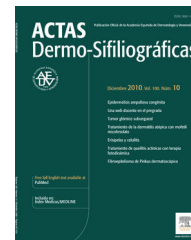


ACTAS Derma-Sifiliográficas

Full English text available at
www.elsevier.es/ad



REVIEW

Ectodermal Dysplasias: A Clinical and Molecular Review[☆]

P. García-Martín, A. Hernández-Martín,* A. Torrelo

Servicio de Dermatología, Hospital Infantil del Niño Jesús, Madrid, Spain

Received 19 December 2011; accepted 20 July 2012

Available online 14 June 2013

KEYWORDS

Genodermatosis;
Ectodermal dysplasia;
Nuclear Factor kappa
B;
Ectodysplasin;
Protein p63

PALABRAS CLAVE

Genodermatosis;
Displasia
ectodérmica;
Factor Nuclear kappa
B;
Ectodisplasia;
Proteína p63

Abstract The ectodermal dysplasias are a large group of hereditary disorders characterized by alterations of structures of ectodermal origin. Although some syndromes can have specific features, many of them share common clinical characteristics. Two main groups of ectodermal dysplasias can be distinguished. One group is characterized by aplasia or hypoplasia of ectodermal tissues, which fail to develop and differentiate because of a lack of reciprocal signaling between ectoderm and mesoderm, the other has palmoplantar keratoderma as its most striking feature, with additional manifestations when other highly specialized epithelia are also involved. In recent decades, the genes responsible for at least 30 different types of ectodermal dysplasia have been identified, throwing light on the pathogenic mechanisms involved and their correlation with clinical findings.

© 2011 Elsevier España, S.L. and AEDV. All rights reserved.

Displasias ectodérmicas: revisión clínica y molecular

Resumen Las displasias ectodérmicas son un amplio grupo de trastornos hereditarios que se caracterizan por la alteración de estructuras derivadas del ectodermo. Aunque algunos de estos síndromes poseen características específicas, determinados rasgos clínicos son comunes en muchos de ellos. De modo general, se diferencian 2 grupos de trastornos: uno caracterizado por la aplasia o hipoplasia de los derivados ectodérmicos, que fracasan en su desarrollo y diferenciación por la ausencia de señales recíprocas específicas entre ectodermo y mesénquima, y otro en el que la característica más llamativa es la queratodermia palmoplantar, que se presenta en asociación con otras manifestaciones cuando se afectan otros epitelios altamente especializados. En las últimas décadas se ha logrado identificar el gen responsable en al menos 30 entidades, permitiéndonos entender los mecanismos patogénicos y su correlación con la clínica.

© 2011 Elsevier España, S.L. y AEDV. Todos los derechos reservados.

[☆] Please cite this article as: García-Martín P, et al. Displasias ectodérmicas: revisión clínica y molecular. Actas Dermosifiliogr. 2013;104:451–70.

* Corresponding Author.

E-mail address: ahernandez_hnj@yahoo.es (A. Hernández-Martín).

Introduction

The ectoderm is one of the primitive embryonic components. At around the third week of development, it undergoes a subdivision into the neuroectoderm, the origin of the nervous system, and the ectoderm, which will envelop the entire embryonic surface and form the epidermis, epidermal appendages, and tooth enamel. The ectoderm therefore gives rise not only to hair, teeth, nails, and sweat glands, but also to the central nervous system, peripheral nervous system, eyes, ears, and nose, as well as the eccrine, mammary, and pituitary glands.¹ During development, the ectoderm undergoes complex interactions with the mesoderm, so ectodermal disorders may lead to abnormalities in mesodermal structures such as the musculoskeletal and genitourinary systems.²

Ectodermal dysplasias (EDs) are a heterogeneous group of hereditary disorders characterized by certain shared structural and functional abnormalities in tissues derived from the ectoderm. Most of these diseases are also associated with an abnormal development of structures derived from the mesoderm and, occasionally, mental retardation. They are considered rare conditions, with an estimated incidence of 7 cases per 10 000 births.³ They can be transmitted by any of the possible Mendelian inheritance patterns,⁴ and although many share certain clinical characteristics, some syndromes have specific clinical findings. To date, approximately 200 such conditions are known, and the causative gene mutation has been identified in around 30. Mutations in only 4 genes (*EDA1*, *EDAR*, *EDARADD*, and *WNT10A*) are responsible for most cases of ED.⁵

Historical Perspective

The first descriptions of clinical cases that might correspond to what we would now classify as ED date from 1792.⁶ In 1848, Thurman defined anhidrotic ectodermal dysplasia (also known as hypohidrotic ectodermal dysplasia [HED]) as a condition in its own right.⁷ Subsequently, similar cases were reported, such as the one presented by Wedderhorn and published in 1875 by the naturalist Charles Darwin: "I may give an analogous case, communicated to me by Mr. W. Weddenburn, of a Hindoo family in Scinde, in which ten men, in the course of four generations, were furnished, in both jaws taken together, with only four small and weak incisor teeth and with eight posterior molars. The men thus affected have very little hair on the body, and become bald early in life. They also suffer much during hot weather from excessive dryness of the skin. It is remarkable that no instance has occurred of a daughter being affected. . . though the daughters in the above family are never affected, they transmit the tendency to their sons: and no case has occurred of a son transmitting it to his sons. The affection thus appears only in alternate generations, or after long intervals."⁸ The above case described by Darwin corresponds to what we would describe today as X-linked HED, a term coined by Weech in 1929.⁹

Classification of Ectodermal Dysplasias

The classification of EDs is complex, and classification systems have come and gone in an attempt to accommodate clinical and genetic data.^{10–15} Biomolecular findings have enabled the identification of the causative mutations that become manifest through 2 broad pathogenic mechanisms, associated with specific clinical features. Using these mechanisms as a starting point, in 2009, Priolo² established a clinical-functional classification, which will form the basis for this review. The author proposed the definition of 2 groups of disorders (Fig. 1).

Group 1

Group 1 corresponds to disorders in which defective interaction between the ectoderm and the mesenchyme is apparent. Two pathophysiologic mechanisms have been identified:

1. Changes in the signaling pathways that modulate activity of nuclear factor (NF) κ B (ectodysplasin/ectodysplasin-A [EDA] receptor [EDAR]/EDAR associated death domain [EDARADD] signaling pathway and NEMO [NF- κ B essential modulator] regulatory pathway).
2. Regulatory changes in transcription and/or expression of genes such as *p63*, *DLX3*, *MSX1*, *EVC2*, and *EVC*.

The resulting clinical phenotype is hypoplasia or aplasia of structures derived from the ectoderm. Development and differentiation of these structures fail due to the absence of specific reciprocal signals between the ectoderm and the mesenchyme (Table 1).

Group 2

Group 2 corresponds to disorders in which there is abnormal function of a structural protein in the cell membrane. Examples of structural proteins include nectin 1, connexins, and plakophilin, whose role in adhesion and cell-cell communication is essential for maintaining tissue homeostasis and controlling cell growth, development, and response to different stimuli.

Clinically, these disorders are mainly characterized by skin abnormalities such as palmoplantar keratoderma, with or without involvement of highly differentiated epithelia associated with deafness or retinal dystrophy (Table 2).

Group 1

Changes in the Signaling Pathways That Modulate NF- κ B Activity

Ectodysplasin/EDAR/EDARADD Pathway

EDA, which consists of 391 amino acids, is a type 2 transmembrane protein belonging to the tumor necrosis factor (TNF) family.¹⁶ The most biologically important isoforms of EDA are EDA-A1 and EDA-A2.¹⁷ EDA-A1 binds to the EDA receptor (EDAR), whereas EDA-A2 binds to the X-linked EDAR A2 ligand (Fig. 2).¹⁸ EDAR is a type 1 transmembrane protein

