (Fig. 5C).⁴⁵ The good response obtained with dapsone in 90% of patients is therefore not surprising.⁴⁵

There are also entities in which the classification as a form of lupus erythematosus is disputed. One of these is antiphospholipid syndrome, often overdiagnosed as lupus erythematosus and, as a result, inappropriately treated. To avoid this, some authors have suggested including the requirement for at least 1 criterion of systemic lupus erythematosus before making a diagnosis of lupus in patients with antiphospholipid syndrome.⁴⁶

As in other diseases with a predominantly autoimmune component, lupus erythematosus has occasionally been associated with entities supposedly of autoinflammatory origin, such as hidradenitis suppurativa.⁴⁷

Recently, the possibility has arose of using ex vivo confocal fluorescent microscopy during the operation for a rapid diagnosis of inflammatory diseases, including lupus, with the aim of reducing delays in treatment.⁴⁸

Skin manifestations of lupus are accompanied by increased skin carcinogenesis for a variety of reasons, including the presence of chronic scarring, persistent inflammation, sensitivity to ultraviolet radiation, and use of immunosuppressants for therapeutic management.⁴⁹ However, both clinical observations of colocalization of multiple independent skin cancers on areas of active inflammation in patients with discoid lupus erythematosus in follow-up for several years, as well as murine experimental models, led to the conclusion that carcinogenesis is closely related to inflammation and that suppression or limitation of this leads to a considerable reduction in the risk of developing tumors.⁴⁹

Lupus and Kikuchi-Fujimoto Disease

Kikuchi-Fujimoto disease (histiocytic necrotizing lymphadenitis) is an idiopathic disorder. The most widely accepted etiopathogenic hypothesis is the viral/autoimmune one.⁵⁰ This is based on clinical findings (self-limiting course), laboratory findings (atypical lymphocytes in peripheral blood and increased cytokines, interferon alfa, and interleukin 6),⁵¹ and pathologic ones (predominance of T-CD8 lymphocytes in the infiltrate with apoptosis and necrotic background).⁵²⁻⁵⁴ This can be considered either an autoimmune disease or an exaggerated immune response to viral infection. Although the process usually resolves by itself, relapses may occur, sometimes many years after the first episode.⁵⁵

Kikuchi-Fujimoto disease has a disputed relationship with systemic lupus erythematosus and, sometimes, both entities are present in the same patient. In these cases, Kikuchi-

Fujimoto disease may occur before, at the same time, or after the presentation of lupus.⁵⁶ At other times, patients diagnosed with Kikuchi-Fujimoto disease meet a series of criteria for lupus in some of their episodes.^{57,58}

In addition, the lymph node changes observed in patients with systemic lupus and patients with Kikuchi-Fujimoto disease overlap, and in many cases cannot be distinguished.⁵⁸ One of the morphological findings that has been considered as providing greatest discrimination between lupus and Kikuchi-Fujimoto disease is the presence of hematoxylin bodies that mimic DNA aggregates and anti-DNA antibodies.⁵⁹ These hematoxylin bodies have mainly been observed in lymph nodes but they may also be present in biopsies of other organs, such as bone marrow or skeletal muscle.^{60,61} However, they are not a frequent feature of skin biopsies from patients with lupus erythematosus.

In contrast, skin biopsy in cases of Kikuchi-Fujimoto disease show perivascular and interstitial lymphocytic infiltrate (Fig. 5D) with presence of histiocytoid CD163+ and CD68+ cells that express myeloperoxidase.⁶² The nature of this infiltrate is currently under investigation. Necrotic foci without mature neutrophils are characteristic, as well as dermal-epidermal junction lesions.^{59,63} These latter lesions are a sign common to lupus, as are also mucin deposits and panniculitis.⁵⁹

Given this overlap, Kikuchi-Fujimoto disease may actually be an incipient form of lupus erythematosus⁶⁴ with a self-limiting and self-resolving course over a period of weeks or months in most cases.^{55,65} There are then a minority of cases with progression to severe systemic forms of Kikuchi-Fujimoto disease, some of which may be fatal, with response only to early and aggressive immunosuppressive treatment as well as cases of overlap with or progression to systemic lupus erythematosus.

Skin Biopsy in Diseases of the Digestive Tract

Probably, the best known digestive disease with skin involvement is gluten intolerance, with herpetiform dermatitis as a skin manifestation. The presence of granular IgA in the papillary dermis is due to the formation of immune complexes between IgA with avidity for the skin and the transglutaminase enzyme.⁶⁶

In 2006, Humbert et al.⁶⁷ proposed classifying skin diseases associated with gluten intolerance into 4 large groups according to their pathogenic mechanism. Those with an allergic mechanism include atopic dermatitis⁶⁸ and urticaria, whether acute or chronic.⁶⁹ Psoriasis figures among the autoimmune diseases.⁷⁰⁻⁷³ Other associated entities include rosacea and aphthous stomatitis.^{74,75} Although the list of associations is quite a lot longer, many entities have been associated only sporadically or by chance.

Another large group of digestive conditions with skin involvement are inflammatory diseases of the large intestine, with cutaneous manifestations of the granulomatous or reactive type, but also at times associated with nutritional deficiencies or iatrogenic factors.⁷⁶ Excluding the last 2 (nutritional or iatrogenic), skin involvement is observed in

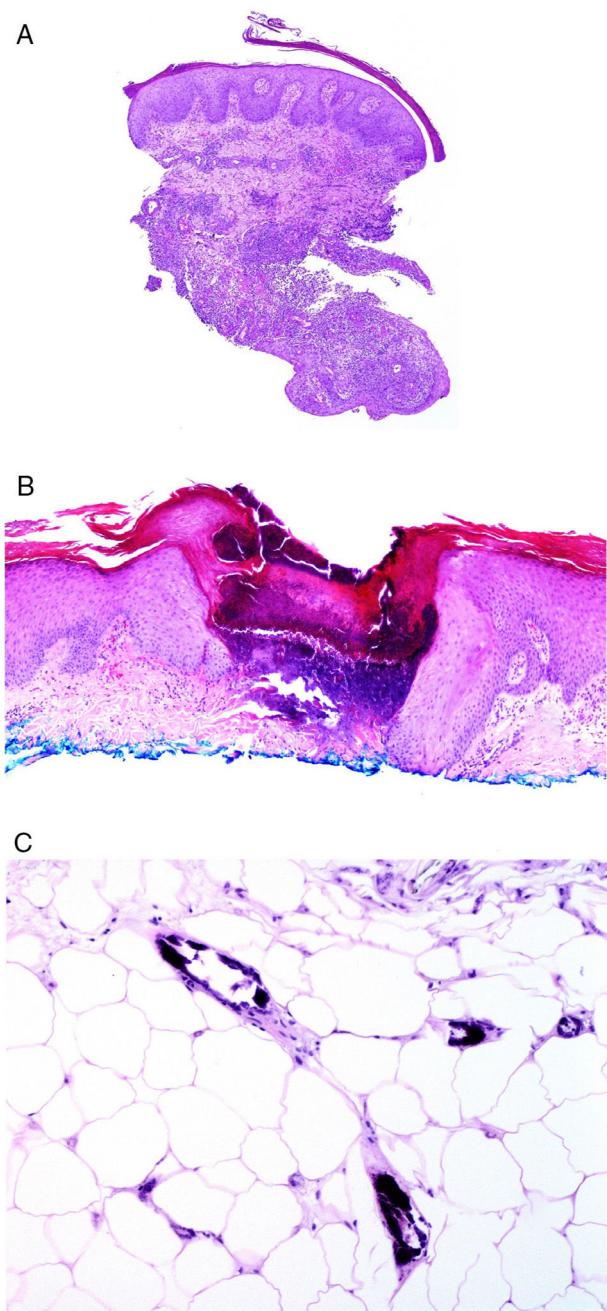


Fig. 6 A, Fistulous trajectory in biopsy of a metastatic Crohn lesion on the leg (hematoxylin and eosin [H&E] $\times 20$). B, Perforating collagenosis with elimination of eosinophilic collagen fibers by a crater of necrotic material (H&E $\times 20$). C, Calciphylaxis showing the calcified walls of small caliber subcutaneous vessels (H&E $\times 200$).

