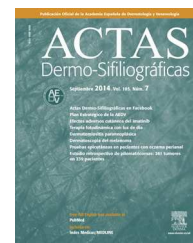




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## REVIEW

# The Role of Filaggrin in the Skin Barrier and Disease Development<sup>☆</sup>



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### PALABRAS CLAVE

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Asma;  
Rinitis alérgica

**Abstract** Filaggrin is a structural protein that is fundamental in the development and maintenance of the skin barrier. The function of filaggrin and its involvement in various cutaneous and extracutaneous disorders has been the subject of considerable research in recent years. Mutations in *FLG*, the gene that encodes filaggrin, have been shown to cause ichthyosis vulgaris, increase the risk of atopic dermatitis and other atopic diseases, and exacerbate certain conditions. The present article reviews the current knowledge on the role of filaggrin in the skin barrier, *FLG* mutations, and the consequences of filaggrin deficiency.

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### Filagrina: papel en la barrera cutánea y en el desarrollo de patología

**Resumen** La filagrina es una proteína estructural fundamental para el desarrollo y mantenimiento de la barrera cutánea. En los últimos años se ha llevado a cabo una extensa investigación sobre su función y su implicación en distintos trastornos cutáneos y extracutáneos. Se ha comprobado que las mutaciones en el gen que la codifica, el gen *FLG*, son la causa de la ictiosis vulgar y confieren un mayor riesgo de desarrollar dermatitis atópica y otras enfermedades atópicas, además de agravar algunas enfermedades. El presente artículo revisa la información existente en cuanto a su papel en la barrera cutánea, así como respecto a las mutaciones en el gen *FLG* y las consecuencias que conlleva el déficit de filagrina.

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## Introduction

The main function of the skin is to act as a barrier that separates the internal environment from the external one, thus protecting against aggression by exogenous agents and minimizing the loss of water and other essential body components to the external space.<sup>1</sup> Filaggrin is particularly important in the formation of the skin barrier, both for its

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**Table 1** Effects of Filaggrin Deficit on the Epidermal Barrier.

	Biochemical and Structural Repercussions	Consequences
Skin surface	Increased pH Increased activity of certain proteases	Increased adhesion and proliferation of staphylococci Release of epithelial proinflammatory mediators
Corneal layer	Decreased concentration of natural moisturizing factor Decreased density of corneodesmosomes and of tight intercellular junctions Abnormal architecture of the extracellular lipid matrix	Xerosis Altered barrier function Increased exposure to allergens
Stratum granulosum-stratum corneum transition region	Altered maturation and excretion of the lamellar bodies Altered aggregation of keratin intermediate filaments	Altered barrier function Increased exposure to allergens
Stratum granulosum	Decreased number of keratohyalin granules	-

fundamental role in terminal epidermal differentiation and for its implication in some of the most common dermatological diseases, such as atopic dermatitis (AD) and ichthyosis vulgaris.<sup>2</sup> Filaggrin is an important structural protein that was first identified in 1977.<sup>3</sup> Later, when it was found to produce aggregation and compaction of keratin intermediate filaments, it was named *filaggrin*, the acronym of *filament-aggregating protein*.<sup>4</sup> This protein is synthesized as a giant precursor protein called profilaggrin, which is the main component of the keratohyalin granules in the stratum granulosum of the epidermis.

### Role of Filaggrin in the Formation of the Epidermal Barrier

The main element of the skin barrier is the stratum corneum. This stratum is the end-product of the differentiation of keratinocytes, which, from the basal layer to the granulosum layer, are viable nucleated cells. These cells express various structural proteins as they mature.<sup>5</sup> In the final steps of differentiation, the keratinocytes undergo marked changes in their structure, leading to their transformation into flat, anucleate squamous cells, the corneocytes. These corneocytes, which remain tightly bound together by corneodesmosomes, are covered by a cellular coating called the *cornified envelope* (CE), which has protein and lipid components that endow the cells with mechanical and chemical resistance.<sup>5</sup> Between the cells there is a hydrophobic, lipid-rich extracellular matrix arranged in a laminar bilayer.<sup>6</sup> This organization of the stratum corneum has been called “bricks and mortar”, in which the keratinocytes are the bricks and the extracellular lipid matrix is the mortar.<sup>7</sup>

A calcium gradient exists in the epidermis, with low concentrations in the basal layer, even lower concentrations in the stratum spinosum, high levels in the stratum granulosum, and very low levels in the stratum corneum.<sup>8</sup> This gradient is important in terminal keratinocyte differentiation.<sup>2,9</sup> The higher concentration of calcium in the stratum granulosum causes the keratohyalin granules to release their contents, leaving the profilaggrin exposed to undergo

processing and fragmentation into active filaggrin monomers.<sup>10</sup> This free filaggrin binds to the intermediate keratin filaments, causing their aggregation and compaction, provoking the collapse and flattening of the cell. Simultaneously, the cell expresses a series of structural proteins that make up the protein portion of the CE.<sup>11,12</sup> The bundles of keratin intermediate filaments aggregated by filaggrin bind to these structural proteins through the action of transglutaminases.<sup>6</sup> In addition, the increase in the calcium concentration also provokes release of the contents of the lamellar bodies, which are granules rich in lipids and enzymes synthesized in the Golgi apparatus. The action of these enzymes on the lipids gives rise to the lipid portion of the CE and the extracellular matrix of the stratum corneum.<sup>6,11,13</sup>

Filaggrin continues to undergo processing by various proteases. This proteolysis leads to the release of hygroscopic amino acids and their derivatives, which form natural moisturizing factor (NMF), responsible for water retention in the stratum corneum.<sup>14</sup> The breakdown of some of these amino acids gives rise to 2 organic acids: trans-urocanic acid (UCA), a histidine derivative; and pyrrolidone-5-carboxylic acid (PCA), a glutamine derivative. These acids are 2 of the main factors responsible for maintaining the acid pH of the stratum corneum,<sup>15</sup> which is essential for its role in the metabolism and organization of the lipids of the extracellular matrix,<sup>16</sup> for its antimicrobial action, and for its regulatory role on enzyme activity and physiologic desquamation.<sup>17</sup> In addition, UCA has a photoprotective effect against UV radiation<sup>18</sup> and has been shown to be a key mediator in UV-B-induced immunosuppression.<sup>19,20</sup>

### The Epidermal Barrier and Filaggrin Deficit

Filaggrin deficit has a major impact on the epidermal barrier (Table 1), affecting the organization of the keratin filaments of the cytoskeleton and the structure of the CE. There is also a fall in the number of keratohyalin granules, a marked fall in NMF concentration (and thus in hydration of the stratum corneum), and alkalization of the skin pH.<sup>5</sup> In addition to these changes, recent ultrastructural studies

have shown that a deficit of filaggrin is also associated with a generalized fall in the density of corneodesmosomes and of tight intercellular junctions, as well as with abnormalities in the architecture of the extracellular lipid matrix; these changes may produce a notable alteration of barrier function, provoked by abnormalities in the organization of the cytoskeleton (which affect lamellar body maturation and exocytosis, leading to a nonuniform distribution of secreted lipids and enzymes) and by the increase in the pH (which modulates the activity of those enzymes).<sup>16</sup> Furthermore, the increase in the activity of certain proteases caused by the persistent elevation of the pH favors the release of proinflammatory mediators by keratinocytes; these mediators induce an inflammatory response mediated by type 2 helper T (Th2) cells even in the absence of allergens.<sup>21</sup> For example, alkalization of the skin pH increases the activity of the proteases responsible for the production of interleukin (IL) 1 $\alpha$  and IL-1 $\beta$  from their inactive proproteins generated by keratinocytes.<sup>22</sup>