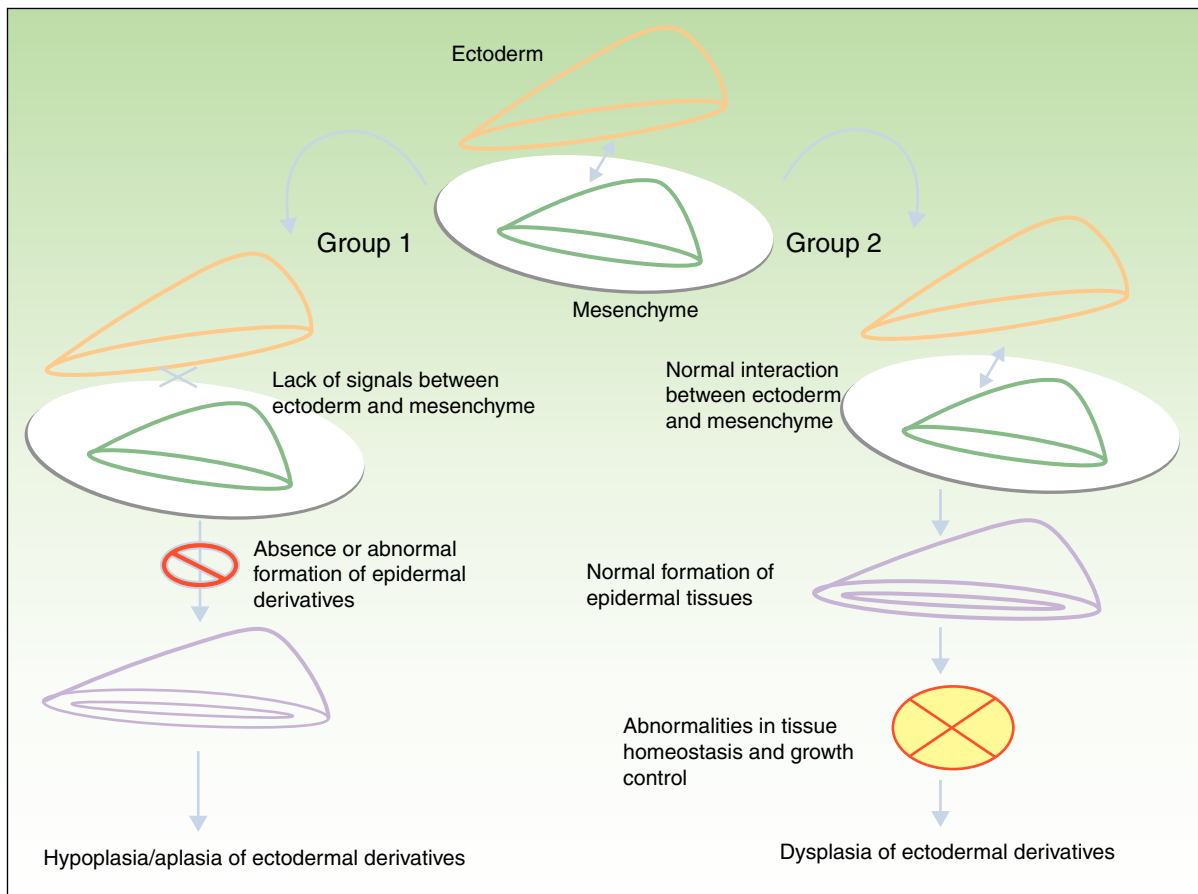


Figure 1 Pathogenic mechanisms in ectodermal dysplasias. In group 1, there is an abnormal interaction between the ectoderm and the mesenchyme, thereby impeding correct differentiation of the epidermal derivatives, which are hypoplastic or aplastic. In group 2, the interaction between the ectoderm and mesenchyme is normal, and the epidermal derivatives differentiate normally, but tissue homeostasis and growth are abnormal, so dysplasia is present in the ectodermal derivatives. Adapted from Priolo.²

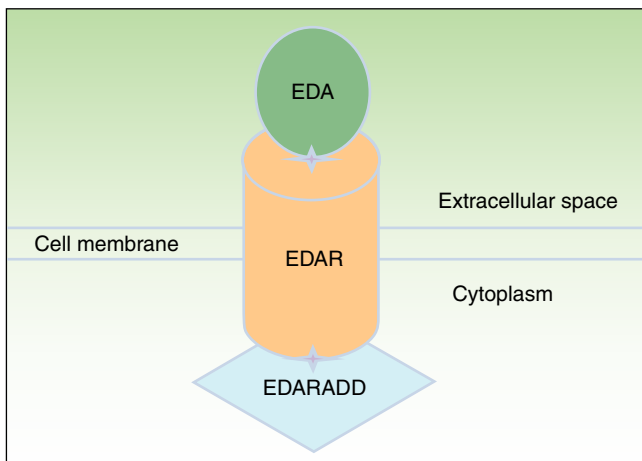


Figure 2 Ectodysplasin-EDAR-EDARADD pathway. The ectodysplasin-A (EDA) protein binds to an ectodysplasin-A receptor (EDAR), located in the extracellular region of EDAR. EDAR has an extracellular region, a transmembrane region, and a death domain in its intracellular region. This death domain binds to the death domain of EDAR-associated death domain (EDARADD). Adapted from Lu et al.³⁶

that consists of 448 amino acids and belongs to the TNF receptor superfamily. It has an extracellular region, a transmembrane region, and a death domain in its intracellular region.¹⁹ A death domain is a protein interaction module that interacts with the death domains of other proteins, thereby triggering metabolic cascades that are often implicated in regulating apoptosis and inflammation through the NF- κ B cascade. The extracellular domain of EDAR is essential for binding to EDA-A1, whereas the death domain on the intracellular region plays an important role in the initiation of apoptotic transduction signals.²⁰ The EDAR death domain has an associated death domain protein (EDARADD), which contains a 208 amino-acid sequence.²¹ The integrity of the death domains of both proteins is thus vital for their interaction²² and a normal regulation of embryonic morphogenesis.¹⁷ Therefore, the EDA-mediated signaling pathway is essential for appropriate organ development and ectoderm-derived structures, such as hair, nails, pituitary gland, mammary glands, sweat glands, nose, eyes, and tooth enamel.²³

The *EDAR* gene, which has 12 exons, is located on chromosome 2 (locus 2q11-q13)¹⁹ and encodes the receptor for EDA-A1 (EDAR).¹⁷ To date, at least 41 *EDAR* mutations have been described. Most of these involve exon 12, which

Table 1 Group 1 Disorders.

Changes in the Signaling Pathways that Modulate NF- κ B Activity						
Pathway	Gene	Locus	Protein	Disease	OMIM	Type of Inheritance
Ectodysplasin-EDAR-EDARRADD signaling pathway	<i>ED-1</i>	Xq12-q13	Ectodysplasin	Anhidrotic ectodermal dysplasia	omim:305100305100	XL
	<i>EDAR</i>	2q13	EDAR	Anhidrotic ectodermal dysplasia	omim:129490129490	AD
	<i>EDARADD</i>	1q42-2-q.43	EDAR-associated death domain	Anhidrotic ectodermal dysplasia	omim:224900224900	AR
NEMO regulatory pathway	<i>NEMO/IκBγ</i>	Xq28	NF- κ B	Incontinentia pigmenti	omim:308300308300	XL
				Anhidrotic ectodermal dysplasia with immunodeficiency	omim:300291300291	XL
	<i>IκBα</i>	14q13	I κ B α	Anhidrotic ectodermal dysplasia with osteopetrosis and immunodeficiency	omim:300301300301	XL
			Anhidrotic ectodermal dysplasia with immunodeficiency	omim:164008164008	AD	
Abnormalities in Gene Transcription/Expression Regulators						
Gene	Locus	Protein	Disease	OMIM	Type of Inheritance	
<i>p63</i>	3q27	P63	EEC syndrome	omim:604292604292	AD	
			AEC syndrome	omim:106260106260	AD	
			ADULT syndrome	omim:103285103285	AD	
			Limb-mammary syndrome	omim:603543603543	AD	
			Rapp-Hodgkin syndrome	omim:603543603543	AD	
<i>DLX3</i>	17q21	DLX3	Tricho-dento-osseous syndrome	omim:190320190320	AD	
<i>MSX1</i>	4p16.1	MSX1	Witkop disease	omim:189500189500	AD	
<i>EVC2</i>	4p16	EVC2	Ellis-van Creveld disease	omim:225500225500	AR	
<i>EVC</i>	4p16	EVC	Weyers acro-dental dysostosis	omim:193530193530	AD	
			Ellis-van Creveld disease	omim:225500225500	AR	

Adapted from Priolo.² Abbreviations: AD, autosomal dominant transmission; ADULT; acro-dermato-ungual-lacrimal-tooth; AEC, ankyloblepharon-ectodermal dysplasia-cleft lip/palate; AR, autosomal recessive transmission; EEC, ectrodactyly-ectodermal dysplasia-cleft lip/palate; OMIM, Online Mendelian Inheritance in Man; XL, X-linked.

encodes the C-terminal region where the death domain is located.²⁴ Some of these mutations have been reported in Spanish patients.^{25,26} The *EDARADD* gene, located on chromosome 1 (locus 1q-42-q43), encodes EDARADD, which is implicated in several diseases of autosomal recessive and dominant inheritance.^{17,27} Finally, certain mutations in the *WNT10A* gene, whose product is a member of the Wnt signaling pathway and implicated in embryonic development and cell differentiation, as well as in certain physiologic processes in adults and certain cancers, have been shown to give rise to several forms of ED of autosomal inheritance, such as HED or odonto-onycho-dermal dysplasia.⁵

X-linked HED. Also known as Christ-Siemens-Touraine syndrome, X-linked HED is the most frequent form of ED, with an incidence of approximately 1 case per 100 000 births.²⁸ Given the X-linked inheritance, affected males show all or most of the typical characteristics of the disease, while female carriers show only partial manifestations. There is no genotype-phenotype relationship, and the phenotype can vary greatly among different families and within the same family group.²⁹ X-linked HED occurs as a result of mutations in the *ED1* gene, also known as *EDA*.³⁰ This gene is located on the long arm of the X chromosome (locus Xq12-q13) and encodes the EDA protein. More than 204 different *ED1* mutations have been identified.²⁴ Although a wide range

Table 2 Group 2 Disorders.

Gene	Locus	Protein	Disease	OMIM	Type of Transmission
GJB6	13q12	Connexin 30	Clouston syndrome	omim:129500129500	AD
PVRL1	11q23-q24	Nectin 1	Ectodermal dysplasia with cleft lip/palate	omim:255060255060	AD
PKP1	1q32	Plakophyllin 1	Ectodermal dysplasia with fragile skin syndrome	omim:604536604536	AR
CDH3	16q22.1	Cadherin 3	Ectodermal dysplasia with ectrodactyly and macular dystrophy	omim:225280225280	AR
WNT10A	2q35	Wnt10A	Odonto-onycho-dermal dysplasia	omim:257980257980	AR

Adapted from Priolo.² Abbreviations: AD, autosomal dominant transmission; AR, autosomal recessive transmission; OMIM, Online Mendelian Inheritance in Man.

of deletions and insertions have been reported, just 1 mutation accounts for 80% of the cases.^{31,32} Recently, a Spanish group has identified a family with a previously unreported mutation in this gene.³³

The distinctive clinical features include skin, tooth, and sweating abnormalities. In the neonate, the characteristic abnormalities are not especially notable, and so diagnosis in the first days of life is difficult. Up to 70% of boys with X-linked HED show skin desquamation during the neonatal period. This finding is also relatively common among female carriers. Cases of presentation as collodion baby have been reported.³⁴ Alopecia is usually the first notable characteristic, although alopecia universalis is rare (Fig. 3).³⁵ During



Figure 3 X-linked hypohidrotic ectodermal dysplasia with alopecia universalis of the scalp, eyebrows, and eyelashes in an infant. Also of note is the frontal bulging, lip eversion, and discrete perioral eczematiform lesions.

childhood, most patients have sparse, fine, blond hair which darkens and thickens as the individuals get older. The eyebrows and beard are also sparse, but the eyelashes and armpit and pubic hair can be normal. Other body hair is also sparse or even absent.³⁶ In some patients, hair shaft abnormalities may be present, though this finding is not specific for X-linked HED.³⁷ Tooth abnormalities may become apparent during lactation as hypoplasia of the alveolar crests (Fig. 4). The number of missing and malformed teeth varies greatly between families and within the same family and there are also substantial variations between sexes.³⁸ Morphologic variations are more evident in the anterior teeth. The most common type of affected tooth has an abnormal crown and takes on a cone or peg shape (Fig. 5).³⁵ An X-ray will usually reveal taurodontism, an abnormality mainly of the molars characterized by a shortening of the root although the total height of the tooth is unchanged, yielding a prismatic form.³⁸ The ability to sweat is reduced or absent, and so patients are predisposed to developing hyperthermia



Figure 4 Alveolar hypoplasia in an infant with X-linked hypohidrotic ectodermal dysplasia.