14.9% of patients with inflammatory bowel disease (Crohn disease or ulcerative colitis).⁷⁶

Involvement of skin closest to the gastrointestinal tract with noncaseating granulomas is considered specific to Crohn disease, sometimes accompanied by fistulas or abscesses (Fig. 6A).⁷⁷ These same granulomatous manifestations can, however, be seen away from the anus and mouth, given rise to the term metastatic Crohn disease. Manifestations can appear even in patients

whose gastrointestinal symptoms are well controlled with treatment.⁷⁸

In contrast, manifestations that do not show the granulomatous characteristic typical of Crohn disease but that probably correspond to skin involvement of the underlying autoinflammatory disorder are considered to be reactive. Many of these therefore show substantial dermal inflammatory infiltrate of polymorphonuclear neutrophils, such as pyoderma gangrenosum; Sweet syndrome; bowel-associated dermatosis-arthritis syndrome, pyodermititis-pyostomatitis vegetans; synovitis, acne, pustulosis, hyperostosis, osteitis (SAPHO) syndrome; and pyogenic arthritis, pyoderma gangrenosum, acne (PAPA) syndrome.⁷⁷ Reactive manifestations also include erythema nodosum of characteristic histopathology with tendency for septal panniculitis.

The forms secondary to nutritional deficiencies are considered to be due mainly to lack of vitamin A, B12, and C.⁷⁹ Finally, the drugs used in the treatment of inflammatory bowel disease are also responsible for a range of skin manifestations.

Hepatic autoimmune diseases such as primary biliary cirrhosis, primary sclerosing cholangitis, and autoimmune hepatitis, show an association with different skin manifestations: the relationship with vitiligo has been demonstrated.⁸⁰ Other probable associations include alopecia areata, psoriasis, pyoderma gangrenosum, and amyloidosis.⁸¹ In contrast to previous opinion, primary biliary cholangitis is not related to lichen planus but to amicrobial pustulosis of the skin folds.⁸⁰

Recently, it has been reported that hepatitis virus E shows endothelial tropism able to guide T lymphocytes to clonality and trigger different types of lymphoma.⁸²

Finally, we should remember that antiviral treatments in hepatitis are able to trigger systemic diseases such as lupus erythematosus⁸³ and cryoglobulemia.⁸⁴

Skin Biopsy in Renal Diseases

Skin manifestations associated with renal diseases can be classified in 3 categories: terminal renal disease, manifestations of uremia, and transplant-associated manifestations. Between 50% and 100% of patients with terminal renal disease have some type of skin manifestation.

Some of the skin manifestations are directly related to the primary underlying disease (Table 1), as is the case with diabetes, lupus erythematosus, Henoch-Schönlein purpura, granulomatosis with polyangiitis, polyarteritis nodosum, bacterial endocarditis, amyloidosis,⁸⁵ viral hepatitis, and systemic sclerosis, among others.

Among the effects associated with uremia are xerosis, pruritus, purpura, skin pigmentation, calciphylaxis, perforating diseases, porphyria and pseudoporphyria, sclerosis related to gadolinium accumulation, and alopecia. Different nail disorders, such as half-and-half nails (Lindsay nails), Muehrcke lines, absence of lunula, nail paleness, and lamina coloration disorders can also be observed.⁸⁶

Finally, the main skin manifestations related to renal transplant, in addition to rejection, are drug-associated disorders (Cushing-like changes, gingival hyperplasia, hair

Table 1 Systemic Diseases with Renal and Skin Involvement.

Systemic Disorder	Skin M	Manifestations
Diabetes mellitus		Skin infections Necrobiosis lipoidica Diabetic dermopathy Bullous diabetorum Skin thickening syndrome Acanthosis nigricans Eruptive xanthomas Kyrle disease Manifestations of diabetic vasculopathy Manifestations of diabetic neuropathy Chronic form
Lupus erythematosus		Subacute form Acute form Bullous forms Tumidus form Mucosal ulcers Purpura Urticaria White atrophy Degos Livedo reticularis Thrombophlebitis Raynaud Cutis laxa Anetoderma Papular mucinosis Amicrobial pustulosis of the folds Perniosis Neutrophilic dermatosis Purpura
Henoch-Schönlein		Macular-papular or urticarial rash Blistering forms Subcutaneous nodules
Granulomatosis with polyangiitis		Ulcers Digital ischemia Vasculitis Panniculitis Livedo reticularis
Polyarteritis nodosa		Painful subcutaneous nodules Ulcers Purpura Skin necrosis Self amputations
Subacute bacterial endocarditis		Janeway palmoplantar lesions
		Osler subcutaneous digital nodules Roth retinal stains Conjunctival and mucosal petechiae Nail splinter bleeding

Table 1 (Continued)

Systemic Disorder	Skin M	Manifestations
Amyloidosis		Periorbital purpura and in large folds White papules Sclerodermiform lesions Nail dystrophy Alopecia Blistering lesions Macroglossia
Viral hepatitis		Purpura Ictericus Palmar erythema Lichen planus Urticarial vasculitis Cryoglobulinemia Erythema nodosum Erythema multiforme Behçet syndrome Porphyria cutanea tarda Hypertrichosis Sclerodermiform plaques Generalized granuloma annulare Porokeratosis Necrolytic acral erythema Polyarteritis nodosa Scleroderma
Systemic sclerosis		Chronic edema Sclerodactyly Scarring on the heel of the hand Raynaud Telangiectasias Cutaneous hypopigmentation and hyperpigmentation Alopecia Anhidrosis

follicle changes), and those related directly to immunosuppression (infections of different types, skin tumors, porokeratosis).

One of the renal disorders with associated impact on the skin is Reed syndrome, with a tendency to renal cell carcinoma. Patients with the syndrome have associated cutaneous leiomyomas in which immune staining with fumarate hydratase is observed.⁸⁷

Chronic renal failure is closely associated with perforating skin diseases, characterized by transepidermal elimination of different dermal components (elastic fibers, collagen, etc). Traditionally, these have been classified into 4 large groups: elastosis perforans serpiginosa, reactive perforating collagenosis, perforating folliculitis, and Kyrle disease, with these 4 groups referred to by several authors with the overarching term *acquired perforating dermatosis*.

The pathogenesis of these perforating dermatoses is unknown although scratching is probably responsible in a

significant number of patients, as shown by the fact that the lesions are usually much less numerous after treatment to mitigate pruritus.

From the pathologic point of view, perforating dermatosis presents as vertical invaginations of the epidermis in the form of a short channel with acanthotic epidermis at either end. The epidermis at the base of the canal may appear preserved or, at times, eroded. The channel contains basophilic degenerated material with neutrophils and connective tissue fibers (collagen in collagenosis, elastic fibers in elastosis, none in Kyrle disease, and even elastic and collagen fibers together) (Fig. 6B). In the case of elastosis perforans serpiginosa induced by D-penicillamine, the elastic fibers adopt a characteristic beady or lumpy-bumpy pattern.

Another crucial pathologic entity in the context of renal disease is calciphylaxis, a disease with a high mortality rate. Although the pathogenesis of this disease is unknown, calcification of capillary vascular walls and small and medium caliber vessels usually in subcutaneous sites (Fig. 6C) appears to be the main cause of chronic flow that leads to acute ischemic dermal and subcutaneous necrosis. This is accompanied by vascular thrombosis which probably translates into a state of hypercoagulability. As a result, dermal angioplasty is a frequent finding in pathologic study.

To demonstrate calciphylaxis, a deep punch biopsy is required to enable assessment of vessels in the hypodermis. However, sometimes, there is a more superficial histopathologic finding that is pathognomonic of calciphylaxis: stippled perieccrine calcifications. In addition, neural calcifications can also be seen.

Finally, we note that the presence of calciphylaxis does not necessarily imply renal disease: cutaneous histopathologic findings of calciphylaxis observed in patients with and without renal disease appear to be similar.

Skin Biopsy in Sarcoidosis: The Concept of Silica Granuloma

One of the most controversial aspects in the literature is the finding of sarcoidal granulomas in skin biopsy, and whether this bears any relation to systemic sarcoidosis.

Many of these granulomas, when examined with polarized light, contain birefringent silica particles (Fig. 7A). This has given rise to the concept of silica granuloma.⁸⁸ Very recently, these deposits of sarcoidal granulomas have been characterized physically and chemically, with crystalline silica present in central areas of granuloma and calcite in the periphery.⁸⁹

For some authors, the mere presence of silica rules out systemic sarcoidosis.^{90–92} In contrast, other studies have demonstrated that silica particles are frequent in cutaneous granulomas of patients with systemic sarcoidosis.^{93,94}

Silica is an ubiquitous material in nature. It probably penetrates our skin at a very early stage in our lives, as a result of mild trauma during childhood, and has a long latency period.⁹⁵ Indeed, it is perhaps surprising that granulomatous reactions to silica are not more common.