### Genetics of filaggrin

Profilaggrin is coded by the *FLG* gene, located in the epidermal differentiation complex on chromosome 1 (locus 1q21), a cluster of genes that code for proteins involved in epidermal differentiation.<sup>23</sup> The *FLG* gene is made up of 3 exons and 2 introns (Fig. 1). Transcription starts at exon 2, but it is exon 3 that codes for the greatest part of the protein, constituting one of the largest exons in the genome.<sup>24</sup> The resulting protein (profilaggrin) is rich in histidine and contains between 10 and 12 repetitions of filaggrin flanked by N- and C-terminus domains (Fig. 1).

These domains are also functional. The N-terminus domain is, in turn, formed by 2 subdomains: A and B. Subdomain A contains 2 calcium binding sites.<sup>25</sup> It is the binding of calcium to this subdomain that produces a series of conformational changes in the profilaggrin molecule that will initiate its processing.<sup>2</sup> The B subdomain contains a nuclear localization signal that facilitates translocation of

the N-terminus domain to the cell nucleus when it is cleaved from the rest of the protein.<sup>26</sup> It has been suggested that this N-terminus domain, once within the nucleus, plays an important role in the loss of the keratinocyte nucleus during transformation of the cell into an anucleate corneocyte<sup>27</sup> and it has also been attributed a role in assembly of the CE.<sup>28</sup> The C-terminus domain is essential for correct processing of the profilaggrin, although its exact function is unknown.<sup>2</sup>

### Mutations of the Filaggrin Gene (*FLG*)

The first mutations identified in the *FLG* gene were R501X and 2282del4 in patients with ichthyosis vulgaris.<sup>29</sup> Many more mutations have been described since that time, but all are loss-of-function mutations (amino acid substitutions or frameshift mutations).<sup>21</sup> Furthermore, the mutations have been shown to display ethnic/geographic specificity, with different mutations being detected in the European and Asian populations.<sup>30</sup> There is also variation in the frequency of the mutations: 2 mutations (R501X and 2282del4) account for 80% of all *FLG* mutations in Ireland, whereas in Singapore no single mutation predominates in any such way over the others.<sup>21</sup> The overall prevalence of mutations in the general population in Europe is 7.7%, but in Asia is 3%.<sup>31</sup> Focusing on European studies, there is a noticeable difference in prevalence between the north and south of the continent. The majority of publications refer to northern populations, with a prevalence of approximately 10% (range, 7%-14%).<sup>31</sup> There are only 2 studies in Mediterranean populations, one in a French population<sup>32</sup> and the other in an Italian population<sup>33</sup>; both studies revealed a much lower prevalence (4%).

This apparently latitude-dependent trans-European gradient, led the Italian authors to put forward that perhaps *FLG* mutations offer some form of survival benefit in the north.<sup>33</sup> It has been suggested that this could be related to a lower level of UCA found in these patients; this would produce greater sensitivity to UV radiation.<sup>18,32,34</sup> Recent population-based studies have revealed that individuals with *FLG* mutations have serum vitamin D levels that are 10%

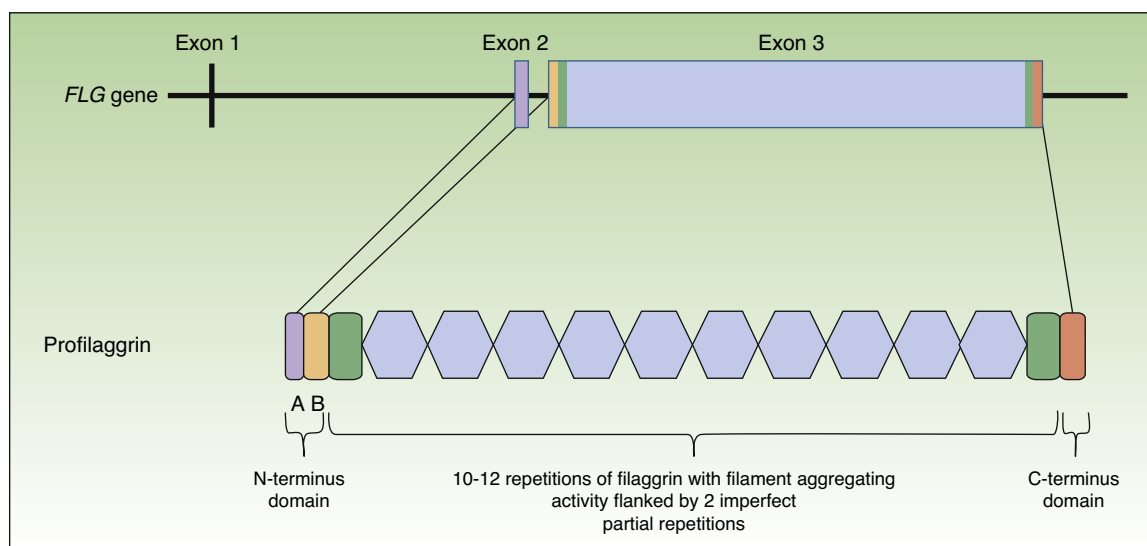


Figure 1 Structure of the *FLG* gene and of the profilaggrin protein.



**Figure 2** Fine, pale desquamation in a patient with ichthyosis vulgaris.

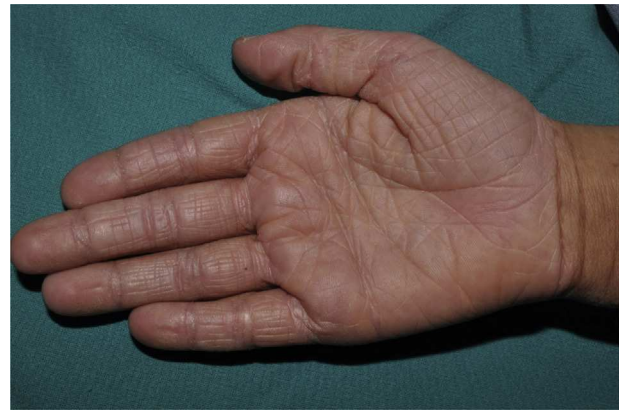
higher than controls.<sup>35</sup> Overall, these data show that in latitudes with a lower intensity of UV radiation, individuals with *FLG* mutations may have greater protection against the onset of diseases such as rickets.<sup>31</sup>

Table 2 lists the disorders that have been associated with a filaggrin deficit caused by mutations in the *FLG* gene; the main diseases are ichthyosis vulgaris and atopic disorders.