Figure 5 Hypohidrotic ectodermal dysplasia with dental agenesis and characteristic cone-shaped teeth.

with physical exercise or high ambient temperatures.³⁶ More than 90% of children have recurrent fever spikes with no apparent cause during the first year of life. Febrile seizures occur in approximately 6% of children with X-linked HED.³⁹ Sweat disorders make a substantial contribution to the morbidity and mortality associated with X-linked HED; thus the mental retardation reported in between 30% and 50% of some series could be due to the damage caused by prolonged fever and febrile seizures.³⁵ Although a genotype-phenotype correlation is apparent in terms of extent of sweat gland dysfunction, the risk of hyperthermia cannot be predicted and there is no correlation with morbidity and mortality.⁴⁰

In the full syndrome, patients have characteristic facies, with a prominent frontal bone and chin, sunken nasal bridge, thick and everted lips, large ears, and a broad and high maxillary bone.³⁵ Most abnormalities in craniofacial morphology can be attributed to the absence of teeth, although some authors have suggested that changes in embryonic morphogenesis could also be responsible.⁴¹ In addition to the typical characteristics, from childhood onwards, patients usually develop a dry, thin, shiny skin, periocular hyperpigmentation and fine periocular wrinkles giving the appearance of premature aging (Fig. 6), as well as small papular lesions reminiscent of sebaceous hyperplasia.³⁵ Up to two-thirds of patients have atopic eczema, which can be difficult to control. Palmoplantar keratoderma is rare in X-linked HED, although it is often found in hidrotic ectodermal dysplasia and Clouston syndrome.⁴² The nails can be normal or fragile, but they are not usually especially dystrophic.³⁵

Abnormalities in the mucosal glands can cause very thick nasal secretions that predispose patients to the development of respiratory tract infections.^{35,36} The decreased salivary secretion could, according to some authors, increase the risk of tooth decay and oral fungal infections, and also hinder food ingestion and speech.⁴³ The reduction or absence of meibomian glands and Moll and Zeiss gland

Figure withdrawn at the patient's request.

Figure 6

dysfunction can lead to the development of eyelid abnormalities from the second decade of life.⁴⁴ Thick cerumen in the ears may lead to obstruction of the external auditory canal and thus hypoacusia.⁴³ Mammary glands may be hypoplastic or even show complete agenesis,⁴⁵ but sexual development is usually normal.³⁵

Female carriers of an *ED1* mutation may be asymptomatic or show mild or moderate clinical manifestations.⁴⁶ A characteristic sign suggestive of the disease is present in 70%.⁴⁷ The most frequent manifestations are tooth abnormalities, mild hypohidrosis, and differing degrees of hypotrichosis.^{46,48} A patchy distribution of body hair can be found, with somewhat depressed alopecic and xerotic areas in a Blaschkoid distribution alternating with areas of normal skin. The affected areas are more evident with tanning and during childhood.⁴⁶ In some carriers, a radial shift in the distal phalange of the index finger has been reported.⁴⁹

HED of autosomal transmission. The autosomal forms of HED, which are much less frequent than the X-linked forms, can follow a dominant or recessive inheritance, and are caused in most cases by mutations in the *EDAR* or *EDARADD* genes.¹⁹ Patients with autosomally transmitted HED are clinically indistinguishable from male patients with X-linked HED.^{50,51} In general, patients with the recessive form have more severe abnormalities, whereas the clinical spectrum of the dominant forms is variable,^{52,53} and at times very mild, with clinical manifestations similar to those observed in X-linked HED carriers.^{26,53} In recessive transmission, heterozygous carriers are clinically indistinguishable from individuals with normal genotypes.⁵⁴ The genetic heterogeneity of HED and the clinical similarity between patients with different transmission modes can be explained by the involvement of ectodysplasin, *EDAR*, and *EDARADD* in the same pathway, as activation of NF- κ B by ectodysplasin is necessary for the initiation, formation, and differentiation of ectodermal derivatives.⁵⁴



Figure 7 Hypohidrotic ectodermal dysplasia. The Minor test and the iodine-starch test show absent or decreased sweating in affected individuals. Almost complete absence of sweating was observed on the trunk of this patient.

Diagnosis of HED. Early diagnosis of HEDs is important to prevent complications essentially resulting from an ineffective control of body temperature during the neonatal period. Diagnosis can be made on clinical grounds in patients with the full syndrome, but it can be difficult in partial cases and in carriers of the disease.^{34,35} In such cases, it is necessary to perform additional tests to demonstrate decreased sweating or a decreased number of eccrine glands.

The Minor test or iodide-starch test is very useful for confirming the absence of sweating, which is generalized in affected individuals and patchy in female carriers, who have normally functioning eccrine glands alternating with eccrine glands with reduced function in areas with a Blaschkoid distribution (Fig. 7). Examination of large areas of the body, such as the back, can more readily reveal this mosaic distribution. Moreover, such an examination is useful for differentiating between X-linked HED carriers and females affected by autosomally transmitted HED, in whom the function of the sweat glands is almost completely absent.⁴⁶ Other methods for assessing sweating include iontophoresis after applying pilocarpine to the forearm, sweat pore count, and measurement of skin conductance or temperature; such methods are useful for screening but are less sensitive in patients with residual gland function.⁴⁰ Skin biopsy is not normally essential for confirmation, but the lack of eccrine glands has a positive predictive value and diagnostic specificity of 100%.³⁷ Molecular analysis is the only way of determining which gene is involved, detecting carriers, and confirming the type of inheritance. This information is vital for genetic counseling.

Prognosis and treatment. For many years, the mortality rate in children with X-linked HED was thought to be around 30% during early childhood,⁴³ but today it is known to be around 13%. Complications appear in the first years of life as a result of hyperthermia and respiratory infections, but after childhood, life expectancy is normal.³⁹ Education of the patients and their families is essential to prevent hyperthermia. Physical activity does not need to be completely avoided, but patients and their families should be aware that high body temperature can cause symptoms such as headache, nausea and vomiting, dizziness, excessive

tiredness, and muscle cramps. They should also be made familiar with techniques for reducing body temperature (immersion in water, air conditioning, cold drinks, use of refrigerated devices, etc.). Water sports are ideal for these patients.

There is no treatment for the associated skin disorders or periorcular hyperpigmentation, and outbreaks of atopic dermatitis may be difficult to treat. Some authors have suggested these patients have an increased risk of melanoma, and so a full physical examination is recommended once a year.⁵⁵ Management of children with HED also includes early dental care to prevent maxillary hypoplasia and gum atrophy, which if severe may hinder chewing and language development in addition to being a notable aesthetic problem. Other specialists may also be involved in the care of these patients, for example, ear-nose-throat specialists when nasal and cerumen secretion is a problem, ophthalmologists when eye dryness or problems with the eyelids are present, pulmonologists in the event of respiratory tract infections and, in some case, psychologists.³⁶ Gene therapy with recombinant *EDA* is still in the experimental phase, but it may offer hope for these patients in the future.⁵⁶⁻⁵⁸ Finally, as is the case in many other genetic diseases, patients diagnosed with HED will need continuous updates about their disease and socioeconomic support. We therefore recommend that patients contact the Spanish Association of Patients With HED (abbreviated as AADE in Spanish): <http://www.displasiaectodermica.org>.

NF- κ B Signaling Pathway

NF- κ B is a transcription factor that regulates the expression of multiple genes implicated in immune and inflammatory responses, reaction to stress, cell adhesion, and protection against apoptosis.⁵⁹⁻⁶¹ In most cells, NF- κ B is kept in an inactive state through cytoplasmic sequestering by the NF- κ B inhibitory protein (I κ B). Several stimuli, such as interleukin (IL) 1, TNF- α , lipopolysaccharides (bacterial endotoxins), and double-stranded RNA (from viral infections) lead to activation of the cell membrane receptors of the TNF family, such as EDAR and the receptor activator of NF- κ B (RANK).⁶² Activation of these receptors leads to degradation of I κ B through the I κ B kinase (I κ K) complex, which is phosphorylated to form I κ B, so allowing NF- κ B translocation to the nucleus. In the nucleus, the NF- κ B induces genetic transcription, triggering critical inflammatory and immune responses in the development of T and B cells and in osteoclast function and growth of epidermal cells.⁶³ The I κ K complex consists of at least 3 subunits: I κ K1/I κ K α , I κ K2/I κ K β , and NEMO/I κ K γ (essential modulator of NF- κ B). I κ K1 and I κ K2 act as catalytic subunits, whereas NEMO is a structural and regulatory subunit, essential for the complex to function as a unit. If NEMO is not present, NF- κ B shows no response to stimuli,^{64,65} as NEMO is the main molecule that provides this signaling from the cytoplasm nucleus (Fig. 8).⁶⁶