Some studies have demonstrated that particles of silica are as frequent in cutaneous sarcoidal granulomas in patients with sarcoidosis as in those without sarcoidosis.⁹⁶ Furthermore, when skin biopsies are studied from patients with different diseases unrelated to sarcoidosis or granulomatous inflammation, silica particles are found in almost 40%.⁹⁶

As a result, it has been suggested that although silica particles could be a trigger, a predisposition is necessary, for example in patients with sarcoidosis, and so these patients are more susceptible to developing granulomas in the face of certain stimuli.⁹⁷ This mechanism has been corroborated by the description of similar granulomatous processes in patients with sarcoidosis as in processes associated with other types of particles, such as for example tattoos,⁹⁸ mesotherapy,⁹⁹ in-dwelling catheters,¹⁰⁰ and, more recently, for example the development of sarcoidal granulomas in predisposed patients at venipuncture sites.¹⁰¹ A similar process has been reported in other organs in response to other types of particles (such as talc) in patients with a tendency to develop other types of granulomatosis, such as for example Crohn disease.¹⁰²

Multiple Trichoepitheliomas and Cowden Syndrome

Cowden syndrome is a autosomal dominant genodermatosis with multiorgan involvement in which there is a risk of developing neoplasms, in particular breast, thyroid, and endometrial tumors.¹⁰³ Clinical expression of the syndrome is variable, as are the causative mutations identified so far.^{104,105} The term is not derived from the clinicians who discovered the entity but from the patient in whom it was described, Rachel Cowden, who later died of breast cancer.¹⁰⁶ Furthermore, the syndrome is characterized by multiple hamartomas at many sites, such as oral mucosa, skin, gastrointestinal tract, bone, eyes, central nervous system, and urinary tract.¹⁰⁷

The cutaneous marker of Cowden syndrome is the presence of multiple facial trichilemmomas (more than 3) (Fig. 7C), which are considered pathognomonic of the syndrome. Other pathognomonic markers are acral keratosis, oral papillomatosis, and Lhermitte-Duclos disease.

Although the etiology is unknown, a mutation in the phosphatase and tensin homolog (*PTEN*) tumor suppressor gene is observed in up to 85% of cases. The function of the gene product, the *PTEN* protein, is not entirely characterized but involves modulation of the cell cycle and cell survival. Inactivation leads to growth and excessive cell survival.

It has been suggested that *PTEN* could be used as a immunohistochemical marker for Cowden syndrome, as this gene is not expressed in up to 83% of cases of the syndrome (Fig. 7D).¹⁰⁸ However, absence of expression is not pathognomonic as it is lacking in between 3% and 13% of sporadic cases.^{108,109} In addition, there are forms of Cowden syndrome that are not associated with the *PTEN* mutation.¹¹⁰

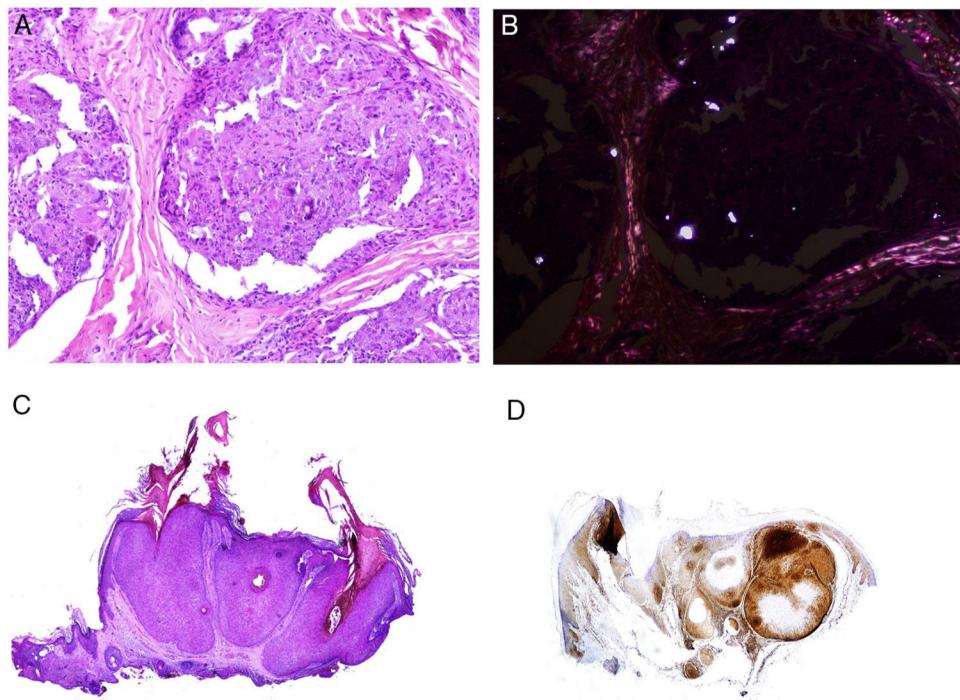


Fig. 7 A, Sarcoid epithelioid granulomatous reaction in the dermis in a case of silica granuloma (hematoxylin and eosin [H&E] $\times 100$). B, In the examination of sarcoid granulomas with polarized light, numerous intensely birefringent particles can be observed (H&E $\times 100$, polarized light). C, Typical example of trichilemmoma (H&E $\times 20$); even at this magnification, characteristic cytoplasmic cellular vacuolizations can be observed. D, Preservation of PTEN expression in a sporadic trichilemmoma, not associated with Cowden syndrome (PTEN $\times 30$).

Recently, a segmental presentation of storiform cholangiomas has been reported as part of the spectrum of the syndrome.¹¹¹

Approach In Patients With Suspicion of Muir-Torre Syndrome

Lynch syndrome, also known as hereditary non-polyposis colorectal cancer, is an autosomal dominant condition associated with mutation of the germ line of DNA mismatch repair genes, responsible for between 5% and 10% of colorectal cancers. In 90% of colorectal cancers that develop in patients with Lynch syndrome, there is a relationship with the pathogenic mechanism of microsatellite instability (MSI). The microsatellites are repeated sequences of 1 to 6 base pairs in length that appear normally in our DNA. Of note is that although the length varies from one person to another, their length is constant in a given person. If there are defects in the DNA mismatch repair proteins, we find a variation in microsatellite length in an individual. This condition is not a direct cause of tumor risk but rather a consequence, a marker, and a warning of the incorrect functioning of the DNA mismatch repair system.

Patients with Lynch syndrome have a functional allele and a defective one for DNA mismatch repair. The syndrome

develops after an incapacitating somatic mutation of the functional allele.¹¹² As this pathway implies deficient repair of genetic errors in general, individuals with Lynch syndrome have a risk of developing other cancers at an early age, with endometrial cancer being the most frequent. To a lesser extent, there is also a risk of ovarian, stomach, small intestine, pancreatic, hepatobiliary tract, urinary tract, prostate, brain, and skin cancers.

The presentation of cutaneous tumors associated with MSI is known as Muir-Torre syndrome, which is therefore considered a variant of Lynch syndrome. Autosomal dominant transmission is observed in 5% of patients, with a high degree of penetrance and variable expression. The cutaneous tumors that develop most frequently are sebaceous tumors and keratoacanthomas. The former include adenoma sebaceum, sebaceous carcinoma, and basal cell carcinoma with sebaceous differentiation. Nevus sebaceus of Jadassohn is not included among these tumors. Although sebaceous hyperplasia bears some relation to Muir-Torre syndrome, it is usually excluded from the list given that it frequently occurs in patients without the syndrome. Similar considerations apply to keratoacanthoma (both solitary and multiple presentation), spindle cell carcinoma, and multiple infundibular cysts, all associated with Muir-Torre syndrome but also frequently observed in the general population and so their usefulness as a marker is reduced. Nevertheless, some authors suggest investigat-

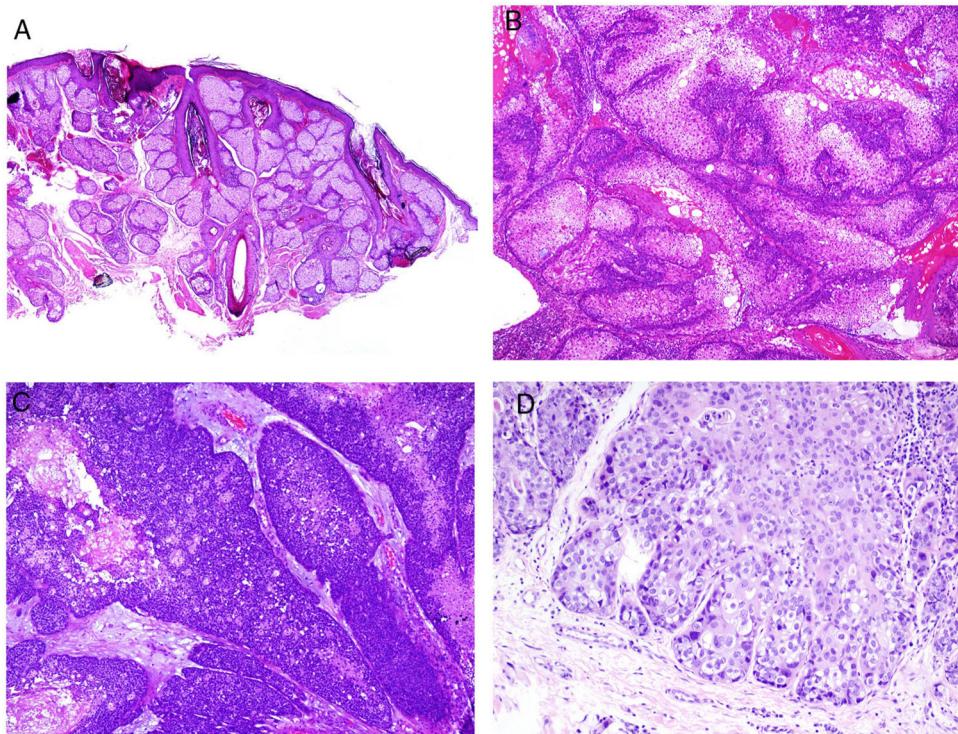


Fig. 8 A, Sebaceous hyperplasia (hematoxylin and eosin [H&E] $\times 20$). Sebaceous Muir-Torre syndrome associated neoplasms. B, Adenoma sebaceum (H&E $\times 40$). C, Sebaceoma (H&E $\times 40$). D, Sebaceous carcinoma (H&E $\times 200$).