### Ichthyosis Vulgaris

Ichthyosis vulgaris is the most common disorder of keratinization, with an estimated prevalence between 1 in 80 and 1 in 250 in school-age English children,<sup>15</sup> and is the disease of mendelian inheritance caused by mutations in the *FLG* gene.<sup>31</sup> The absence (or marked reduction) of keratohyalin granules in biopsies from patients with ichthyosis vulgaris was first observed in the 1980s, together with a decrease in filaggrin expression detected using immunostaining techniques.<sup>36</sup> However, it was not until 2006 that loss-of-function mutations in the *FLG* gene were identified as the cause of the condition<sup>29</sup>; this lag was the result of the long length of the gene and its highly repetitive sequence, which made it difficult to sequence using conventional polymerase chain reaction techniques.<sup>15</sup> The discovery of these mutations made it possible to clarify the pattern of disease inheritance; this is autosomal semidominant, which explains the phenotypic variability of ichthyosis vulgaris. Heterozygotic patients present haploinsufficiency, which is a 50% reduction in filaggrin expression. In this situation, patients develop less serious disease (or may even be asymptomatic) and the manifestations respond better to external influences, such as the application of emollient creams or environmental moisture. Homozygotic individuals, on the other hand, develop all the manifestations of the disease.<sup>21,29</sup>

Clinically, ichthyosis vulgaris is characterized by the appearance of xerosis, keratosis pilaris, palmar hyperlinearity, and atopy in the postnatal period (typically in the first year of life).<sup>21</sup> Xerosis is seen as a fine, sometimes polygonal desquamation that mainly affects the extensor surface of the limbs (Fig. 2), scalp, central facial region, and trunk, while the folds are usually spared. The level of moisture



**Figure 3** Palmar hyperlinearity in a 7-year-old child with ichthyosis vulgaris.

modifies filaggrin processing,<sup>14,37</sup> which explains the tendency to sparing of the skin folds and why, in the majority of patients, the disease worsens in winter, when environmental humidity is lower.<sup>31</sup>

Painful fissures on the hands, heels, fingers, and toes are common (present in up to 76% of patients with ichthyosis vulgaris). This is closely related to the level of environmental moisture.<sup>31</sup> Furthermore, mutations in the *FLG* gene have been associated with the onset of fissured dermatitis on the dorsum of the hands and fingers in patients with AD and even in the general population.<sup>31,38</sup>

Palmar hyperlinearity (Fig. 3) and keratosis pilaris are typical findings not only in patients with clinical manifestations of ichthyosis vulgaris but also in all individuals with *FLG* mutations. One study found that palmar hyperlinearity had a positive predictive value of 71% and a negative predictive value of 90% for mutations in the *FLG* gene. That is, 71% of children with palmar hyperlinearity have a mutation, whereas mutations are highly unlikely in patients without palmar hyperlinearity. In the case of keratosis pilaris, the positive predictive value was 53% and the negative predictive value was 90%.<sup>39</sup>

Finally, 35% to 70% of patients with ichthyosis vulgaris develop atopic disease, particularly dermatitis but also allergic rhinitis and asthma.<sup>40</sup> This finding has led to extensive research into the relationship between mutations in the *FLG* gene and atopic diseases, detecting interesting data on the pathogenesis of these complex disorders.

### Atopic Diseases: Atopic March

Atopy is defined as a personal or familial tendency to sensitization and immunoglobulin (Ig) E antibody production in response to exposure to common environmental allergens to which everyone is exposed but to which the majority develop no response.<sup>41</sup> This tendency predisposes to the development of the so-called *atopic diseases*, which affect 20% of the population in developed countries.<sup>42</sup> The concept of *atopic march* was introduced to describe the tendency of AD to precede the sequential onset of asthma and allergic rhinitis,<sup>43</sup> which suggests that AD has an initiator role in the process. It has recently been suggested that food allergies may also form part of this atopic march.<sup>44</sup>

## Mutations in *FLG* and the Initiation of Atopic March

In 2006, the same year that the relationship between ichthyosis vulgaris and mutations in the *FLG* gene was described, a significant association was reported between the mutations and the onset of AD, the first step of atopic march.<sup>45</sup> Traditionally, AD has been considered an immune-mediated disorder associated with a secondary alteration of the skin barrier. However, particularly since the discovery of the role of filaggrin in many patients, the currently predominant thought is that the primary disorder is of the skin barrier.<sup>40</sup> According to this new hypothesis on the pathogenesis of AD, all patients with the disease have an inherent defect of the skin barrier, and such a defect has actually been demonstrated in both diseased and disease-free skin.<sup>46,47</sup> This defect can be caused by various different molecular mechanisms, one of the most important and frequent of which is filaggrin deficit secondary to mutations in the *FLG* gene.<sup>15</sup> The alteration of the skin barrier, combined with the immunological changes that it induces, will give rise to the clinical manifestations of the disease.<sup>21,40,42</sup>

## Influence of *FLG* Mutations on the Immune System

The alteration of the stratum corneum allows allergens to pass through and be taken up and processed by the Langerhans cells of the epidermis. The Langerhans cells migrate to the lymph nodes where they interact with T cells and induce a Th2-mediated immune response.<sup>40,42,48</sup> Percutaneous sensitization through a defective skin barrier has been demonstrated in mice with mutations in *FLG*<sup>49–51</sup> and it has also been shown that patients with AD and mutations in *FLG* have a significantly higher frequency of allergen-specific Th2 responses.<sup>52</sup> The continual entry of allergens will end up causing polarization of the adaptive immunity towards Th2,<sup>42</sup> characterized by the local production of Th2 cytokines (IL-4, IL-5, and IL-13), increased eosinophil and mast cell production, survival, and activation, and the production of allergen-specific IgE with an increase in total IgE.<sup>40</sup> In addition, this polarization towards Th2 is also favored by a series of cytokines released by the altered keratinocytes,<sup>40</sup> such as IL-1 and thymic stromal lymphopoietin (TSLP), as a result of the increased activity of endogenous proteases<sup>22</sup> and the action of exogenous proteases (including those released by sources of allergens [e.g., dust mites, roaches, fungi, and pollens] and those produced by *Staphylococcus aureus*, which frequently colonizes the skin of these patients).<sup>53</sup>

## Mutations in *FLG* and the Progression of Atopic March

Recent studies have demonstrated that carriers of *FLG* mutations have a higher risk of progression of atopic march,<sup>54</sup> and have shown a significantly higher risk of developing asthma,<sup>55</sup> allergic rhinitis,<sup>55</sup> and peanut allergy<sup>56</sup> compared with noncarriers.