Both activation and inhibition of NF- κ B have been associated with the development of inflammatory skin conditions.⁶⁷ Mutations in 2 genes, *NEMO* and *I κ B α* , have been shown to give rise to a heterogeneous group of genetic disorders that include incontinentia pigmenti; X-linked HED with immune deficiency; osteopetrosis, lymphedema, and HED with immune deficiency; and autosomal dominant HED with immunodeficiency.^{61,68,69}

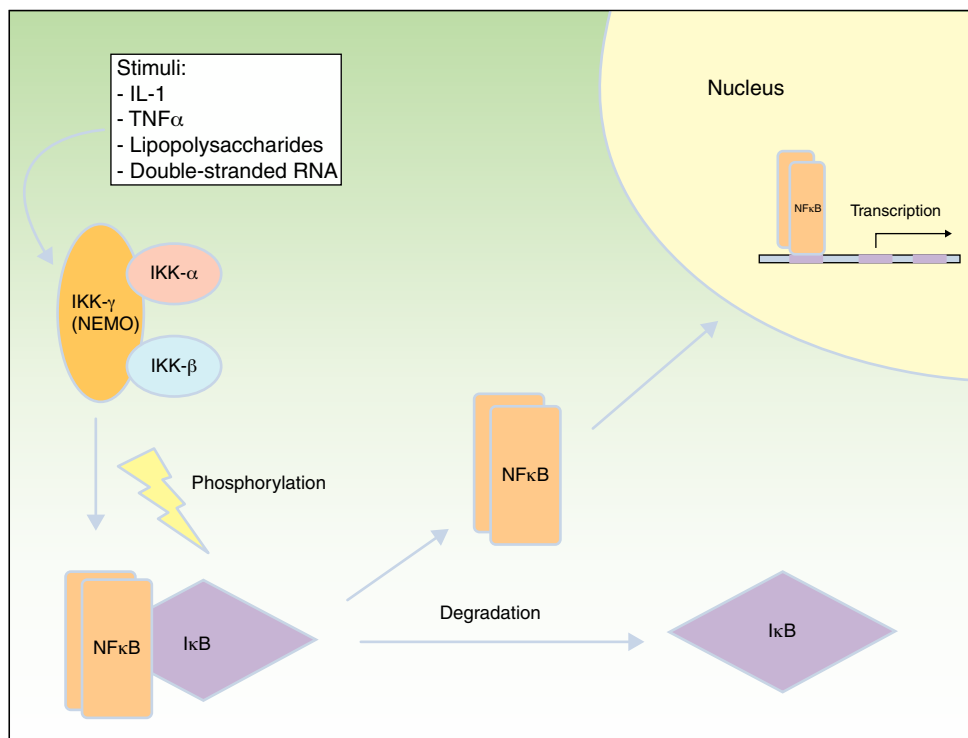


Figure 8 NEMO (nuclear factor [NF] κ B essential modulator) and NF- κ B pathway. NEMO (also known as NF- κ B inhibitory protein [IKB] kinase [IKK] γ) is a regulatory component of the Ikk complex. I κ B is activated and phosphorylated with different stimuli, leading to its degradation. NF- κ B is released and transported to the cell nucleus, where it activates the transcription genes. IL indicates interleukin; TNF, tumor necrosis factor. Adapted from Nelson.⁷³

Incontinentia pigmenti. Incontinentia pigmenti is a rare disease whose exact prevalence is not known. It is caused by mutations in the *NEMO* gene, which is located on the X chromosome (locus Xq28).^{70,71} Approximately 97% of the patients are female as the condition causes intrauterine death in most males. Most female patients survive due to the selective elimination of cells that express the X chromosome with the mutation.⁷² The clinical expression of the disease is variable.^{70,73} Nevertheless, male patients can also have the condition in the case of somatic mosaicism or XXY trisomy.^{65,74,75}

The skin manifestations are the most striking but not the most serious. They are present in almost all patients, with onset during the neonatal period. The lesions are distributed along the Blaschkoid lines and are usually divided into 4 stages: vesicular, verrucous, hyperpigmented, and atrophic (Fig. 9).⁶⁵ The first stages may go unnoticed, and the patients only show mild hyperpigmented lesions that are not detected until the patient gives birth to an affected daughter. Some women might not even show any clinical manifestations despite being carriers of the mutation.⁷³ The most important clinical problems are vision disorders and neurological deficits, but fortunately, these are less common than the skin manifestations, appearing in 40% and 30% of the patients, respectively.⁷⁶ In addition, patients may show other problems such as alopecia, tooth abnormalities (cone or peg-shaped teeth, hypodontia, or anodontia), and nail dystrophy.⁷³ Given that a detailed review of this disorder is beyond the scope of this article, we refer the interested reader to other literature sources.

X-Linked HED With Immunodeficiency. X-linked hypohidrotic ectodermal dysplasia with immunodeficiency is a rare X-linked recessive disorder that affects mainly males,⁷⁷ although some cases have been reported in female patients.^{78,79} The estimated incidence is 1 case per 250 000 newborns.⁸⁰ Most patients show small deletions or non-sense mutations in the zinc finger domain (zinc-bound region that allows interaction with other molecules) of *NEMO* that do not lead to complete loss of NF- κ B activation, as occurs with incontinentia pigmenti, but rather an altered or reduced function of the NF- κ B pathway.⁷⁷

Both males and females with X-linked HED with immunodeficiency have mothers with skin lesions reminiscent of incontinentia pigmenti, as well as variable manifestations of HED, including cone-shaped teeth.^{81,82} Some patients have prominent superficial veins.⁸³ A skin phenotype has also been described with initial involvement of intertriginous areas. The lesions, which have a seborrheic appearance, progress to erythroderma and are reminiscent of lesions in patients with congenital immunodeficiencies such as combined severe immunodeficiency or Omenn syndrome.⁸⁴ In some cases, incontinentia pigmenti lesions have been observed, thereby illustrating the complexity and overlap between different diseases derived from *NEMO* mutations.⁸¹ Patients with X-linked HED with immunodeficiency have a poor inflammatory response caused by abnormalities in cell response to proinflammatory cytokines (IL-1 β , IL-18, and TNF- α).⁶⁰ The most frequently reported immunodeficiency is dysgammaglobulinemia, with normal or low IgG levels, although elevated IgA, IgM, and IgE levels can also



Figure 9 Different clinical aspects of incontinentia pigmenti. A, Vesicular phase in a neonate. B, Linear hyperpigmented lesions on the legs in a later stage. C, Dental agenesis and presence of cone-shaped teeth. D, Nail dystrophy, with trachyonychia and pitting of the nail plate.

be observed.⁸⁵ In addition, there have been reports of defects in natural killer (NK) cells, decreased TNF and IL-12 production, abnormal immune response to polysaccharide stimuli with the inability to form specific antibodies to *Streptococcus pneumoniae*, and a delayed or absent production of isohemagglutinins.^{77,85,86} Thus, in addition to the characteristic HED phenotype, these patients have serious and recurrent bacterial infections in the lower respiratory tract, skin, soft tissues, bones, gastrointestinal tract, and meninges.^{65,87} The pathogens are usually pyogenic bacteria such as *Spneumoniae*, *Haemophilus influenzae*, *Staphylococcus aureus*, *Klebsiella*, *Salmonella*, and *Pseudomonas* species, as well as mycobacteria, cytomegalovirus, herpes simplex virus, and *Pneumocystis jiroveci*.⁸⁰ Although the infectious complications are generally considered the most life-threatening, defects in NF- κ B signaling also increase the risk of inflammatory diseases and inflammatory colitis in particular.⁸⁰

Female carriers have a range of manifestations, from normal teeth to mild hypodontia or cone-shaped teeth. There has been an anecdotal report of 2 women with mottled skin pigmentation.⁷⁷

Dominant X-linked HED with immunodeficiency. In cases of dominant transmission, the patients have an *IkB α* mutation that prevents phosphorylation and degradation of the *IkB α* protein resulting in abnormal activation of NF- κ B.⁸⁸ These patients have the classic characteristics of HED, along with T-cell immunodeficiency, giving rise to recurrent infections and immune deficiencies.⁸⁹ Although patients with *NEMO* and *IkB α* mutations share certain clinical manifestations, their immunologic phenotypes are different.⁹⁰ These patients have marked T-cell deficiency characterized by loss of CD45RO⁺ cells and abnormal T-cell receptor-mediated

stimulation of lymphocytes when studied in vitro. They are unable to respond to TNF- α stimulation, and have abnormal antibody production and greater susceptibility to infections by gram-positive and gram-negative bacteria, as in patients with *NEMO* mutations. However, these patients have normal NK cell activity, and so are not susceptible to mycobacterial infections.⁸⁹ It seems that hypomorphic mutations in the stop codon of *IkB α* could produce a small change in NF- κ B activation, giving rise to less severe immunodeficiency.⁸⁸

Mutations in the stop codon of the *NEMO* gene yield osteopetrosis, lymphedema, and HED with immunodeficiency.⁷⁷ As with children with X-linked HED with immunodeficiency, these patients have HED skin manifestations, may have skin lesions similar to those of incontinentia pigmenti, and have an abnormal inflammatory response.^{85,91} Immunodeficiency is particularly severe, and so these patients experience unusual, aggressive, and often fatal infections from early childhood.^{60,65} The distinctive clinical characteristics are osteopetrosis and lymphedema, which are thought to be due to abnormal signaling involving RANK, a receptor of the TNF family present in osteoclast progenitor cells.^{87,92} Differentiation and function of these 2 types of cells depends on NF- κ B.⁶⁹ In these patients, osteoclast differentiation is abolished or severely reduced, and so the bones formed are dense but fragile.⁹² The *NEMO* gene encodes the vascular endothelial growth factor receptor 3, an activator of the NF- κ B pathway.⁹³ Such a *NEMO* mutation would be associated with interference in this pathway, giving rise to lymphatic vessel dysfunction and characteristic lymphedema.⁶⁹