ing whether Muir-Torre syndrome is present in cases of multiple keratoacanthoma or keratoacanthoma with sebaceous differentiation. Recently, a subgroup of tumors has been described with a particular morphological pattern (carcinoid-like) which does not seem to be related to Muir-Torre syndrome.¹¹³

Adenoma sebaceum is the sebaceous neoplasm most frequently associated with Muir-Torre syndrome. This is a multilobed dermal tumor. Each lobe appears to be surrounded by a pseudocapsule of connective tissue and is made up primarily of multivacuolated cells with a mature sebaceous appearance (Fig. 8B). At the periphery of each lobe, several layers of basophilic germ cells can be observed (in contrast with sebaceous hyperplasia which, at most, shows 2 layers of germ cells; Fig. 8A). These latter cells may sometimes have a palisade arrangement, although this is not the most frequent. Unlike adenoma sebaceum, sebaceoma mainly consists of mature basaloid cells and does not usually have a lobed architecture (Fig. 8C). However, there is a morphological continuum between adenoma sebaceum and sebaceoma, and so some authors prefer to use the unifying term of sebomatricoma. It is important to note that both neoplasms generally have a symmetric architecture (unlike sebaceous carcinoma) and, from the cytological point of view, do not show nuclear pleomorphism (Fig. 8D). Necrosis is also an important indicator of malignancy in sebaceous neoplasms, unlike mitosis, which can be seen in abundance even in benign sebaceous tumors.

Distinction should be made between the 3 aforementioned sebaceous neoplasms and basal cell carcinoma with sebaceous differentiation, which is a tumor with many of the classic basal cell attributes (peripheral palisading, stromal retraction, stroma with mucin, among others). Sebaceous differentiation is usually observed in the form of mature sebocytes in a variable quantity.

It should be clarified that sebaceous tumors associated with Muir-Torre syndrome often show ambiguous morphological characteristics that do not permit a clear classification as benign or malignant. However, this question is not usually critical if the neoplasm has been fully resected. Muir-Torre syndrome is an example of MSI.^{114,115}

To sample this condition of risk, the microsatellites of the individual can be sequenced but this is costly and laborious for a screening process. There are other more practical alternatives. Specifically, determination of DNA mismatch repair protein expression. The National Cancer Institute recommends the inclusion of 5 markers for detection of individuals with MSI¹¹⁶: *MutL homolog 1* (MLH1), *MutS protein homolog 2* (MSH2), *MutS homolog 6* (MSH6), *Postmeiotic segregation increased 2* (PMS2), and *MutL homolog 3* (MLH3). The genes that encode these proteins belong to the mismatch repair system. This is a very important system in evolutionary terms, and one that is highly preserved from bacteria to humans.

MLH1 and MSH2 defects are detected in 90% of cases of Lynch syndrome, while MSH6 defects are detected in 7%–10% and PMS2 defects in 5%.¹¹⁷ The association

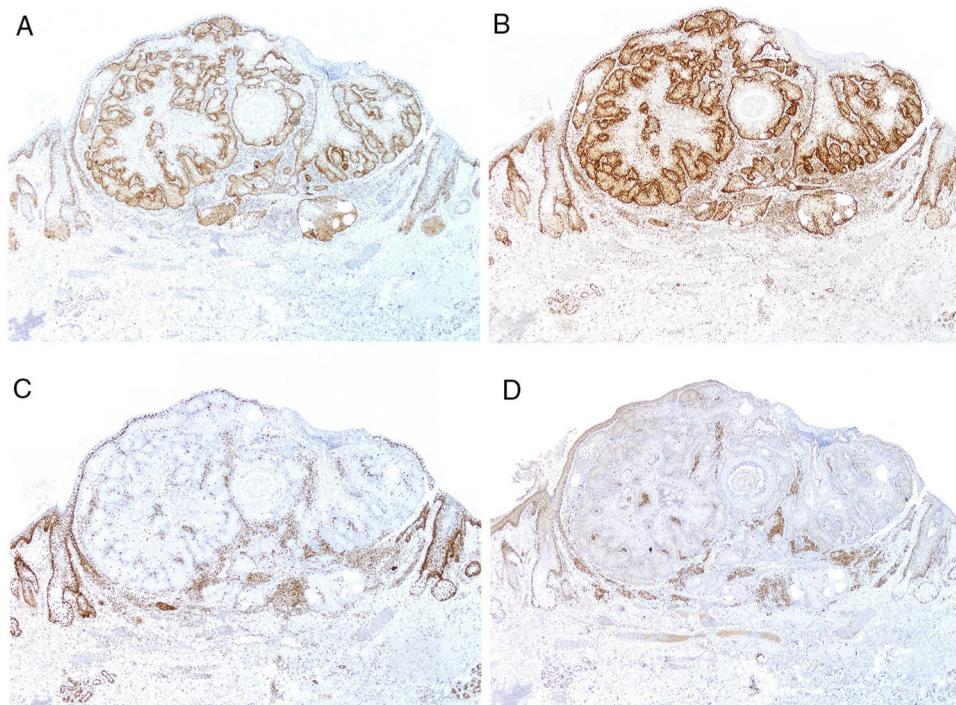


Fig. 9 Immunohistochemical study of mismatch repair markers in an adenoma sebaceum associated with Muir-Torre syndrome, demonstrating preservation of PMS2 expression (A, $\times 20$) and MLH1 expression (B, $\times 20$) with loss of MSH2 expression (C, $\times 20$) and MSH6 expression (D, $\times 20$).

with MLH3 is very small (less than 3%) and so most laboratories do not include this marker in the study. Alternatively, Lynch syndrome can be the result of epigenetic alterations,¹¹² or of sporadic somatic mutations in both alleles (rare).¹¹⁸

Interestingly, most patients with brain tumors show mutations in MSH2 with preservation of MLH1 and present a family history of brain tumors,^{119,120} and so this possibility should be investigated in clinical questioning when this phenotype is present.

Currently, most hospitals perform a first systematic screening study in all colorectal, endometrial, and small intestinal carcinomas through immunohistochemical staining for MSH2, MSH6, MLH1, and PMS2 (Fig. 9), as well as for sebaceous adenomas, sebaceous carcinomas, sebaceomas, and basal cell carcinomas with sebaceous differentiation. It is still subject of debate whether keratoacanthomas or sebaceous hyperplasia should be included.

Regardless of the result of this screening, patients with one of the aforementioned tumors should be questioned as to whether they meet the Bethesda or Amsterdam criteria or, alternatively, the Mayo Clinic risk score for Muir-Torre syndrome should be calculated.¹²¹ If this investigation is positive, a first step could be study of the mismatch repair markers by immunohistochemistry, and if these results are inconclusive, genetic and molecular analysis should be performed to rule out MSI.

The inactivity of some of the genes that encode mismatch repair proteins may also be due to hypermethylation

of their promotor. In this case, the search for a mutation will be futile. Given that BRAF mutations are frequent in sporadic colorectal cancers but infrequent in those associated with Lynch syndrome, it has been suggested that study of these mutations would be a useful step prior to study of mismatch repair markers.¹²² However, study of the BRAF V600E mutation (the most frequently investigated by laboratories) showed wild-type BRAF was present in all sebaceous neoplasms associated with Muir-Torre syndrome investigated.¹²³

The use of BRAF has also been suggested after immunohistochemistry for mismatch repair proteins in cases of loss of MLH1 expression not associated with MSH2, given the strong association between BRAF V600E mutation and methylation of the MLH1 promotor.¹²⁴

The key question, and one subject to ongoing debate in the literature, is whether systematic study is required of all resected sebaceous tumors or only those associated with Muir-Torre syndrome. The most accepted approach is to only include study of adenoma sebaceum, sebaceoma, and sebaceous carcinoma. As mentioned above, the rationale for excluding sebaceous hyperplasia and keratoacanthomas from the study is largely based on the high frequency of presentation in the population without the syndrome. However, immunohistochemical study with the 4 mismatch repair markers is currently inexpensive, and so it would be justified to include at least sebaceous hyperplasia in these studies.