Filaggrin is not expressed in the bronchial, nasal, or gastrointestinal mucosa and there will therefore be no alteration of barrier function at this level.<sup>40,57,58</sup> The mechanism by which *FLG* mutations promote allergic responses in these

mucosas is therefore through systemic sensitization to allergens that have penetrated through a defective skin barrier, rather than through the mucosa. This explains the tendency of AD to precede the other atopic disorders of atopic march.<sup>5</sup> A correct treatment of AD in children that restores and maintains the epidermal barrier would therefore prevent the subsequent onset of asthma, allergic rhinitis, or food allergy.<sup>42</sup> This supports the growing tendency to maintain a proactive (rather than reactive) therapeutic attitude towards AD, particularly in cases with moderate or severe manifestations.<sup>59,60</sup>

## Atopic Dermatitis

### *FLG* Mutations: Role in the Risk, Severity, Manifestations, and Epidemiology of Atopic Dermatitis

AD affects 20% of children and has a marked impact on their quality of life. It is the most prevalent chronic disease of childhood.<sup>61</sup> More than 30 studies have confirmed its association with mutations in *FLG*<sup>5</sup> and 2 recent meta-analyses have reported an odds-ratio (OR) for AD in association with *FLG* mutations of 3.12<sup>62</sup> and 4.78,<sup>63</sup> respectively. Carriers of *FLG* mutations therefore have a 4-times higher risk of developing AD than noncarriers. However, there appears to be not only a higher risk, but also greater severity.<sup>64</sup> Overall, "only" 15% to 20% of European patients with AD have mutations in *FLG*.<sup>10</sup> However, if European patients with AD are grouped according to clinical severity, it is found that the mutations are much more common in the group with moderate or severe AD (50%) than in those with mild AD (4%–15%).<sup>65</sup>

Based on the combined data from all the studies performed to date, the clinical profile described for AD associated with mutations in *FLG* (*AD<sub>FLG</sub>*) differs from the profile of those who do not have mutations (*AD<sub>non-FLG</sub>*).<sup>5</sup> *AD<sub>FLG</sub>* patients have more severe disease,<sup>64</sup> an earlier onset,<sup>66,67</sup> a greater tendency of the disease to persist into adulthood,<sup>68</sup> more marked alkalinization of the pH of the stratum corneum,<sup>69</sup> much lower levels of NMF,<sup>5</sup> greater IL-1 $\beta$  production in the stratum corneum,<sup>22</sup> palmar hyperlinearity,<sup>39</sup> more common fissured dermatitis on the dorsum of hands,<sup>38</sup> higher serum IgE levels,<sup>48</sup> greater allergic sensitization,<sup>48</sup> the development of multiple allergies,<sup>63</sup> a higher risk of asthma,<sup>55,70</sup> and a 10-fold risk of eczema herpeticum.<sup>71</sup>

In addition, this division also appears to be important from an epidemiological research point of view. The prevalence of AD is much higher in industrialized countries, leading to a suspicion of the influence of environmental factors.<sup>5</sup> Recent studies have shown that children who live with cats (but not with dogs) at early ages,<sup>72,73</sup> and those with an older sibling (which may imply greater exposure to pathogens and allergens)<sup>74</sup> have a significantly higher risk of developing AD. But this risk is only present in those children with mutations in *FLG*. Similarly, clinical severity correlates with the presence of specific IgE for dust mites or cat dander among patients with *AD<sub>FLG</sub>* but not among those with *AD<sub>non-FLG</sub>*.<sup>10</sup> It would thus appear important that future epidemiological studies of AD should stratify patients into carriers and noncarriers of mutations in *FLG*.<sup>5</sup>

## FLG Mutations and *S aureus* in Atopic Dermatitis

According to some studies, more than 90% of patients with AD are colonized by *S aureus* (vs 5% of healthy subjects)<sup>5,75</sup> and the clinical severity of the dermatitis correlates with the number of *S aureus* present on the skin.<sup>76</sup> This colonization is dependent on the integrity of the skin barrier and on the expression of bacterial adhesion proteins, which also contribute to the inflammatory state.<sup>5</sup> It has been shown in vitro that an acid pH of the stratum corneum inhibits the expression of these proteins on the bacterial surface and that the filaggrin degradation products, UCA and PCA, are themselves able to inhibit the expression of some of these proteins independently of the pH.<sup>77</sup> *FLG* mutations will thus cause greater *S aureus* colonization not only by altering the skin barrier but also by increasing the expression of adhesion proteins by the bacteria, through alkalization of the pH of the stratum corneum and reduced levels of UCA and PCA. In addition, the Th2 environment that exists in the lesions of these patients (and that is due in part to the deficit of filaggrin) inhibits the expression of antimicrobial peptides.<sup>78</sup>

Furthermore, *S aureus* secretes a wide variety of virulence factors that exacerbate the inflammation, worsening the skin lesions and interfering with their healing.<sup>77</sup> The most important of these virulence factors is  $\alpha$ -toxin, which induces cell death by creating pores in the plasma membrane that provoke cytolysis. A recent study has shown in vitro that keratinocytes with filaggrin deficit are more vulnerable to the action of this toxin than keratinocytes with normal filaggrin expression.<sup>79</sup> This is because cells that do not express filaggrin also have reduced sphingomyelinase expression, which leads to an increased presence of sphingomyelin, a lipid of the plasma membrane that serves as a receptor for  $\alpha$ -toxin.<sup>79</sup>

In summary, *FLG* mutations will not only facilitate colonization by *S aureus* but will also make keratinocytes more vulnerable to the action of *S aureus* toxins. On this subject, in 1 study it was observed that AD patients with an *FLG* mutation (compared with noncarriers) had a 7-fold risk of developing more than 4 episodes a year of skin infection that required antibiotic treatment.<sup>80</sup>

Therapeutically, these data support the need for measures that reduce colonization by *S aureus* and that acidify the skin pH to impede colonization.<sup>77</sup>

## Other Mechanisms of Filaggrin Deficit in Atopic Dermatitis

Although *FLG* mutations can explain many cases of AD (particularly the severe cases), a large proportion of these patients do not have any mutation.<sup>10</sup> However, it has been observed that the expression of filaggrin is also reduced in these patients without mutations, though to a lesser degree.<sup>10</sup> It would thus appear that additional mechanisms are involved in the deficit of filaggrin.

## Th2 Cytokines

Acute AD lesions have a predominantly Th2 inflammatory response to the entry of external antigens through the altered skin barrier.<sup>5,81</sup> It has been shown that in vitro

exposure of keratinocytes to Th2 cytokines produces a significant reduction in the expression of filaggrin.<sup>82</sup> Many patients with AD but without mutations in *FLG* may thus have an acquired filaggrin deficit because of the Th2 environment. This would in turn cause greater alteration of the stratum corneum, with increased entry of antigens that would finally lead to greater Th2 immune polarization, creating a vicious circle. To this situation it must be added that Th2 cytokines also cause a reduction in the expression of CE proteins such as loricrin and involucrin, further altering the structure of the stratum corneum.<sup>83</sup> Again, this would justify an active therapeutic approach to break these vicious circles.<sup>59</sup>

## Variation in the Number of Copies

As stated above, profilaggrin coded by the *FLG* gene contains between 10 and 12 filaggrin monomers.<sup>2</sup> This is called *copy number variation* and a recent study has demonstrated that this has clinical relevance.<sup>84</sup> In that study in Ireland it was observed that the most common variant of profilaggrin was the one containing 11 monomers (51.5%), followed by the one with 10 monomers (33.9%) and the one with 12 monomers (14.6%); those authors found that the variation in the number of copies gave rise to a dose-dependent decrease in the risk of AD. A subject whose 2 alleles coded for 10 monomers had a risk of developing AD 1.67 times higher than a patient whose 2 alleles coded for 12 monomers; the risk fell with each additional monomer.<sup>84</sup> Furthermore, a lower number of copies also correlated with a lower level of UCA in the stratum corneum.<sup>84</sup> Using the data obtained in that study, the authors estimated that an increase of 5% to 10% in filaggrin levels can have clinical repercussions. This would appear to be a good starting point for the development of new treatments.