Female carriers of a hypomorphic mutation in the *NEMO* gene may be asymptomatic or have several manifestations of incontinentia pigmentaria.^{69,94}

Diagnosis of NEMO/IkB α -derived disorders. Mutations in the *NEMO* gene should be considered in children with refractory extensive seborrheic or atopic dermatitis, particularly when the facial characteristics of HED are present or the mother has a history of incontinentia pigmenti.⁸⁴ A biopsy of the skin lesions is recommended in patients with HED with immunodeficiency, as they can develop similar lesions to incontinentia pigmenti after childhood.⁸¹

Treatment. The management of patients with HED manifestations is similar to that presented in the preceding section. Treatment of immunodeficiency may include immune therapies and an aggressive approach to infections, including prophylaxis against gram-positive and gram-negative bacteria, mycobacteria, herpes simplex virus, and *Pneumocystis jiroveci*.⁸⁰ Even with such an approach, morbidity and mortality are high.⁷⁷ Hematopoietic cell transplantation offers the possibility of immune reconstitution, but it brings with it the inherent risks of immunosuppression.⁶⁹ Little has been published on the subject. Allogeneic transplantation of hematopoietic progenitors could correct the immunodeficiency associated with diseases due to NEMO or IkB α mutation, but the causative mutations do not exclusively affect the hematopoietic system, and so constitutional manifestations not related to immunodeficiency are not corrected by transplantation. Inflammatory colitis might therefore even worsen on correcting immunodeficiency.⁹⁵

Changes in the Transcription and/or Expression of Regulators of Certain Genes

Disorders Derived From *p63* Mutations

The *p63* gene, also known as *TP63* (tumor protein 63), is located on chromosome 3 (locus 3q27) and encodes the transcription factor p63, involved in ectodermal development. The regions of greatest biological importance are the DNA binding domain (a region that allows transactivation through binding of p63 to DNA), the sterile alpha motif (SAM), which is thought to participate in protein-protein interactions, and the transactivation inhibition domain (TID), which is located next to the SAM domain and could be involved in balancing the effects of different isoforms of TP63.⁹⁶ The p63 protein is expressed very early during embryogenesis and plays an essential role in inducing epidermal differentiation and proliferation and in other processes including facial and limb development.⁹⁶⁻⁹⁸ Lack of expression of this protein during early development of ectodermal structures might block a chain of interactions between the epithelium and the mesenchyme, thereby interfering in normal morphogenesis.⁹⁹ In addition, p63 regulates expression of P-cadherin, a critical regulator of hair development.¹⁰⁰

Heterozygous mutations in the *p63* gene are responsible for at least 6 different syndromes that combine ED, orofacial clefts, and limb malformations.¹⁰¹ The ectrodactyly-ectodermal dysplasia-cleft lip/palate syndrome (EEC) syndrome is the prototype syndrome. Other syndromes include ankyloblepharon-ectodermal dysplasia-cleft lip/palate (AEC) syndrome, limb-mammary syndrome, acro-dermato-ungual-lacrimal-tooth (ADULT) syndrome, Rapp-Hodgkin syndrome, and split-hand/foot malformation. All these syndromes present at least 1 of the key characteristics of EEC syndrome.¹⁰¹ There is a strong

genotype-phenotype correlation that is dependent on the location of the *p63* mutation.⁹⁶ The most frequently mutated amino acid residues are R204, R227, R279, R280, and R304. The phenotype related to the R204 mutation is very similar to the complete phenotype of EEC syndrome, but the patients have a lower frequency of hypohidrosis and orofacial cleft; the R227 mutations are rarely associated with orofacial cleft or syndactyly, but the incidence of renal problems and hypohidrosis is higher whereas hearing disorders are absent; the R279 mutation is the only one that gives rise to ankyloblepharon, which is usually associated with ectrodactyly (deformity of the limbs in which part or all of the fingers are missing, giving the hands or feet the form of a lobster claw); patients with the R280 mutation frequently have skin manifestations and syndactyly but hypohidrosis and hearing or renal disorders are absent; and finally, the R304 mutation is associated with a higher percentage of orofacial cleft, syndactyly, and hearing disorders.¹⁰¹

Ectrodactyly-Ectodermal Dysplasia-Cleft Lip/Palate Syndrome. EEC syndrome is relatively common. Although it can appear in sporadic cases, autosomal dominant inheritance can be detected in most cases.¹⁰² Mutations in the *p63* gene are present in 98% of patients with the classic phenotype; these mutations are generally point mutations in the DNA binding domain, while mutations in the SAM and TID domains are rare.⁹⁶ At least 30 different mutations have been identified, of which 5 (mutations in amino acids R204, R227, R279, R280, and R304 of *p63*) are responsible for 86% of the cases.¹⁰¹

The most frequent abnormalities are malformations of the limbs, ED, and orofacial cleft (Fig. 10), followed by tear duct abnormalities, genital malformations, and deafness, although the clinical manifestations vary greatly within and among families.¹⁰³ Among the most representative limb malformations are ectrodactyly and syndactyly (Fig. 11). Ectodermal dysplasia may be associated with sparse, hypopigmented hair, absence of eyebrows and eyelashes, and alopecia. The skin is usually fine, dry, and of an atopic appearance, while the nails are dystrophic and may appear pitted. Perioral lesions and angular cheilitis may be present in the oral commissures as a result of anatomic changes caused by reconstructive surgery for cleft lip/palate.^{104,105} Tooth abnormalities such as hypodontia or anodontia have also been reported, as well as a propensity



Figure 10 Anodontia and dental dysgenesis in a patient with ectrodactyly-ectodermal dysplasia-cleft lip/palate syndrome.



Figure 11 Two limb abnormalities most characteristic of ectrodactyly-ectodermal dysplasia-cleft lip/palate syndrome.

to tooth decay due to defective enamel and changes in the salivary gland function. Orofacial cleft is common and may be accompanied by maxillary and malar hypoplasia. Tear duct stenosis contributes to keratitis, which is sometimes associated with photophobia.¹⁰¹ Urogenital and anogenital abnormalities (micropenis, hypospadias, vaginal septum, and female genital hypoplasia), hypothalamic-hypophyseal insufficiency, thymic hypoplasia, and mental retardation may also be present.^{106,107} Isolated cases have been reported of an association with white sponge nevus,¹⁰⁸ as well as micrognathia, cleft soft palate, and glosptosis (Pierre Robin sequence), but the relevance of this association is not known.¹⁰⁹

Prenatal ultrasound diagnosis is an important aspect of diagnosis of EEC syndrome, not only because ectrodactyly and cleft lip/palate may alert the physician, but also because severe associated genitourinary abnormalities can be detected.¹⁰⁷

Ankyloblepharon-ectodermal dysplasia-cleft lip/palate syndrome. The AEC disorder follows an autosomal dominant transmission pattern and results from non-sense mutations in the SAM domain of protein p63, implicated in the interaction with other proteins participating in the regulation of transcription and development of skin appendages. The characteristic clinical triad consists of ankyloblepharon, ectodermal defects, and cleft lip/palate (Fig. 12).¹¹⁰ Ankyloblepharon is a condition in which fibrous bands between the eyelids prevent these from opening or moving normally. The ectodermal abnormalities are similar to those of other EDs.⁹⁶ In addition to these abnormalities, skin erosions can be observed in the scalp, head, neck, skin folds, palms, and soles. The erosions heal poorly and tend to become superinfected. On the upper part of the trunk, they heal to leave residual scarring of a cribriform, reticular, stellate, or punctate pattern. Although the exact reasons for the erosions and delayed wound healing are not well known, a contributing factor could be the role of protein p63 in the formation of basal cells and epidermal differentiation.¹¹¹ At birth, patients may present manifestations such as erosive lesions, congenital erythroderma, ichthyosiform scaling, and even collodion membrane which may lead to an initial suspicion of epidermolysis bullosa or a keratinization disorder.¹¹¹ Changes in pigmentation (hyperpigmentation or



Figure 12 Shorter eyelid skin crease, sparse hair, and (surgically repaired) cleft lip in a patient with ankyloblepharon-ectodermal dysplasia-cleft lip/palate syndrome.

hypopigmentation) are also frequent (Fig. 13). Reticular hyperpigmentation, which becomes more accentuated with age, can be observed in the large skin folds, whereas characteristic hypopigmentation, which improves as the patient gets older, may be present around the eyes giving a mask-like appearance. Almost all patients have sweating disorders.¹¹¹ The extent of alopecia is variable and does not seem to be related to age or severity of prior erosions in the scalp. Hair may be thick, wiry, fragile, or matt, and have variable or even 2-tone pigmentation (pigmented and pale hairs). Hair shaft abnormalities, such as pili annulati, pili torti, and pili canaliculi, and irregular indentation may be present.¹¹² Nail involvement is also variable (Fig. 14) and ranges from complete anonychia to mild scaling of the nail plate.¹¹¹ Other skin abnormalities occasionally reported include absence of



Figure 13 Abnormal pigmentation in the inguinal fold, with areas of hypopigmentation in the central part of the fold and areas of reticular hyperpigmentation in the peripheral areas and on the vulva in a patient with ankyloblepharon-ectodermal dysplasia-cleft lip/palate syndrome.