BAPoma Concept

BAP-1 is the protein associated with the *BCRA-1* (*BreastCancer-1*) tumor suppressor gene. This gene encodes the mismatch repair protein BCRA-1. BAP-1 binds to BRCA-1 to form a repair complex with antitumor function.¹²⁵

Defects in the *BAP-1* gene can present as an autosomal dominant hereditary condition with as yet undetermined penetrance, in which affected individuals develop several types of somatic mutation of the functional allele,¹²⁶ showing predisposition to uveal and cutaneous melanoma and also to mesothelioma, cholangiocarcinoma, renal carcinoma, and basal cell carcinomas.

BAP-1 negative nevi were described as a cutaneous marker for the syndrome. They present as multiple, papular lesions (between 5 and 50) usually during the second decade of life.¹²⁶ The interesting point from the anatomopathologic point of view is that they can be identified by their morphological characteristics (although before the entity had been described, probably most if not all were classed as Spitz nevi). These are nevi that are preferentially dermal with extensive cytoplasm and prominent nucleoli.¹²⁷ Immunohistochemical study for *BAP-1* shows loss of nuclear positivity, leading to the paradox of using the term BAPomas when they actually do not express BAP. An alternative to this term is the eponymous Wiesner nevus, as Wiesner was the first author to describe the lesion.^{126,128}

Likewise, an uncommon presentation is as a combined nevus in which one part that is negative for *BAP-1* coexists with another part with another non-Spitzoid conventional *BAP-1* positive part.¹²⁹

Unlike in Spitz nevi, Wiesner nevi have *BRAF* mutations.^{130,131} However, as in almost all skin lesions, all information should be put into context, as not all melanocytic tumors with loss of *BAP-1* are Wiesner nevi; conventional Spitz nevi have been reported with loss of *BAP-1* expression.¹²⁹

As with many other markers, Wiesner nevi do not always present in the context of familial disease but rather can present sporadically.

Recently, a multicenter study was published of the common clinical and dermoscopic characteristics of 48 Wiesner nevi from 31 patients.¹³² The authors concluded that the entity should be suspected in the presence of cupuliform papules with amorphous pinkish-brown areas and periphery that show either irregular globules-dots or an irregular lattice.

From the histopathological point of view, Wiesner nevi are symmetric under low magnification, although as the magnification is increased, these are clearly comprised of different types of cell population with predominance of epithelioid melanocytes with broad cytoplasm and atypical hyperchromatic nucleus. Melanocytic maturation is not observed, that is, the melanocytes of the deeper areas are not smaller or more monotone than those from the upper parts of the nevus. Frequently, the contrary paradox may even arise; there appears to be an inverse maturation process, with populations of very small cells in the upper parts, reflecting a certain chaotic structuring in these nevi. The melanocytic population can be accompanied by a moderately inflammatory infiltrate.

Conclusions

Skin manifestations in systemic inflammatory processes enable an early diagnosis of these entities, allowing subsequent appropriate therapy. A different approach in recent years with some of these manifestations has allowed better management, classification, and understanding of the underlying systemic inflammatory processes.

Conflicts of Interest

The author declares that he has no conflicts of interest.

References

1. Böyükbaş B, Krause K. Cutaneous manifestations of systemic autoinflammatory disorders. Clin Dermatol. 2015;33(5):520–6.
2. Coutinho I, Pereira N, Gouveia M, Cardoso JC, Tellechea O. Interstitial granulomatous dermatitis: a clinicopathological study. Am J Dermatopathol. 2015;37(8):614–9.
3. Marzano AV. Hidradenitis suppurativa, neutrophilic dermatoses and autoinflammation: what's the link? Br J Dermatol. 2016;174(3):482–3.
4. Braun-Falco M, Kovnerysty O, Lohse P, Ruzicka T. Pyoderma gangrenosum, acne, and suppurative hidradenitis (PASH)—a new autoinflammatory syndrome distinct from PAPA syndrome. J Am Acad Dermatol. 2012;66(3):409–15.
5. Kazlouskaya V, Junkins-Hopkins JM. Lymphocytes in Sweet syndrome: a potential diagnostic pitfall. J Cutan Pathol. 2018;45(3):217–22.
6. Magro CM, Kiani B, Li J, Crowson AN. Clonality in the setting of Sweet's syndrome and pyoderma gangrenosum is not limited to underlying myeloproliferative disease. J Cutan Pathol. 2007;34(7):526–34.
7. Gerfaud-Valentin M, Maucort-Boulch D, Hot A, Iwaz J, Ninet J, Durieu I, et al. Adult-onset still disease: manifestations, treatment, outcome, and prognostic factors in 57 patients. Medicine (Baltimore). 2014;93(2):91–9.
8. Efthimiou P, Georgy S. Pathogenesis and management of adult-onset Still's disease. Semin Arthritis Rheum. 2006;36(3):144–52.
9. Narváez García FJ, Pascual M, López de Recalde M, Juarez P, Morales-Ivorra I, Notario J, et al. Adult-onset Still's disease with atypical cutaneous manifestations. Medicine (Baltimore). 2017;96(11):e6318.
10. Kikuchi N, Satoh M, Ohtsuka M, Yamamoto T. Persistent pruritic papules and plaques associated with adult-onset Still's disease: report of six cases. J Dermatol. 2014;41(5):407–10.
11. Lübbe J, Hofer M, Chavaz P, Saurat JH, Borradori L. Adult-onset Still's disease with persistent plaques. Br J Dermatol. 1999;141(4):710–3.
12. Kaur S, Bamberg P, Dhar S. Persistent dermal plaque lesions in adult onset Still's disease. Dermatology. 1994;188(3):241–2.
13. Kavusi S, Paravar T, Hasteh F, Lee R. Atypical eruption but still Still's: case report and review of the literature. Int J Dermatol. 2015;54(5):e154–159.
14. Nassereddine H, Fite C, Kottler D, Descamps vV, Couvelard A, Marot L, et al. An atypical persistent eruption of adult-onset Still's disease with neutrophilic urticarial dermatosis-like dermal features: a case report and review of the literature. J Cutan Pathol. 2018;45:793–9.
15. Setterfield JF, Hughes GR, Kobza Black A. Urticaria as a presentation of adult Still's disease. Br J Dermatol. 1998;138(5):906–8.