## Epigenetic Regulatory Mechanisms

Epigenetic mechanisms alter the expression of a gene without changing its nucleotide sequence. At present, the only data on this subject relevant to filaggrin come from a study in which it was observed that, in individuals with haploinsufficiency, the higher the degree of methylation of the *FLG* gene, the higher the risk of developing AD.<sup>85</sup> It is believed that the increased risk of AD is due to a lower expression of the gene caused by the methylation, but that parameter was not evaluated.<sup>85</sup>

## Asthma, Allergic Rhinitis, and Food Allergy

Forty percent of children with AD will finally develop asthma or allergic rhinitis.<sup>10</sup> In the case of asthma, the risk correlates with the severity of the skin disease, with 70% of patients with severe AD developing asthma, while this only occurs in 20% to 30% of those with mild AD and 8% of the general population.<sup>42</sup> Population-based studies have demonstrated that carriers of *FLG* mutations have a higher risk of developing asthma, with an overall OR of 1.8.<sup>55,70</sup> But that risk is limited to those with AD, in whom the OR rises to 3.3 (95% confidence interval, 3.16-3.49).<sup>55,70</sup> Carriers of a mutation also have a higher risk of allergic rhinitis (OR, 2.64),

**Table 2** Disorders That Have Been Associated With Mutations in *FLG*.

<b>Cutaneous</b>
<i>Ichthyosis vulgaris</i> <sup>29</sup>
<i>Atopic dermatitis</i> <sup>45</sup> and its complications:
<i>Recurrent Staphylococcus aureus infection</i> <sup>80</sup>
<i>Eczema herpeticum</i> <sup>71</sup>
<i>Irritant contact dermatitis</i> <sup>86,87</sup>
<i>Nickel sensitization and earlier onset allergic contact dermatitis to nickel</i> <sup>88,89</sup>
<i>Chronic hand eczema</i> <sup>90</sup>
<i>Alopecia areata: more aggressive course in patients with atopic dermatitis</i> <sup>91</sup>
<i>More severe phenotype of recessive X-linked ichthyosis</i> <sup>92</sup>
<i>More severe phenotype of pachyonychia congenita</i> <sup>93</sup>
<b>Extracutaneous</b>
<i>Asthma</i> <sup>55,70</sup>
<i>Allergic rhinitis</i> <sup>55</sup>
<i>Peanut allergy</i> <sup>56</sup>
<i>Type 2 diabetes mellitus</i> <sup>94</sup>

Source: Adapted from Thyssen et al.<sup>51</sup>

although in this case it is independent of whether or not the patient has eczema.<sup>55</sup> Finally, concerning the onset of food allergy, it has been shown that carriers of *FLG* mutations have a significantly higher risk of developing peanut allergy; this risk is greater if the patient has AD, but is also present independently of AD (overall OR, 5.3, with a residual OR of 3.8 after adjustment for AD).<sup>56</sup>

### Role of *FLG* Mutations in Other Disorders

Since the discovery of mutations in the *FLG* gene, numerous studies have been performed to look for a possible relationship with various skin and extracutaneous diseases, whether due to an association with AD, to a similar underlying pathogenesis, or to a shared susceptibility locus.<sup>15</sup> A summary of the results of these studies is presented in Tables 2 and 3. Given the high frequency of carriers in the general population, it is likely that *FLG* mutations can act as a modifying factor in numerous disorders, particularly in those related to abnormalities of keratinization and of the skin barrier.<sup>21</sup>

**Table 3** Disorders That Do NOT Appear to Be Related to Mutations in *FLG*.

<b>Cutaneous</b>
<i>Acne vulgaris</i> <sup>95</sup>
<i>Psoriasis vulgaris</i> <sup>96,97</sup>
<b>Extracutaneous</b>
<i>Rheumatoid arthritis and psoriatic arthritis</i> <sup>98</sup>
<i>Inflammatory bowel disease</i> <sup>99,100</sup>
<i>Sarcoidosis</i> <sup>100</sup>
<i>Hearing disturbances</i> <sup>101</sup>

Source: Adapted from Thyssen et al.<sup>51</sup>

## Conclusions

Filaggrin is an essential protein for the correct formation and function of the skin barrier. Mutations in the *FLG* gene are responsible for ichthyosis vulgaris and are associated with a higher risk of developing AD, asthma, allergic rhinitis, and food allergy. The discovery of its function has enabled us to better understand the pathogenesis of various disorders associated with alterations of the skin barrier, and it is likely that its mutations influence the onset or clinical severity of other dermatologic diseases. In addition, the study of its functions and of the consequences of its deficit may have important therapeutic implications in the future, with the possibility of developing new specific treatments for disorders in which this gene is altered.

## Conflicts of Interest

The authors declare that they have no conflicts of interest.

## References

- Madison KC. Barrier function of the skin: "la raison d'être" of the epidermis. *J Invest Dermatol.* 2003;121:231–41.
- Sandilands A, Sutherland C, Irvine AD, McLean WH. Filaggrin in the frontline: Role in skin barrier function and disease. *J Cell Sci.* 2009;122:1285–94.
- Dale BA. Purification and characterization of a basic protein from the stratum corneum of mammalian epidermis. *Biochim Biophys Acta.* 1977;491:193–204.
- Steinert PM, Cantieri JS, Teller DC, Lonsdale-Eccles JD, Dale BA. Characterization of a class of cationic proteins that specifically interact with intermediate filaments. *Proc Natl Acad Sci USA.* 1981;78:4097–101.
- McAleer MA, Irvine AD. The multifunctional role of filaggrin in allergic skin disease. *J Allergy Clin Immunol.* 2013;131:280–91.
- Proksch E, Brandner JM, Jensen J-M. The skin: An indispensable barrier. *Exp Dermatol.* 2008;17:1063–72.
- Nemes Z, Steinert PM. Bricks and mortar of the epidermal barrier. *Exp Mol Med.* 1999;31:5–19.
- Menon GK, Grayson S, Elias PM. Ionic calcium reservoirs in mammalian epidermis: Ultrastructural localization by ion-capture cytochemistry. *J Invest Dermatol.* 1985;84:508–12.
- Lee SH, Elias PM, Proksch E, Menon GK, Mao-Quiang M, Feingold KR. Calcium and potassium are important regulators of barrier homeostasis in murine epidermis. *J Clin Invest.* 1992;89:530–8.
- Heimall J, Spergel JM. Filaggrin mutations and atopy: Consequences for future therapeutics. *Expert Rev Clin Immunol.* 2012;8:189–97.
- Kalinin A, Marekov LN, Steinert PM. Assembly of the epidermal cornified cell envelope. *J Cell Sci.* 2001;114:3069–70.
- Kalinin AE, Kajava AV, Steinert PM. Epithelial barrier function: Assembly and structural features of the cornified cell envelope. *Bioessays.* 2002;24:789–800.
- Candi E, Schmidt R, Melino G. The cornified envelope: A model of cell death in the skin. *Nat Rev Mol Cell Biol.* 2005;6:328–40.
- Rawlings AV, Harding CR. Moisturization and skin barrier function. *Dermatol Ther.* 2004;17 Suppl 1:43–8.
- Brown SJ, McLean WH. One remarkable molecule: Filaggrin. *J Invest Dermatol.* 2012;132:751–62.
- Gruber R, Elias PM, Crumrine D, Lin T-K, Brandner JM, Hachem J-P, et al. Filaggrin genotype in ichthyosis vulgaris predicts abnormalities in epidermal structure and function. *Am J Pathol.* 2011;178:2252–63.