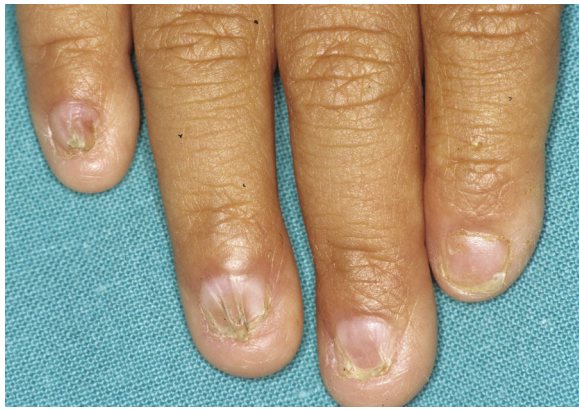


Figure 14 Fingernail abnormalities of varying severity in a patient with ankyloblepharon-ectodermal dysplasia-cleft lip/palate syndrome.

dermatoglyphs, palmoplantar hyperkeratosis, punctate keratoderma, hyperlinearity, and hyperkeratosis on the elbows and knees.¹¹¹

Facial dysmorphism includes, in addition to cleft palate and lips, small or malformed ears, maxillary hypoplasia, lack of permanent teeth, and decreased length of the eyelid skin crease. Failure to thrive, syndactyly, nasolacrimal atresia, recurrent otitis media, hearing loss, and hypospadias are also common.⁹⁶ Some individuals may experience neutropenia of unknown origin and have recurrent skin and ear infections, which may progress to bacteremia and sepsis.⁹⁶

AEC syndrome should be suspected in any neonate with erythroderma and cleft lip/palate, particularly when other skin manifestations are present, such as erosions in the scalp or skin, although ankyloblepharon is not always present. Skin biopsy, although not very specific, shows mild hyperkeratosis, epidermal atrophy, basal pigmentation, and/or incontinentia pigmenti, and a prominent superficial vascular plexus with mild perivascular infiltrate consisting mainly of lymphocytes.¹¹²

The first objective of treatment is to prevent skin erosions, and so energetic cleansing of the skin should be avoided. Secondary wound infection can be hard to treat, and so extensive daily hygiene and application of antiseptics to the erosions are recommended. The administration of antibiotics for long periods would only seem to provide a minimal or temporal improvement in the lesions. Some authors have suggested that the use of corticosteroids^{113,114} or low doses of doxycycline¹¹⁵ may be useful for reducing inflammation and improving healing, but the risk of side effects should be taken into consideration. Looking to the future, other therapeutic strategies, such as gene therapy and use of epidermal stem cells to regenerate affected skin, are under investigation.¹¹¹

Acro-dermato-ungual-lacrimal-tooth syndrome. EEC and ADULT syndromes are considered allelic disorders with overlapping clinical characteristics, such as ectodermal, limb, and tooth abnormalities. The main difference lies in the absence of orofacial cleft from ADULT syndrome.¹¹⁶ ADULT syndrome arises through mutations that affect the TID domain, implicated in transactivation of the *p63* gene.¹¹⁷ To date, 5 mutations in this gene have been reported; 2 of these have also been identified in patients with EEC syndrome.¹¹⁸

Limb-mammary syndrome. The limb-mammary syndrome, with an autosomal dominant inheritance, is caused by mutations in the SAM and TID domains of the *p63* gene.¹¹⁹ Its clinical characteristics overlap with those of EEC, AEC, ADULT and Rapp-Hodgkin syndromes, as well as with ulnar-mammary syndrome (caused by mutations in the *TBX3* gene at locus 12q24.1).¹²⁰ The syndrome is characterized by ectrodactyly, hypoplasia of the mammary glands and nipples, cleft palate (without a cleft lip), and the absence of skin or hair abnormalities. Other manifestations include lacrimal duct stenosis, hearing loss, urogenital abnormalities, nose dysplasia, hypohidrosis, hypodontia, and gonadal dysplasia.^{99,120}

Differentiation between limb-mammary syndrome and EEC syndrome is based on 3 findings¹²⁰: hypoplasia of the mammary glands and nipples, present in all cases of limb-mammary syndrome but only occasionally in EEC syndrome; hair and skin abnormalities, absent in patients with limb-mammary syndrome; and cleft lip, present only in patients with EEC syndrome.

Rapp-Hodgkin syndrome. Rapp-Hodgkin syndrome is a disorder of autosomal dominant transmission,¹²¹ produced by mutations that, like in AEC syndrome, affect the SAM domain, thereby explaining the considerable clinical and molecular overlap between the 2 syndromes.^{96,122} Some authors consider these 2 syndromes as the same disorder.¹²¹⁻¹²⁷

Tricho-dento-osseus syndrome

The tricho-dento-osseus syndrome is a condition of autosomal dominant transmission caused by different mutations in the *DLX3* gene, located on chromosome 17 (locus 17q21). This gene, which is also responsible for imperfect amelogenesis, encodes the DLX3 protein,¹²⁸ which is expressed during embryogenesis and which participates in the differentiation of tissue derived from the ectoderm, bone tissue, and cartilaginous tissue.¹²⁹ The clinical manifestations are variable and include abnormalities in tooth enamel, nail abnormalities, blond curly hair, sclerosis and thickening of cranial bones, radiographic abnormalities such as hypocalcification, and taurodontism.¹³⁰

Witkop Syndrome

Witkop syndrome, an autosomal dominant condition also known as the tooth and nail syndrome and nail dysgenesis and hypodontia, is caused by a mutation in the *MSX1* gene, located on chromosome 4 (locus 4p16.1). The *MSX1* gene participates in the formation of certain teeth (premolars, first molars, and third molars) and nails, by determining the thickness and integrity of the nail plate.¹³¹ Typical clinical characteristics are nail dysplasia and hypodontia, although the clinical manifestations are very varied. In some cases, hair abnormalities (fine or thick hair) have been reported, though most individuals have normal hair and sweat gland function.¹³²

Ellis-van Creveld Syndrome

Ellis-van Creveld syndrome follows an autosomal recessive transmission pattern and is caused by mutations in the *EVC* or *EVC2* genes; the clinical phenotype is the same for both types of mutation.¹³³⁻¹³⁵ Approximately 30% of patients

with Ellis-van Creveld syndrome have no mutations in either of these genes, suggesting that greater genetic heterogeneity may be present.¹³⁶ The syndrome is characterized by bone abnormalities, nail dysplasia, orofacial abnormalities, and cardiovascular malformations.¹³⁷ Patients have a short stature, acromesomelic limb shortening (more prominent in the distal region), and a narrow chest. Polydactyly, syndactyly, genus valgum, and various types of tooth abnormalities are common.^{137,138}

Weyers Acrodistal Dysostosis

Weyers acrodistal dysostosis is an autosomal dominant disorder whose causative mutations are located in the *EVC* and *EVC2* genes.^{133,134,137} The transmission pattern and milder phenotype distinguish this condition from Ellis-van Creveld syndrome.¹³³ Clinical expression is variable and characterized by short stature, hypotelorism, prominent ears, postaxial polydactyly, oral abnormalities (irregular, small, peg-shaped teeth and hypodontia), and onychodystrophy (dysplastic or hypoplastic nails).^{133,137}

Group 2

Hidrotic Ectodermal Dysplasia

Hidrotic ectodermal dysplasia, also known as Clouston syndrome, is caused by mutations in the *GJB6* gene which is located on chromosome 13 (locus 13q11-q12.1) and which encodes connexin 30.¹³⁹ This autosomal dominant disorder is particularly common among French-Canadian individuals,¹⁴⁰ and only appears very exceptionally de novo.¹⁴¹ Mutations in this gene can also give rise to other disorders such as autosomal dominant and autosomal recessive sensorineural deafness.¹⁴²

Connexins are transmembrane proteins that facilitate intercellular communication.¹⁴⁰ Connexin molecules form hexamers called connexons, which interact with other connexons of adjacent cells to form intercellular gap junctions.¹⁴² These channels allow the diffusion of small molecules between cells and mediate signal and nutrient exchange, thereby coordinating cell activities and response to stimuli. More than 10 different connexins have been identified in skin. Each connexin seems to have its own specific properties, and mutations are responsible for a distinct skin disorder. Thus, hidrotic ectodermal dysplasia and KID syndrome¹⁴⁰ are caused by mutations in the *GJB6* and *GBJ2* genes, which encode connexin 30 and connexin 26, respectively.¹⁴² These 2 connexins share 76% of their amino acid sequence and are coexpressed in the stratum corneum, sweat glands, and hair follicles.¹⁴⁰ This overlap, despite the different tissue expression of connexin 26 and connexin 30, implies that certain functions are shared and that there is perhaps a direct interaction between the 2 proteins in many ectodermal epithelia. In fact, both are associated with certain clinical characteristics such as nail dystrophy, hair loss, and plantar keratoderma, and recently there has been a report of a patient with a connexin 30 mutation whose clinical manifestations were similar to those of keratitis-ichthyosis-deafness (KID) syndrome.¹⁴⁰

The 3 main clinical characteristics of hidrotic ectodermal dysplasia are hair loss, nail dystrophy, and palmoplantar



Figure 15 Hidrotic ectodermal dysplasia. Sparse hair of coarse appearance (courtesy of Dr. Isabel Febrer).

keratoderma (Figs. 15 and 16).¹⁴⁰ Unlike patients with HED, patients with hidrotic ectodermal dysplasia have normal teeth and sweat and sebaceous gland function.^{140,143} Hair abnormalities are manifest as atrichia or hypotrichosis, which may progressively worsen¹⁴⁰; the fine, slow-growing hair has a disorganized structure and reduced birefringence.¹⁴⁴ Women are completely bald, whereas men show expression that varies from fair hair with focal alopecia to complete baldness.¹⁴⁵ The eyebrows and eyelashes are scant or absent, as is pubic and axillary hair.¹⁴⁰ Nail disorders range from an almost normal appearance to micronychchia or onychia; the nail plate may show thickening, desquamation, color changes, striation, and onycholysis.¹⁴⁰ Nail abnormalities in these patients may be reminiscent



Figure 16 Hidrotic ectodermal dysplasia. Toenail dystrophy (courtesy of Dr. Isabel Febrer).

of congenital pachyonychia.¹⁴⁶ Some patients also present diffuse palmoplantar keratoderma and discrete skin hyperpigmentation, which is particularly evident underneath the free edge of the nails and on the finger and toe joints, knees, and elbows.^{140,147} Eccrine syringofibroadenomas have been reported in several patients,^{148,149} and there has also been an isolated case of congenital pseudoainhum.¹⁵⁰ These clinical manifestations may be accompanied by strabism, conjunctivitis, pterygium, cataracts,¹⁴³ sensorineural deafness, polydactyly, and syndactyly,¹⁴⁷ but facial dysmorphism is not present and general physical development is normal.