16. Cho YT, Liao YH. Prurigo pigmentosa-like persistent papules and plaques in a patient with adult-onset Still's disease. *Acta Derm Venereol.* 2014;94(1):102–3.
17. Khanna T, Yang CC, Yamany T, Silvers DN, Lauren CT, Lewin JM. Atypical Still disease with necrotic keratinocytes: a histologic mimicker of erythema multiforme. *JAAD Case Rep.* 2018;4(4):301–4.
18. Lee JY, Yang CC, Hsu MM. Histopathology of persistent papules and plaques in adult-onset Still's disease. *J Am Acad Dermatol.* 2005;52(6):1003–8.
19. Sun NZ, Brezinski EA, Berliner J, Haemel A, Connolly MK, Gensler L, et al. Updates in adult-onset Still disease: atypical cutaneous manifestations and associations with delayed malignancy. *J Am Acad Dermatol.* 2015;73(2):294–303.
20. Wolgamot G, Yoo J, Hurst S, Gardner G, Olerud J, Argenyi Z. Unique histopathologic findings in a patient with adult-onset Still disease. *Am J Dermatopathol.* 2007;29(2):194–6.
21. Brance ML, Neffen EL. Oral mucosa lesions as atypical manifestation of adult-onset Still's disease. *An Bras Dermatol.* 2018;93(2):271–3.
22. Stone JH. IgG4-related disease: pathophysiologic insights drive emerging treatment approaches. *Clin Exp Rheumatol.* 2016;34 Suppl 98:66–8.
23. Khosroshahi A, Deshpande V, Stone JH. The clinical and pathological features of IgG4-related disease. *Curr Rheumatol Rep.* 2011;13(6):473–81.
24. Umebara H, Kawano M. IgG4-related disease. *JOP.* 2015;16(2):217.
25. Strehl JD, Hartmann A, Agaimy A. Numerous IgG4-positive plasma cells are ubiquitous in diverse localised non-specific chronic inflammatory conditions and need to be distinguished from IgG4-related systemic disorders. *J Clin Pathol.* 2011;64(3):237–43.
26. Ota M, Katsuyama Y, Hamano H, Umemura T, Kimura A, Yoshizawa K, et al. Two critical genes (HLA-DRB1 and ABCF1) in the HLA region are associated with the susceptibility to autoimmune pancreatitis. *Immunogenetics.* 2007;59(1):45–52.
27. Okazaki K, Uchida K, Ohana M, Nakase H, Uose S, Inai M, et al. Autoimmune-related pancreatitis is associated with autoantibodies and a Th1/Th2-type cellular immune response. *Gastroenterology.* 2000;118(3):573–81.
28. Zen Y, Fujii T, Harada K, Kawano M, Yamada K, Takahira M, et al. Th2 and regulatory immune reactions are increased in immunoglobulin G4-related sclerosing pancreatitis and cholangitis. *Hepatology.* 2007;45(6):1538–46.
29. King C, Tangye SG, Mackay CR. T follicular helper (TFH) cells in normal and dysregulated immune responses. *Annu Rev Immunol.* 2008;26:741–66.
30. Daszkiewicz R, Szymoniak M, Gąsior Ł, Polański Z. Prediction of developmentally competent chromatin conformation in mouse antral oocytes. *Folia Biol (Krakow).* 2016;64(2):59–65.
31. Masaki Y, Kurose N, Yamamoto M, Takahashi H, Saeki T, Azumi A, et al. Cutoff values of serum IgG4 and histopathological IgG4+ plasma cells for diagnosis of patients with IgG4-related disease. *Int J Rheumatol.* 2012;2012:580814.
32. Charrow A, Imadojemu S, Stephen S, Ogunleye T, Takeshita J, Lipoff JB. Cutaneous manifestations of IgG4-related disease (RD): a systematic review. *J Am Acad Dermatol.* 2016;75(1):197–202.
33. Watanabe S, Ohara K, Kukita A, Mori S. Systemic plasmacytosis. A syndrome of peculiar multiple skin eruptions, generalized lymphadenopathy, and polyclonal hypergammaglobulinemia. *Arch Dermatol.* 1986;122(11):1314–20.
34. Tokura Y, Yagi H, Yanaguchi H, Majima Y, Kasuya A, Ito T, et al. IgG4-related skin disease. *Br J Dermatol.* 2014;171(5):959–67.
35. Miyagawa-Hayashino A, Matsumura Y, Kawakami F, Asada H, Tanioka M, Yoshizawa A, et al. High ratio of IgG4-positive plasma cell infiltration in cutaneous plasmacytosis—is this a cutaneous manifestation of IgG4-related disease? *Hum Pathol.* 2009;40(9):1269–77.
36. Kato K, Satoh T, Tanaka-Fujimoto T, Ueda N, Yokozeki H. IgG4-positive cells in skin lesions of cutaneous and systemic plasmacytosis. *Eur J Dermatol.* 2013;23(2):255–6.
37. Nakayama S, Matsuda M, Adachi T, Sueda S, Ohashi Y, Awaji S, et al. Adult T cell leukemia/lymphoma with different pathological features in each tumor site. *Ann Hematol.* 2018;97(6):1095–6.
38. Schmitt V, Meuth AM, Amler S, Kuehn E, Haust M, Messer G, et al. Lupus erythematosus tumidus is a separate subtype of cutaneous lupus erythematosus. *Br J Dermatol.* 2010;162(1):64–73.
39. Chen X, Wang S, Li L. A case report of lupus erythematosus tumidus converted from discoid lupus erythematosus. *Medicine (Baltimore).* 2018;97(16):e0375.
40. Ackerman A, Chongchitnant N, Sánchez J, Guo Y, Bennin B, Reichel MB, et al. Inflammatory diseases. In: Ackerman B, editor. *Histologic diagnosis of inflammatory skin diseases.* 2nd ed. Philadelphia: Williams & Wilkins; 1997. p. 170–786.
41. Li Q, Wu H, Liao W, Zhao M, Chan V, Li L, et al. A comprehensive review of immune-mediated dermatopathology in systemic lupus erythematosus. *J Autoimmun.* 2018;93:1–15.
42. al-Masri AN, Werfel T, Jakobson D, von Wussow P. Intracellular staining of Mx proteins in cells from peripheral blood, bone marrow and skin. *Mol Pathol.* 1997;50(1):9–14.
43. Baima B, Sticherling M. Apoptosis in different cutaneous manifestations of lupus erythematosus. *Br J Dermatol.* 2001;144(5):958–66.
44. Barkauskaite V, Ek M, Popovic K, Harris HE, Wahren-Herlenius M, Nyberg F. Translocation of the novel cytokine HMGB1 to the cytoplasm and extracellular space coincides with the peak of clinical activity in experimentally UV-induced lesions of cutaneous lupus erythematosus. *Lupus.* 2007;16(10):794–802.
45. de Risi-Pugliese T, Cohen Aubert F, Haroche J, Moguet P, Grootenhuis-Mignot S, Mathian A, et al. Clinical, histological, immunological presentations and outcomes of bullous systemic lupus erythematosus: 10 new cases and a literature review of 118 cases. *Semin Arthritis Rheum.* 2018;48(1):83–9.
46. Paule R, Morel N, Le Guern V, Fredi M, Coutte L, Belhocine M, et al. Classification of primary antiphospholipid syndrome as systemic lupus erythematosus: analysis of a cohort of 214 patients. *Autoimmun Rev.* 2018;17(9):866–72.
47. Ben David C, Bragazzi NL, Watad A, Sharif K, Whitby A, Amital H, et al. Hidradenitis suppurativa associated with systemic lupus erythematosus: a case report. *Medicine (Baltimore).* 2018;97(12):e0186.
48. Bertoni L, Azzoni P, Reggiani C, Pisciotta A, Carnevale G, Chester J, et al. Ex vivo fluorescence confocal microscopy for intraoperative, real-time diagnosis of cutaneous inflammatory diseases: a preliminary study. *Exp Dermatol.* 2018;27(10):1152–9.
49. Zaalberg A, Moradi Tuchayi S, Ameri AH, Ngo KH, Cunningham TJ, Eliane JP, et al. Chronic inflammation promotes skin carcinogenesis in cancer-prone discoid lupus erythematosus. *J Invest Dermatol.* 2019;139:62–70.
50. Yu HL, Lee SS, Tsai HC, Huang CK, Chen YS, Lin HH, et al. Clinical manifestations of Kikuchi's disease in southern Taiwan. *J Microbiol Immunol Infect.* 2005;38(1):35–40.
51. Găman M, Vlăduceanu AM, Dobrea C, Onisăi M, Marinescu C, Voican I, et al. A challenging case of Kikuchi-Fujimoto disease associated with systemic lupus erythematosus and review of the literature. *Case Rep Hematol.* 2018;2018:1791627.
52. Kuo TT. Kikuchi's disease (histiocytic necrotizing lymphadenitis). A clinicopathologic study of 79 cases with an analysis of