17. Ali SM, Yosipovitch G. Skin pH: From basic science to basic skin care. *Acta Derm Venereol.* 2013;93:261–7.
18. Barresi C, Stremnitzer C, Mlitz V, Kezic S, Kammeyer A, Ghanadan M, et al. Increased sensitivity of histidinemic mice to UVB radiation suggests a crucial role of endogenous urocanic acid in photoprotection. *J Invest Dermatol.* 2011;131:188–94.
19. Noonan FP, de Fabo EC. Immunosuppression by ultraviolet B radiation: Initiation by urocanic acid. *Immunol Today.* 1992;13:250–4.
20. Walterscheid JP, Nghiem DX, Kazimi N, Nutt LK, McConkey DJ, Norval M, et al. Cis-urocanic acid, a sunlight-induced immunosuppressive factor, activates immune suppression via the 5-HT2A receptor. *Proc Natl Acad Sci USA.* 2006;103:1742–5.
21. Irvine AD, McLean WHI, Leung DYM. Filaggrin mutations associated with skin and allergic diseases. *N Engl J Med.* 2011;365:1315–27.
22. Kezic S, O'Regan GM, Lutter R, Jakasa I, Koster ES, Saunders S, et al. Filaggrin loss-of-function mutations are associated with enhanced expression of IL-1 cytokines in the stratum corneum of patients with atopic dermatitis and in a murine model of filaggrin deficiency. *J Allergy Clin Immunol.* 2012;129:1031–9.
23. Mischke D, Korge BP, Marenholz I, Volz A, Ziegler A. Genes encoding structural proteins of epidermal cornification and S100 calcium-binding proteins form a gene complex ("epidermal differentiation complex") on human chromosome 1q21. *J Invest Dermatol.* 1996;106:989–92.
24. Presland RB, Haydock PV, Fleckman P, Nirunskisiri W, Dale BA. Characterization of the human epidermal profilaggrin gene. Genomic organization and identification of an S-100-like calcium binding domain at the amino terminus. *J Biol Chem.* 1992;267:23772–81.
25. Presland RB, Bassuk JA, Kimball JR, Dale BA. Characterization of two distinct calcium-binding sites in the amino-terminus of human profilaggrin. *J Invest Dermatol.* 1995;104:218–23.
26. Aho S, Harding CR, Lee JM, Meldrum H, Bosko CA. Regulatory role for the profilaggrin N-terminal domain in epidermal homeostasis. *J Invest Dermatol.* 2012;132:2376–85.
27. Ishida-Yamamoto A, Takahashi H, Presland RB, Dale BA, Iizuka H. Translocation of profilaggrin N-terminal domain into keratinocyte nuclei with fragmented DNA in normal human skin and loricrin keratoderma. *Lab Invest.* 1998;78:1245–53.
28. Yoneda K, Nakagawa T, Lawrence OT, Huard J, Demitsu T, Kubota Y, et al. Interaction of the profilaggrin N-terminal domain with loricrin in human cultured keratinocytes and epidermis. *J Invest Dermatol.* 2012;132:1206–14.
29. Smith FJD, Irvine AD, Terron-Kwiatkowski A, Sandilands A, Campbell LE, Zhao Y, et al. Loss-of-function mutations in the gene encoding filaggrin cause ichthyosis vulgaris. *Nat Genet.* 2006;38:337–42.
30. Chen H, Common JE, Haines RL, Balakrishnan A, Brown SJ, Goh CS, et al. Wide spectrum of filaggrin-null mutations in atopic dermatitis highlights differences between Singaporean Chinese and European populations. *Br J Dermatol.* 2011;165:106–14.
31. Thyssen JP, Godoy-Gijon E, Elias PM. Ichthyosis vulgaris - the filaggrin mutation disease. *Br J Dermatol.* 2013.
32. Mlitz V, Latreille J, Gardinier S, Jdid R, Drouault Y, Hufnagl P, et al. Impact of filaggrin mutations on Raman spectra and biophysical properties of the stratum corneum in mild to moderate atopic dermatitis. *J Eur Acad Dermatol Venereol.* 2012;26:983–90.
33. Cascella R, Foti Cuzzola V, Lepre T, Galli E, Moschese V, Chini L, et al. Full sequencing of the FLG gene in Italian patients with atopic eczema: Evidence of new mutations, but lack of an association. *J Invest Dermatol.* 2011;131:982–4.
34. Mildner M, Jin J, Eckhart L, Kezic S, Gruber F, Barresi C, et al. Knockdown of filaggrin impairs diffusion barrier function and increases UV sensitivity in a human skin model. *J Invest Dermatol.* 2010;130:2286–94.
35. Thyssen JP, Thuesen B, Huth C, Standl M, Carson CG, Heinrich J, et al. Skin barrier abnormality caused by filaggrin (FLG) mutations is associated with increased serum 25-hydroxyvitamin D concentrations. *J Allergy Clin Immunol.* 2012;130:1204–7.
36. Sybert VP, Dale BA, Holbrook KA. Ichthyosis vulgaris: Identification of a defect in synthesis of filaggrin correlated with an absence of keratohyaline granules. *J Invest Dermatol.* 1985;84:191–4.
37. Katagiri C, Sato J, Nomura J, Denda M. Changes in environmental humidity affect the water-holding property of the stratum corneum and its free amino acid content, and the expression of filaggrin in the epidermis of hairless mice. *J Dermatol Sci.* 2003;31:29–35.
38. Thyssen JP, Ross-Hansen K, Johansen JD, Zachariae C, Carlsen BC, Linneberg A, et al. Filaggrin loss-of-function mutation R501X and 22824 carrier status is associated with fissured skin on the hands: Results from a cross-sectional population study. *Br J Dermatol.* 2012;166:46–53.
39. Brown SJ, Relton CL, Liao H, Zhao Y, Sandilands A, Wilson IJ, et al. Filaggrin null mutations and childhood atopic eczema: A population-based case-control study. *J Allergy Clin Immunol.* 2008;121:940–6.
40. De Benedetto A, Kubo A, Beck LA. Skin barrier disruption: A requirement for allergen sensitization? *J Invest Dermatol.* 2012;132:949–63.
41. Johansson SGO, Bieber T, Dahl R, Friedmann PS, Lanier BQ, Lockey RF, et al. Revised nomenclature for allergy for global use: Report of the Nomenclature Review Committee of the World Allergy Organization, October 2003. *J Allergy Clin Immunol.* 2004;113:832–6.
42. Zheng T, Yu J, Oh MH, Zhu Z. The atopic march: Progression from atopic dermatitis to allergic rhinitis and asthma. *Allergy Asthma Immunol Res.* 2011;3:67–73.
43. Spergel JM, Paller AS. Atopic dermatitis and the atopic march. *J Allergy Clin Immunol.* 2003;112:S118–27.
44. Allen KJ, Dharmage SC. The role of food allergy in the atopic march. *Clin Exp Allergy.* 2010;40:1439–41.
45. Palmer CNA, Irvine AD, Terron-Kwiatkowski A, Zhao Y, Liao H, Lee SP, et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. *Nat Genet.* 2006;38:441–6.
46. Werner Y, Lindberg M. Transepidermal water loss in dry and clinically normal skin in patients with atopic dermatitis. *Acta Derm Venereol.* 1985;65:102–5.
47. Seidenari S, Giusti G. Objective assessment of the skin of children affected by atopic dermatitis: A study of pH, capacitance and TEWL in eczematous and clinically uninvolved skin. *Acta Derm Venereol.* 1995;75:429–33.
48. Weidinger S, Rodríguez E, Stahl C, Wagenpfeil S, Klopp N, Illig T, et al. Filaggrin mutations strongly predispose to early-onset and extrinsic atopic dermatitis. *J Invest Dermatol.* 2007;127:724–6.
49. Fallon PG, Sasaki T, Sandilands A, Campbell LE, Saunders SP, Mangan NE, et al. A homozygous frameshift mutation in the mouse FLG gene facilitates enhanced percutaneous allergen priming. *Nat Genet.* 2009;41:602–8.
50. Oyoshi MK, Murphy GF, Geha RS. Filaggrin-deficient mice exhibit TH17-dominated skin inflammation and permissiveness to epicutaneous sensitization with protein antigen. *J Allergy Clin Immunol.* 2009;124:485–93.
51. Kawasaki H, Nagao K, Kubo A, Hata T, Shimizu A, Mizuno H, et al. Altered stratum corneum barrier and enhanced percutaneous immune responses in filaggrin-null mice. *J Allergy Clin Immunol.* 2012;129:1538–46.