Zlotogora-Ogur Syndrome or Ectodermal Dysplasia With Cleft Palate

The autosomal recessive disorder Zlotogora-Ogur syndrome is caused by mutations in the poliovirus receptor-like 1 (*PVRL1*) gene, located on chromosome 11 (locus 11q23.2), which encodes the nectin-1 protein.¹⁵¹ Nectins are calcium-independent cell-cell adhesion molecules that act at cell-cell junctions sometimes in conjunction with cadherins. So far, 4 different types of nectins have been reported.¹⁵² Nectin-1 is expressed in several ectodermal tissues, including the skin, teeth, and hair, essentially within the stratum spinosum.¹⁵³ Mutation in this gene also gives rise to nonsyndromic orofacial cleft type 7.¹⁵⁴ Characteristic clinical findings include manifestations of ectodermal dysplasia, bilateral cleft lip/palate, mental retardation, and syndactyly.¹⁵⁵ Hair is sparse and short, and when individuals are over 40 years, they may become completely bald. Eyebrows are also sparse, especially in the lateral regions. Xerosis, hypoplastic dermatoglyphs, and progressive palmoplantar hyperkeratosis are also observed. Tooth abnormalities include delay in dental eruption, microdontia, hypodontia, and anodontia. Nails are normal or mildly dysplastic. Patients have peculiar facial features, with an oval face and large anteverted ears. Syndactyly, which may be partial, is often present in the second, third, and fourth fingers, and can sometimes affect both hands.¹⁵¹ Other manifestations include deafness, genitourinary or renal abnormalities, nipple abnormalities and lumbar lordosis, and variable mental retardation.¹⁵⁶ Differential diagnosis should be established mainly with EEC syndrome, with which it shares most manifestations. The main difference is in the transmission pattern (EEC syndrome follows autosomal dominant inheritance) and limb malformations, mainly ectrodactyly, which are present in 85% of the patients with EEC syndrome.^{103,156}

In August 2010, a second nectin-derived condition was reported arising from an abnormality in nectin 4, which is encoded by *PVRL4*.¹⁵² This syndrome is similar to the one described above, but there is no cleft palate and it is known as ectodermal dysplasia-syndactyly syndrome.

Ectodermal Dysplasia-Fragile Skin Syndrome

Ectodermal dysplasia-fragile skin syndrome is an autosomal recessive condition that was recently reclassified as a epidermolysis bullosa simplex. It is caused by mutation in the plakophilin gene (*PKP1*), located on chromosome 1 (locus 1q32).¹⁵⁷ *PKP1* is a structural component of desmosomes



Figure 17 Ectodermal dysplasia-fragile skin syndrome. The patient has short, sparse, curly hair, diffuse erythema, fissured cheilitis, and areas of erosions. Currently, this syndrome has been reclassified as a simple epidermolysis bullosa.

and is expressed in the stratified squamous epithelium, the myocardium, the meninges, and part of the lymph nodes.¹⁵⁸ Like other types of epidermolysis bullosa, the condition is characterized by substantial trauma-induced skin fragility, generalized erythema, alopecia, nail dystrophy, and focal keratoderma with painful fissures (Fig. 17). Some patients present hypohidrosis, but the teeth are normal in all cases.¹⁵⁷ Histologic study shows widened intercellular spaces, separation of keratinocytes, intraepidermal clefts, acantholytic keratinocytes, and different degrees of dyskeratosis,¹⁵⁹ but to date the development of skin carcinomas has not been reported.

Ectodermal Dysplasia-Ectrodactyly-Macular Dystrophy Syndrome

Ectodermal dysplasia-ectrodactyly-macular dystrophy (EEM) syndrome is an autosomal recessive disorder caused by a mutation in the *CDH3* gene, located on chromosome 16 (locus 16q22.1); this gene encodes the cadherin-3 protein.^{160,161} Cadherins are calcium-dependent adhesion molecules with several extracellular domains, a transmembrane region, and an intracellular region. The intracellular region binds to β -catenin, which is implicated in transcription and cellular adhesion. Expression of *CDH3* during embryogenesis is important for normal development. It is expressed at least in the orofacial region and pharyngeal arches, as well as in the limbs, and it may play an important role in the morphology of the human hand.¹⁶¹ EEM syndrome is characterized by syndactyly, retinal degeneration, and sparse hair,¹⁶⁰ although the clinical manifestations

are variable. In addition to syndactyly, which may be bilateral, 1 or several phalanges may be missing or fingers or toes may be completely hypoplastic. The hands are usually more severely affected than the feet.¹⁶¹ Another key sign is progressive retinal degeneration, with gradual vision loss. Prominent pigmentation is observed in the posterior pole of the retina, along with macular atrophy. Patients may also have hypotrichosis with sparsely populated eyebrows, few eyelashes, and tooth abnormalities (hypodontia and small and very separated teeth).¹⁶²

Odonto-Onycho-Dermal Dysplasia

The autosomal recessive disorder, odonto-onycho-dermal dysplasia, is due to mutations in the *WNT10A* gene, located on chromosome 2 (locus 2q35).¹⁶³ The *WNT* genes encode a large family of glycoproteins implicated in a signaling pathway crucial for the development of ectodermal-derived tissue during embryogenesis and adult life.^{165,165} Thus, *WNT10A* is important for tooth and hair follicle formation, and for epidermal regeneration, lingual papillae formation, and sweat gland function.¹⁶⁴ Although the condition is termed odonto-onycho-dermal dysplasia, tricho-onycho-dermal dysplasia would be more appropriate given the hair involvement.¹⁶³ Characteristic skin lesions include the appearance in the facial region of reticular, erythematous, telangiectatic, atrophic plaques, which become more intense with heat.^{166,167} Another frequent observation is palmoplantar hyperkeratosis, accompanied by hyperhidrosis and painful fissuring.^{168,169} Other abnormalities include the presence of smooth tongue with reduced or absent papillae, pilaris keratosis,¹⁷⁰ hypotrichosis, and tooth and nail abnormalities.¹⁶⁷ Some patients may be slightly mentally retarded.¹⁶⁹ Different mutations in this same gene are responsible for another autosomal recessive ectodermal dysplasia, Schopf-Schulz-Passarge syndrome. This syndrome shares certain clinical features (hypodontia, nail dystrophy, palmoplantar keratoderma, smooth tongue, hyperhidrosis, and hypotrichosis), but in addition, patients have other features such as cysts in the eyelids and an increased risk of developing skin tumors.^{165,171}

In conclusion, ectodermal dysplasias are a heterogeneous group of hereditary disorders that bear many similarities and are difficult to classify. Biomolecular findings in recent years have brought us closer to a useful clinical-functional classification in clinical practice given that different genetic abnormalities in different functional pathways can be linked to a given phenotype. However, many disorders within the group of ectodermal dysplasias have yet to be studied or identified. Nevertheless, the dermatologist should be aware of the main signs and symptoms of these disorders when searching for a diagnosis. In addition, carriers should be identified so that genetic counseling can be offered.

Ethical responsibilities

Protection of human and animal subjects.

The authors declare that no experiments were performed on humans or animals for this investigation.

Confidentiality of data.

The authors declare that they have followed their hospital's protocol on the publication of data concerning patients and that all patients included in the study have received sufficient information and have given their written informed consent to participate in the study.

Right to privacy and informed consent.

The authors obtained the informed consent of patients and/or subjects mentioned in this article. The informed consent form is located in the archives of the corresponding author.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

1. Sadler TW. Langman embriología médica: con orientación clínica. 10th ed. Buenos Aires: Panamericana; 2008. p. 69–90.
2. Priolo M. Ectodermal dysplasias: an overview and update of clinical and molecular-functional mechanism. *Am J Med Genet A*. 2009;149A:2003–13.
3. McKusick V. Mendelian Inheritance in Man: A catalog of Human Genes and Genetic Disorders. 12th ed. Baltimore: The Johns Hopkins University Press; 1998.
4. Itin PH. Rationale and background as basis for a new classification of the ectodermal dysplasias. *Am J Med Genet A*. 2009;149A:1973–6.
5. Cluzeau C, Hadj-Rabia S, Jambou M, Mansour S, Guique P, Mas-moudi S, et al. Only four genes (*EDA1*, *EDAR*, *EDARADD* and *WNTA10A*) account for 90% of hypohidrotic/anhidrotic ectodermal dysplasia cases. *Hum Mutat*. 2011;32:70–2.
6. Danz DFG. Sechste Bemerkung. Von Menschen ohne Haare und Zahne. *Stark Arch Geb Frauen Neugeb Kinderkr*. 1792;4:684.
7. Thurnam J. Two cases in which the skin, hair and teeth were imperfectly developed. *Med Chir Trans*. 1848;31:71–82.
8. Darwin C. The variation of animals and plants under domestication. 2nd ed. London: John Murray; 1975. p. 319.
9. Weech AA. Hereditary ectodermal dysplasia (congenital ectodermal defect): a report of two cases. *Am J Dis Child*. 1929;37:766–90.
10. Pinheiro M, Freire-Maia N. Ectodermal dysplasias: a clinical classification and a causal review. *Am J Med Genet*. 1994;53:153–62.
11. Priolo M, Silengo M, Lerone M, Ravazzolo R. Ectodermal dysplasias: not only «skin» deep. *Clin Genet*. 2000;58:415–30.
12. Priolo M, Laganà C. Ectodermal dysplasias: a new clinical-genetic classification. *J Med Genet*. 2001;38:579–85.
13. Lamartine J. Towards a new classification of ectodermal dysplasia. *Clin Exp Dermatol*. 2003;28:351–5.
14. Irvine AD. Towards a unified classification of the ectodermal dysplasias: Opportunities outweigh challenges. *Am J Med Genet A*. 2009;149A:1970–2.
15. DiGiovanna JJ, Priolo M, Itin P. Approach towards a new classification for ectodermal dysplasias: integration of the clinical and molecular knowledge. *Am J Med Genet A*. 2009;149A:2068–70.
16. Bayés M, Hartung AJ, Ezer S, Pispas J, Thesleff I, Srivastava AK, et al. The anhidrotic ectodermal dysplasia gene (*EDA*) undergoes alternative splicing and encodes ectodysplasin-A with