- histologic subtypes, immunohistology, and DNA ploidy. *Am J Surg Pathol.* 1995;19(7):798–809.
53. Kubota M, Tsukamoto R, Kurokawa K, Imai T, Furusho K. Elevated serum interferon gamma and interleukin-6 in patients with necrotizing lymphadenitis (Kikuchi's disease). *Br J Haematol.* 1996;95(4):613–5.
54. Kato K, Ohshima K, Anzai K, Suzumiya J, Kikuchi M. Elevated serum-soluble Fas ligand in histiocytic necrotizing lymphadenitis. *Int J Hematol.* 2001;73(1):84–6.
55. Litwin MD, Kirkham B, Henderson DR, Milazzo SC. Histiocytic necrotising lymphadenitis in systemic lupus erythematosus. *Ann Rheum Dis.* 1992;51(6):805–7.
56. Baenas DF, Diehl FA, Haye Salinas MJ, Riva V, Diller A, Lemos PA. Kikuchi-Fujimoto disease and systemic lupus erythematosus. *Int Med Case Rep J.* 2016;9:163–7.
57. Sopeña B, Rivera A, Vázquez-Triñanes C, Fluiters E, González-Carreró J, del Pozo M, et al. Autoimmune manifestations of Kikuchi disease. *Semin Arthritis Rheum.* 2012;41(6):900–6.
58. Sopeña B, Rivera A, Chamorro A, Freire M, Alende V, Seco E, et al. Clinical association between Kikuchi's disease and systemic lupus erythematosus: a systematic literature review. *Semin Arthritis Rheum.* 2017;47(1):46–52.
59. Kim JH, Kim YB, In SI, Kim YC, Han JH. The cutaneous lesions of Kikuchi's disease: a comprehensive analysis of 16 cases based on the clinicopathologic, immunohistochemical, and immunofluorescence studies with an emphasis on the differential diagnosis. *Hum Pathol.* 2010;41(9):1245–54.
60. McMullan DT, Powers JM, Nussbaum AI. Hematoxylin-stained bodies and tissue LE cells in a skeletal muscle biopsy. *Am J Clin Pathol.* 1988;90(6):731–3.
61. Xu M, Chisholm KM, Fan G, Stevens AM, Rutledge JC. Hematoxylin bodies in pediatric bone marrow aspirates and their utility in the diagnosis of systemic lupus erythematosus. *Pediatr Dev Pathol.* 2018;21(3):300–7.
62. Fernandez-Flores A, Bouso M, Alonso A, Manjon JA. The histiocytic component of cutaneous manifestations of Kikuchi disease expresses myeloperoxidase. *Appl Immunohistochem Mol Morphol.* 2008;16:202–3.
63. Atwater AR, Longley BJ, Aughenbaugh WD. Kikuchi's disease: case report and systematic review of cutaneous and histopathologic presentations. *J Am Acad Dermatol.* 2008;59(1):130–6.
64. Cramer J, Schmiedel S, Alegre NG, Schäfer H, Burchard GD, Merz H. Necrotizing lymphadenitis: Kikuchi-Fujimoto disease alias lupus lymphadenitis? *Lupus.* 2010;19(1):89–92.
65. Eisner MD, Amory J, Mullaney B, Tierney L, Browner WS. Necrotizing lymphadenitis associated with systemic lupus erythematosus. *Semin Arthritis Rheum.* 1996;26(1):477–82.
66. Reunala T, Salmi TT, Hervonen K, Kaukinen K, Collin P. Dermatitis herpetiformis: a common extraintestinal manifestation of coeliac disease. *Nutrients.* 2018;10(5).
67. Humbert P, Pelletier F, Dreno B, Puzenat E, Aubin F. Gluten intolerance and skin diseases. *Eur J Dermatol.* 2006;16(1):4–11.
68. Ciacci C, Cavallaro R, Iovino P, Sabbatini F, Palumbo A, Amoruso D, et al. Allergy prevalence in adult celiac disease. *J Allergy Clin Immunol.* 2004;113(6):1199–203.
69. Ludvigsson JF, Lindelöf B, Rashtak S, Rubio-Tapia A, Murray JA. Does urticaria risk increase in patients with celiac disease? A large population-based cohort study. *Eur J Dermatol.* 2013;23(5):681–7.
70. Ludvigsson JF, Lindelöf B, Zingone F, Ciacci C. Psoriasis in a nationwide cohort study of patients with celiac disease. *J Invest Dermatol.* 2011;131(10):2010–6.
71. Egeberg A, Griffiths CEM, Mallbris L, Gislason GH, Skov L. The association between psoriasis and coeliac disease. *Br J Dermatol.* 2017;177(6):e329–30.
72. Ungprasert P, Wijarnpreecha K, Kittanamongkolchai W. Psoriasis and risk of celiac disease: a systematic review and meta-analysis. *Indian J Dermatol.* 2017;62(1):41–6.
73. Bhatia BK, Millsop JW, Debbaneh M, Koo J, Linos E, Liao W. Diet and psoriasis, part II: celiac disease and role of a gluten-free diet. *J Am Acad Dermatol.* 2014;71(2):350–8.
74. Nieri M, Tofani E, Defraia E, Giuntini V, Franchi L. Enamel defects and aphthous stomatitis in celiac and healthy subjects: systematic review and meta-analysis of controlled studies. *J Dent.* 2017;65:1–10.
75. Egeberg A, Weinstock LB, Thyssen JP, Gislason GH, Thyssen JP. Rosacea and gastrointestinal disorders: a population-based cohort study. *Br J Dermatol.* 2017;176(1):100–6.
76. Vide J, Osório F, Costa-Silva M, Lopes S, Azevedo F, Camila Dias C, et al. Cutaneous morbidity among inflammatory bowel disease patients: a cohort study. *J Crohns Colitis.* 2018;12(4):442–51.
77. Keyal U, Liu Y, Bhatta AK. Dermatologic manifestations of inflammatory bowel disease: a review. *Discov Med.* 2018;25(139):225–33.
78. Woody MM, Holliday AC, Gavino ACP, McReynolds A, Soldano AC. Metastatic vulvovaginal Crohn disease in the setting of well-controlled intestinal disease. *Cutis.* 2018;102(2):E16–8.
79. Trost LB, McDonnell JK. Important cutaneous manifestations of inflammatory bowel disease. *Postgrad Med J.* 2005;81(959):580–5.
80. Terzioli Beretta-Piccoli B, Invernizzi P, Gershwin ME, Mainetti C. Skin manifestations associated with autoimmune liver diseases: a systematic review. *Clin Rev Allergy Immunol.* 2017;53(3):394–412.
81. Yan X, Jin J. Primary cutaneous amyloidosis associated with autoimmune hepatitis-primary biliary cirrhosis overlap syndrome and Sjögren syndrome: a case report. *Medicine (Baltimore).* 2018;97(8):e0004.
82. Mallet V, Bruneau J, Zuber J, Alanio C, Leclerc-Mercier S, Roque-Afonso AM, et al. Hepatitis E virus-induced primary cutaneous CD30(+) T cell lymphoproliferative disorder. *J Hepatol.* 2017;67(6):1334–9.
83. Prenner A, Blum R, Beltraminelli H, Stirnimann G, Borradori L. Subacute cutaneous lupus erythematosus triggered by an antiviral treatment combination for hepatitis C virus infection. *J Eur Acad Dermatol Venereol.* 2019;33:e129–31.
84. Montero N, Favà A, Rodriguez E, Barrios C, Cruzado JM, Pascual J, et al. Treatment for hepatitis C virus-associated mixed cryoglobulinaemia. *Cochrane Database Syst Rev.* 2018;5:CD011403.
85. Fernandez-Flores A. Cutaneous amyloidosis: a concept review. *Am J Dermatopathol.* 2012;34:1–14, quiz15–17.
86. Rashpa RS, Mahajan VK, Kumar P, Mehta KS, Chauhan PS, Rawat R, et al. Mucocutaneous manifestations in patients with chronic kidney disease: a cross-sectional study. *Indian Dermatol Online J.* 2018;9(1):20–6.
87. Llamas-Velasco M, Requena L, Kutzner H, Schärer L, Rütten A, Hantschke M, et al. Fumarate hydratase immunohistochemical staining may help to identify patients with multiple cutaneous and uterine leiomyomatosis (MCUL) and hereditary leiomyomatosis and renal cell cancer (HLRCC) syndrome. *J Cutan Pathol.* 2014;41(11):859–65.
88. Hou M, Zhu W, Ye Y. Cutaneous silica granuloma with generalized involvement of lymph nodes. *J Dermatol.* 2011;38(7):697–701.
89. Colboc H, Moguelet P, Bazin D, Bachmeyer C, Frochot V, Weil R, et al. Physicochemical characterization of inorganic deposits associated with granulomas in cutaneous sarcoidosis. *J Eur Acad Dermatol Venereol.* 2019;33:198–203.
90. Mowry RG, Sams WM, Caulfield JB. Cutaneous silica granuloma. A rare entity or rarely diagnosed? Report of two cases with review of the literature. *Arch Dermatol.* 1991;127(5):692–4.