52. McPherson T, Sherman VJ, Aslam A, Crack L, Chan H, Lloyd-Lavery A, et al. Filaggrin null mutations associate with increased frequencies of allergen-specific CD4+ T-helper 2 cells in patients with atopic eczema. *Br J Dermatol*. 2010;163:544–9.
53. Takai T, Ikeda S. Barrier dysfunction caused by environmental proteases in the pathogenesis of allergic diseases. *Allergol Int*. 2011;60:25–35.
54. Marenholz I, Nickel R, Rüschemdorf F, Schulz F, Esparza-Gordillo J, Kerscher T, et al. Filaggrin loss-of-function mutations predispose to phenotypes involved in the atopic march. *J Allergy Clin Immunol*. 2006;118:866–71.
55. Weidinger S, O'Sullivan M, Illig T, Baurecht H, Depner M, Rodriguez E, et al. Filaggrin mutations, atopic eczema, hay fever, and asthma in children. *J Allergy Clin Immunol*. 2008;121:1203–9.
56. Brown SJ, Asai Y, Cordell HJ, Campbell LE, Zhao Y, Liao H, et al. Loss-of-function variants in the filaggrin gene are a significant risk factor for peanut allergy. *J Allergy Clin Immunol*. 2011;127:661–7.
57. Ying S, Meng Q, Corrigan CJ, Lee TH. Lack of filaggrin expression in the human bronchial mucosa. *J Allergy Clin Immunol*. 2006;118:1386–8.
58. De Benedetto A, Qualia CM, Baroody FM, Beck LA. Filaggrin expression in oral, nasal, and esophageal mucosa. *J Invest Dermatol*. 2008;128:1594–7.
59. Wollenberg A, Bieber T. Proactive therapy of atopic dermatitis—an emerging concept. *Allergy*. 2009;64:276–8.
60. Wollenberg A, Ehmann LM. Long term treatment concepts and proactive therapy for atopic eczema. *Ann Dermatol*. 2012;24:253–60.
61. Garnacho-Saucedo G, Salido-Vallejo R, Moreno-Giménez JC. Atopic dermatitis: Update and proposed management algorithm. *Actas Dermosifiliogr*. 2013;104:4–16.
62. Rodríguez E, Baurecht H, Herberich E, Wagenpfeil S, Brown SJ, Cordell HJ, et al. Meta-analysis of filaggrin polymorphisms in eczema and asthma: Robust risk factors in atopic disease. *J Allergy Clin Immunol*. 2009;123:1361–70.
63. Van den Oord RA, Sheikh A. Filaggrin gene defects and risk of developing allergic sensitisation and allergic disorders: Systematic review and meta-analysis. *BMJ*. 2009;339:b2433.
64. Brown SJ, Relton CL, Liao H, Zhao Y, Sandilands A, McLean WHI, et al. Filaggrin haploinsufficiency is highly penetrant and is associated with increased severity of: Further delineation of the skin phenotype in a prospective epidemiological study of 792 school children. *Br J Dermatol*. 2009;161:884–9.
65. Brown SJ, McLean WHI. Eczema genetics: Current state of knowledge and future goals. *J Invest Dermatol*. 2009;129:543–52.
66. Brown SJ, Sandilands A, Zhao Y, Liao H, Relton CL, Meggitt SJ, et al. Prevalent and low-frequency null mutations in the filaggrin gene are associated with early-onset and persistent atopic eczema. *J Invest Dermatol*. 2008;128:1591–4.
67. Stemmler S, Parwez Q, Petrasch-Parwez E, Epplen JT, Hoffjan S. Two common loss-of-function mutations within the filaggrin gene predispose for early onset of atopic dermatitis. *J Invest Dermatol*. 2007;127:722–4.
68. Margolis DJ, Apter AJ, Gupta J, Hoffstad O, Papadopoulos M, Campbell LE, et al. The persistence of atopic dermatitis and filaggrin (FLG) mutations in a US longitudinal cohort. *J Allergy Clin Immunol*. 2012;130:912–7.
69. Jungersted JM, Scheer H, Mempel M, Baurecht H, Cifuentes L, Høgh JK, et al. Stratum corneum lipids, skin barrier function and filaggrin mutations in patients with atopic eczema. *Allergy*. 2010;65:911–8.
70. Henderson J, Northstone K, Lee SP, Liao H, Zhao Y, Pembrey M, et al. The burden of disease associated with filaggrin mutations: A population-based, longitudinal birth cohort study. *J Allergy Clin Immunol*. 2008;121:872–7.
71. Gao PS, Rafaels NM, Hand T, Murray T, Boguniewicz M, Hata T, et al. Filaggrin mutations that confer risk of atopic dermatitis confer greater risk for eczema herpeticum. *J Allergy Clin Immunol*. 2009;124:507–13.
72. Bisgaard H, Simpson A, Palmer CNA, Bønnelykke K, McLean I, Mukhopadhyay S, et al. Gene-environment interaction in the onset of eczema in infancy: Filaggrin loss-of-function mutations enhanced by neonatal cat exposure. *PLoS Med*. 2008;5:e131.
73. Schuttelaar MLA, Kerkhof M, Jonkman MF, Koppelman GH, Brunekreef B, de Jongste JC, et al. Filaggrin mutations in the onset of eczema, sensitization, asthma, hay fever and the interaction with cat exposure. *Allergy*. 2009;64:1758–65.
74. Cramer C, Link E, Horster M, Koletzko S, Bauer C-P, Berdel D, et al. Elder siblings enhance the effect of filaggrin mutations on childhood eczema: Results from the 2 birth cohort studies LIS-Aplus and GINIplus. *J Allergy Clin Immunol*. 2010;125:1254–60.
75. Bieber T. Atopic dermatitis. *N Engl J Med*. 2008;358:1483–94.
76. Williams RE, Gibson AG, Aitchison TC, Lever R, Mackie RM. Assessment of a contact-plate sampling technique and subsequent quantitative bacterial studies in atopic dermatitis. *Br J Dermatol*. 1990;123:493–501.
77. Miajlovic H, Fallon PG, Irvine AD, Foster TJ. Effect of filaggrin breakdown products on growth of and protein expression by *Staphylococcus aureus*. *J Allergy Clin Immunol*. 2010;126:1184–90.
78. Kisich KO, Carspecken CW, Fiéve S, Boguniewicz M, Leung DY. Defective killing of *Staphylococcus aureus* in atopic dermatitis is associated with reduced mobilization of human beta-defensin-3. *J Allergy Clin Immunol*. 2008;122:62–8.
79. Brauweiler AM, Bin L, Kim BE, Oyoshi MK, Geha RS, Goleva E, et al. Filaggrin-dependent secretion of sphingomyelinase protects against staphylococcal  $\alpha$ -toxin-induced keratinocyte death. *J Allergy Clin Immunol*. 2013;131:421–7.
80. Cai SC, Chen H, Koh WP, Common JE, van Bever HP, McLean WHI, et al. Filaggrin mutations are associated with recurrent skin infection in Singaporean Chinese patients with atopic dermatitis. *Br J Dermatol*. 2012;166:200–3.
81. Oyoshi MK, He R, Kumar L, Yoon J, Geha RS. Cellular and molecular mechanisms in atopic dermatitis. *Adv Immunol*. 2009;102:135–226.
82. Howell MD, Kim BE, Gao P, Grant AV, Boguniewicz M, de Benedetto A, et al. Cytokine modulation of atopic dermatitis filaggrin skin expression. *J Allergy Clin Immunol*. 2009;124:R7–12.
83. Kim BE, Leung DYM, Boguniewicz M, Howell MD. Loricrin and involucrin expression is down-regulated by Th2 cytokines through STAT-6. *Clin Immunol*. 2008;126:332–7.
84. Brown SJ, Kroboth K, Sandilands A, Campbell LE, Pohler E, Kezic S, et al. Intragenic copy number variation within filaggrin contributes to the risk of atopic dermatitis with a dose-dependent effect. *J Invest Dermatol*. 2012;132:98–104.
85. Ziyab AH, Karmaus W, Holloway JW, Zhang H, Ewart S, Arshad SH. DNA methylation of the filaggrin gene adds to the risk of eczema associated with loss-of-function variants. *J Eur Acad Dermatol Venereol*. 2012;27:e420–3.
86. De Jongh CM, Khrenova L, Verberk MM, Calkoen F, van Dijk FJH, Voss H, et al. Loss-of-function polymorphisms in the filaggrin gene are associated with an increased susceptibility to chronic irritant contact dermatitis: A case-control study. *Br J Dermatol*. 2008;159:621–7.
87. Visser MJ, Landeck L, Campbell LE, McLean WHI, Weidinger S, Calkoen F, et al. Impact of atopic dermatitis and loss-of-function mutations in the filaggrin gene on the development of occupational irritant contact dermatitis. *Br J Dermatol*. 2013;168:326–32.

88. Thyssen JP, Johansen JD, Linneberg A, Menné T, Nielsen NH, Meldgaard M, et al. The association between null mutations in the filaggrin gene and contact sensitization to nickel and other chemicals in the general population. *Br J Dermatol*. 2010;162:1278–85.
89. Ross-Hansen K, Menné T, Johansen JD, Carlsen BC, Linneberg A, Nielsen NH, et al. Nickel reactivity and filaggrin null mutations-evaluation of the filaggrin bypass theory in a general population. *Contact Derm*. 2011;64:24–31.
90. Molin S, Vollmer S, Weiss EH, Ruzicka T, Prinz JC. Filaggrin mutations may confer susceptibility to chronic hand eczema characterized by combined allergic and irritant contact dermatitis. *Br J Dermatol*. 2009;161:801–7.
91. Betz RC, Pforr J, Flaquer A, Redler S, Hanneken S, Eigelshoven S, et al. Loss-of-function mutations in the filaggrin gene and alopecia areata: Strong risk factor for a severe course of disease in patients comorbid for atopic disease. *J Invest Dermatol*. 2007;127:2539–43.
92. Liao H, Waters AJ, Goudie DR, Aitken DA, Graham G, Smith FJD, et al. Filaggrin mutations are genetic modifying factors exacerbating X-linked ichthyosis. *J Invest Dermatol*. 2007;127:2795–8.
93. Gruber R, Wilson NJ, Smith FJD, Grabher D, Steinwender L, Fritsch PO, et al. Increased pachyonychia congenita severity in patients with concurrent keratin and filaggrin mutations. *Br J Dermatol*. 2009;161:1391–5.
94. Thyssen JP, Linneberg A, Carlsen BC, Johansen JD, Engkilde K, Hansen T, et al. A possible association between a dysfunctional skin barrier (filaggrin null-mutation status) and diabetes: A cross-sectional study. *BMJ Open*. 2011;1:e000062.
95. Common JEA, Brown SJ, Haines RL, Goh CS, Chen H, Balakrishnan A, et al. Filaggrin null mutations are not a protective factor for acne vulgaris. *J Invest Dermatol*. 2011;131:1378–80.
96. Zhao Y, Terron-Kwiatkowski A, Liao H, Lee SP, Allen MH, Hull PR, et al. Filaggrin null alleles are not associated with psoriasis. *J Invest Dermatol*. 2007;127:1878–82.
97. Thyssen JP, Johansen JD, Carlsen BC, Linneberg A, Meldgaard M, Szecsi PB, et al. The filaggrin null genotypes R501X and 22824 seem not to be associated with psoriasis: Results from general population study and meta-analysis. *J Eur Acad Dermatol Venereol*. 2012;26:782–4.
98. Hüffmeier U, Böiers U, Lascorz J, Reis A, Burkhardt H. Loss-of-function mutations in the filaggrin gene: No contribution to disease susceptibility, but to autoantibody formation against citrullinated peptides in early rheumatoid arthritis. *Ann Rheum Dis*. 2008;67:131–3.
99. Van Limbergen J, Russell RK, Nimmo ER, Zhao Y, Liao H, Drummond HE, et al. Filaggrin loss-of-function variants are associated with atopic comorbidity in pediatric inflammatory bowel disease. *Inflamm Bowel Dis*. 2009;15:1492–8.
100. Ruether A, Stoll M, Schwarz T, Schreiber S, Fölster-Holst R. Filaggrin loss-of-function variant contributes to atopic dermatitis risk in the population of Northern Germany. *Br J Dermatol*. 2006;155:1093–4.
101. Rodriguez S, Hall AJ, Granell R, McLean WH, Irvine AD, Palmer CN, et al. Carrier status for the common R501X and 22824 filaggrin mutations is not associated with hearing phenotypes in 5,377 children from the ALSPAC cohort. *PLoS ONE*. 2009;4:e5784.