91. Walsh NM, Hanly JG, Tremaine R, Murray S. Cutaneous sarcoidosis and foreign bodies. *Am J Dermatopathol.* 1993;15(3):203–7.
92. Kim YC, Triffet MK, Gibson LE. Foreign bodies in sarcoidosis. *Am J Dermatopathol.* 2000;22(5):408–12.
93. Lofgren S, Snellman B, Nordenstam H. Foreign-body granulomas and sarcoidosis; a clinical and histo-pathological study. *Acta Chir Scand.* 1955;108(6):405–18.
94. Marcoval J, Mañá J, Moreno A, Gallego I, Fortuño Y, Peyrí J. Foreign bodies in granulomatous cutaneous lesions of patients with systemic sarcoidosis. *Arch Dermatol.* 2001;137(4):427–30.
95. Eskeland G, Langmark F, Husby G. Silicon granuloma of skin and subcutaneous tissue. *Acta Pathol Microbiol Scand Suppl.* 1974; Suppl 248:69–73.
96. Fernandez-Flores A. Birefringent particles in granulomatous dermatitis, sarcoidal-type, as well as in other non-granulomatous skin disorders in patients without sarcoidosis. *Bratisl Med J.* 2009;110(6):328–31.
97. Fernandez-Flores A, Montero MG. Does cutaneous silica granuloma develop mainly in predisposed patients? *Eur J Dermatol.* 2006;16:321–2.
98. Baumgartner M, Feldmann R, Breier F, Steiner A. Sarcoidal granulomas in a cosmetic tattoo in association with pulmonary sarcoidosis. *J Dtsch Dermatol Ges.* 2010;8(11):900–2.
99. Gokdemir G, Küçükünal A, Sakiz D. Cutaneous granulomatous reaction from mesotherapy. *Dermatol Surg.* 2009;35(2):291–3.
100. Kenmochi A, Satoh T, Igawa K, Yokozeki H. Silica granuloma induced by indwelling catheter. *J Am Acad Dermatol.* 2007;57 2 Suppl:S54–5.
101. Marcoval J, Penín RM, Mañá J. Specific skin lesions of sarcoidosis located at venipuncture points for blood sample collection. *Am J Dermatopathol.* 2018;40(5):362–6.
102. Weiner J, Chandak T, Caperna A, Malieckal D, Xu W, Reder I, et al. Diffuse talc granulomatosis in a patient with Crohn's disease. *Am J Respir Crit Care Med.* 2012;186(7):e11.
103. Pilarski R, Burt R, Kohlman W, Pho L, Shannon KM, Swisher E. Cowden syndrome and the PTEN hamartoma tumor syndrome: systematic review and revised diagnostic criteria. *J Natl Cancer Inst.* 2013;105(21):1607–16.
104. Machado RA, Paranaíba LMR, Martins L, Melo-Filho MR, de Souza TT, Picciani BLS, et al. Variable expressivity and novel PTEN mutations in Cowden syndrome. *Oral Surg Oral Med Oral Pathol Oral Radiol.* 2019;127:55–61.
105. Adachi T, Takigawa H, Nomura T, Watanabe Y, Kowa H. Cowden syndrome with a novel PTEN mutation presenting with partial epilepsy related to focal cortical dysplasia. *Intern Med.* 2018;57(1):97–9.
106. Reddy KV, Anusha A, Maloth KN, Sunitha K, Thakur M. Mucocutaneous manifestations of Cowden's syndrome. *Indian Dermatol Online J.* 2016;7(6):512–5.
107. Seol JE, Park IH, Lee W, Kim H, Seo JK, Oh SH. Cowden syndrome with a novel germline PTEN mutation and an unusual clinical course. *Ann Dermatol.* 2015;27(3):306–9.
108. Al-Zaid T, Ditelberg JS, Prieto VG, Lev D, Luthra R, Davies MA, et al. Trichilemmomas show loss of PTEN in Cowden syndrome but only rarely in sporadic tumors. *J Cutan Pathol.* 2012;39(5):493–9.
109. Jin M, Hampel H, Pilarski R, Zhou X, Peters S, Frankel WL. Phosphatase and tensin homolog immunohistochemical staining and clinical criteria for Cowden syndrome in patients with trichilemmoma or associated lesions. *Am J Dermatopathol.* 2013;35(6):637–40.
110. Yehia L, Ni Y, Sesock K, Niazi F, Fletcher B, Chen HJL, et al. Unexpected cancer-predisposition gene variants in Cowden syndrome and Bannayan-Riley-Ruvalcaba syndrome patients without underlying germline PTEN mutations. *PLoS Genet.* 2018;14(4):e1007352.
111. Habeshian K, Huppmann A, Ferreira C, Kirkorian AY. Segmental storiform collagenomas: expanding the spectrum of PTEN hamartoma tumor syndrome in children. *Pediatr Dermatol.* 2018;35(4):e253–4.
112. Steinke V, Engel C, Büttner R, Schackert HK, Schmiegel WH, Propping P. Hereditary nonpolyposis colorectal cancer (HNPCC)/Lynch syndrome. *Dtsch Arztebl Int.* 2013;110(3):32–8.
113. Wiedemeyer K, Kyrychova L, Işıkci Ö, Spagnolo DV, Kutzner H, Rütten A, et al. Sebaceous neoplasms with rippled, labyrinthine/sinusoidal, petaloid, and carcinoid-like patterns: a study of 57 cases validating their occurrence as a morphological spectrum and showing no significant association with Muir-Torre syndrome or DNA mismatch repair protein deficiency. *Am J Dermatopathol.* 2018;40(7):479–85.
114. Lorente-Lavirgen AI, Morillo-Andújar M, Zulueta-Dorado T, Conejo-Mir J. Microsatellite and genetic instability in patients with Muir-Torre syndrome. *Actas Dermosifiliogr.* 2013;104(7):643–4.
115. Schwartz RA, Torre DP. The Muir-Torre syndrome: a 25-year retrospect. *J Am Acad Dermatol.* 1995;33(1):90–104.
116. Boland CR, Thibodeau SN, Hamilton SR, Sidransky D, Eshleman JR, Burt RW, et al. A National Cancer Institute Workshop on Microsatellite Instability for cancer detection and familial predisposition: development of international criteria for the determination of microsatellite instability in colorectal cancer. *Cancer Res.* 1998;58(22):5248–57.
117. Mangold E, Pagenstecher C, Leister M, Mathiak M, Rütten A, Friedl W, et al. A genotype-phenotype correlation in HNPCC: strong predominance of msh2 mutations in 41 patients with Muir-Torre syndrome. *J Med Genet.* 2004;41(7):567–72.
118. Roberts ME, Riegert-Johnson DL, Thomas BC, Thomas CS, Heckman MG, Krishna M, et al. Screening for Muir-Torre syndrome using mismatch repair protein immunohistochemistry of sebaceous neoplasms. *J Genet Couns.* 2013;22(3):393–405.
119. Lehrer MD, Lynch H, Glembotski DJ, Patel NB. Glioblastoma multiforme as initial internal malignancy in Muir-Torre syndrome (MTS). *JAAD Case Rep.* 2015;1(6):381–3.
120. Kurtzman DJ, Fabiano AJ, Qiu J, Zeitouni NC. Muir-Torre syndrome and central nervous system malignancy: highlighting an uncommon association. *Dermatol Surg.* 2015;41(7):856–9.
121. Le S, Ansari U, Mumtaz A, Malik K, Patel P, Doyle A, et al. Lynch syndrome and Muir-Torre syndrome: an update and review on the genetics, epidemiology, and management of two related disorders. *Dermatol Online J.* 2017;23(11).
122. Lynch HT, Lynch JF, Lynch PM. Toward a consensus in molecular diagnosis of hereditary nonpolyposis colorectal cancer (Lynch syndrome). *J Natl Cancer Inst.* 2007;99(4):261–3.
123. Cornejo KM, Hutchinson L, Deng A, Tomaszewicz K, Welch M, Lyle S, et al. BRAF/KRAS gene sequencing of sebaceous neoplasms after mismatch repair protein analysis. *Hum Pathol.* 2014;45(6):1213–20.
124. Parsons MT, Buchanan DD, Thompson B, Young JP, Spurdle AB. Correlation of tumour BRAF mutations and MLH1 methylation with germline mismatch repair (MMR) gene mutation status: a literature review assessing utility of tumour features for MMR variant classification. *J Med Genet.* 2012;49(3):151–7.
125. Jensen DE, Rauscher FJ. BAP1, a candidate tumor suppressor protein that interacts with BRCA1. *Ann N Y Acad Sci.* 1999;886:191–4.
126. Wiesner T, Obenau AC, Murali R, Fried I, Griewank KG, Ulz P, et al. Germline mutations in BAP1 predispose to melanocytic tumors. *Nat Genet.* 2011;43(10):1018–21.
127. Llamas-Velasco M, Pérez-González YC, Requena L, Kutzner H. Histopathologic clues for the diagnosis of Wiesner nevus. *J Am Acad Dermatol.* 2014;70(3):549–54.

128. Busam KJ, Wanna M, Wiesner T. Multiple epithelioid Spitz nevi or tumors with loss of BAP1 expression: a clue to a hereditary tumor syndrome. *JAMA Dermatol.* 2013;149(3):335–9.
129. Requena C, Sanz V, Nagore E, García-Casado Z, Rubio L, Guillén C, et al. BAP1-deficient and VE1-negative atypical Spitz tumor. *J Cutan Pathol.* 2015;42(8):564–7.
130. Aggarwal R. Diagnosis of hepatitis E. *Nat Rev Gastroenterol Hepatol.* 2013;10(1):24–33.
131. Wiesner T, Murali R, Fried I, Cerroni L, Busam K, Kutzner H, et al. A distinct subset of atypical Spitz tumors is characterized by BRAF mutation and loss of BAP1 expression. *Am J Surg Pathol.* 2012;36(6):818–30.
132. Yélamos O, Navarrete-Dechent C, Marchetti MA, Rogers T, Apalla Z, Bahadoran P, et al. Clinical and dermoscopic features of cutaneous BAP1 inactivated melanocytic tumors: results of a multicenter case-control study by the International Dermoscopy Society (IDS). *J Am Acad Dermatol.* 2018, <http://dx.doi.org/10.1016/j.jaad.2018.09.014> [Epub ahead of print].