- deletion mutations in collagenous repeats. *Hum Mol Genet.* 1998;7:1661–9.
17. Yan M, Wang LC, Hymowitz SG, Schilbach S, Lee J, Goddard A, et al. Two-amino acid molecular switch in an epithelial morphogen that regulates binding to two distinct receptors. *Science.* 2000;290:523–7.
 18. Headon DJ, Emmal SA, Ferguson BM, Tucker AS, Justice MJ, Sharpe PT, et al. Gene defect in ectodermal dysplasia implicates a death domain adapter in development. *Nature.* 2001;414:913–6.
 19. Monreal AW, Ferguson BM, Headon DJ, Street SL, Overbeek PA, Zonana J. Mutations in the human homologue of mouse dl cause autosomal recessive and dominant hypohidrotic dysplasia. *Nat Genet.* 1999;22:366–9.
 20. Hofmann K. The molecular nature of apoptotic signaling proteins. *Cell Mol Life Sci.* 1999;55:1113–28.
 21. Bonnert TP, Garka KE, Parnet P, Sonoda G, Testa JR, Sims JE. The cloning and characterization of human MyD88: a member of an IL-1 receptor related family. *FEBS Lett.* 1997;402:81–4.
 22. Yan M, Zhang Z, Brady JR, Schilbach S, Fairbrother WJ, Dixit VM. Identification of a novel death domain-containing adaptor molecule for ectodysplasin-A receptor that is mutated in crinkled mice. *Curr Biol.* 2002;12:409–13.
 23. Cui CY, Schlessinger D. EDA signaling and skin appendage development. *Cell Cycle.* 2006;5:2477–83.
 24. HGMD database [cited 7 Mar 2012]. Available from: <http://www.hgmd.cf.ac.uk/ac/gene.php?gene=EDAR>
 25. Moya-Quiles MR, Ballesta-Martínez MJ, López-González V, Glover G, Guillén-Navarro E. A compound heterozygous mutation in the EDAR gene in a Spanish family with autosomal recessive hypohidrotic ectodermal dysplasia. *Arch Dermatol Res.* 2010;302:307–10.
 26. Valcuende-Cavero F, Martínez F, Pérez-Pastor G, Oltra S, Ferrer I, Tomás-Cabedo G, et al. Autosomal-dominant hypohidrotic ectodermal dysplasia caused by a novel mutation. *J Eur Acad Dermatol Venereol.* 2008;22:1508–10.
 27. Bal E, Baala L, Cluzeau C, El Kerch F, Ouldik M, Hadj-Rabia S, et al. Autosomal dominant anhidrotic ectodermal dysplasias at the EDARADD locus. *Hum Mutat.* 2007;28:703–9.
 28. Zonana J. Hypohidrotic (anhidrotic) ectodermal dysplasia: molecular genetic research and its clinical applications. *Semin Dermatol.* 1993;12:241–6.
 29. Kobiela K, Kobiela A, Roszkiewicz J, Wierzba J, Limon J, Trzeciak WH. Mutations in the EDAR gene in three unrelated families reveal no apparent correlation between phenotype and genotype in the patients with an X-linked anhidrotic ectodermal dysplasia. *Am J Med Genet.* 2001;100:191–7.
 30. Kere J, Srivastava AK, Montonen O, Zonana J, Thomas N, Ferguson B, et al. X-linked anhidrotic (hypohidrotic) ectodermal dysplasia is caused by a mutation in a novel transmembrane protein. *Nat Genet.* 1996;13:409–16.
 31. Pääkkönen K, Cambiaghi S, Novelli G, Ouzts LV, Penttinen M, Kere J, et al. The mutation spectrum of the EDAR gene in X-linked anhidrotic ectodermal dysplasia. *Hum Mutat.* 2001;17:349.
 32. Schneider P, Street SL, Gaide O, Hertig S, Tardivel A, Tschopp J, et al. Mutations leading to X-linked hypohidrotic ectodermal dysplasia affect three major functional domains in the tumor necrosis factor family member ectodysplasin-A. *J Bio Chem.* 2001;276:18819–27.
 33. Cañueto J, Zafra-Cobo MI, Ciria S, Unamuno P, González-Sarmiento R. A novel EDAR gene mutation in a Spanish family with X-linked hypohidrotic ectodermal dysplasia. *Actas Dermosifiliogr.* 2011;102:722–5.
 34. The Executive and Scientific Advisory Boards of the National Foundation for Ectodermal Dysplasias. Scaling skin in the neonate: a clue to the early diagnosis of X-linked hypohidrotic ectodermal dysplasia (Christ-Siemens-Touraine syndrome). *J Pediatr.* 1989;114:600–2.
 35. Anoop TM, Simi S, Mini PN, Ramachandran M, Jabbar PK, Rajakumari PK, et al. Hypohidrotic ectodermal dysplasia. *J Assoc Physicians India.* 2008;56:268–70.
 36. Lu PD, Schaffer JV. Hypohidrotic ectodermal dysplasia. *Dermatol Online J.* 2008;14:22.
 37. Rouse C, Siegfried E, Breer W, Nahass G. Hair and sweat glands in families with hypohidrotic ectodermal dysplasia: further characterization. *Arch Dermatol.* 2004;140:850–5.
 38. Crawford PJ, Aldred MJ, Clarke A. Clinical and radiographic dental findings in X linked hypohidrotic ectodermal dysplasia. *J Med Genet.* 1991;28:181–5.
 39. Blüschke G, Nüsken KD, Schneider H. Prevalence and prevention of severe complications of hypohidrotic ectodermal dysplasia in infancy. *Early Hum Dev.* 2010;86:397–9.
 40. Schneider H, Hammersen J, Preisler-Adams S, Huttner K, Rascher W, Bohring A. Sweating ability and genotype in individuals with X-linked hypohidrotic ectodermal dysplasia. *J Med Genet.* 2011;48:426–32.
 41. Lexner MO, Bardow A, Bjorn-Jorgensen J, Hertz JM, Almer L, Kreiborg S. Anthropometric and cephalometric measurements in X-linked hypohidrotic ectodermal dysplasia. *Orthod Craniofac Res.* 2007;10:203–15.
 42. Sandhu K, Handa S, Kanwar AJ. Anhidrotic ectodermal dysplasia with palmoplantar keratoderma: an unusual presentation. *Int J Dermatol.* 2007;46:631–3.
 43. Clarke A, Phillips DI, Brown R, Harper PS. Clinical aspects of X-linked hypohidrotic ectodermal dysplasia. *Arch Dis Child.* 1987;62:989–96.
 44. Allali J, Roche O, Monnet D, Brezin A, Renard G, Dufier JL. Anhidrotic ectodermal dysplasia: «congenital ameibomia». *J Fr Ophtalmol.* 2007;30:525–8.
 45. Alcón Saez JJ, Elía Martínez MA, Elía Martínez I, Pont Colomer M, Lurbe Ferrer E. Amastia and athelia as an exceptional presentation of hypohidrotic ectodermal dysplasia in an adolescent female. *An Pediatr (Barc).* 2008;69:289–90.
 46. Cambiaghi S, Restano L, Pääkkönen K, Caputo R, Kere J. Clinical findings in mosaic carriers of hypohidrotic ectodermal dysplasia. *Arch Dermatol.* 2000;136:217–24.
 47. Freire-Maia N, Pinheiro M. Carrier detection in Christ-Siemens-Touraine syndrome (X-linked hypohidrotic ectodermal dysplasia). *Am J Hum Genet.* 1982;34:672–4.
 48. Kerr CB, Wells RS, Cooper KE. Gene effect in carriers of anhidrotic ectodermal dysplasia. *J Med Genet.* 1966;3:169–76.
 49. Pinheiro M, Freire-Maia N. Christ-Siemens-Touraine syndrome - a clinical and genetic analysis of a large Brazilian kindred: I. Affected females. *Am J Med Genet.* 1979;4:113–22.
 50. Munoz F, Lestringant G, Sybert V, Frydman M, Alswaini A, Frossard PM, et al. Definitive evidence for an autosomal recessive form of hypohidrotic ectodermal dysplasia clinically indistinguishable from the more common X-linked disorder. *Am J Hum Genet.* 1997;61:94–100.
 51. Tariq M, Wasif N, Ahmad W. A novel deletion mutation in the EDAR gene in a Pakistani family with autosomal recessive hypohidrotic ectodermal dysplasia. *Br J Dermatol.* 2007;157:207–9.
 52. Chassaing N, Bourthoumieu S, Cossee M, Calvas P, Vincent MC. Mutations in EDAR account for one-quarter of non-ED1-related hypohidrotic ectodermal dysplasia. *Hum Mutat.* 2006;27:255–9.
 53. Lind LK, Stecksén-Blicks C, Lejon K, Schmitt-Egenolf M. EDAR mutations in autosomal dominant hypohidrotic ectodermal dysplasia in two Swedish families. *BMC Med Genet.* 2006;7:80.
 54. Bibi N, Ahmad S, Ahmad W, Naeem M. Molecular genetic analysis of consanguineous Pakistani families with autosomal recessive hypohidrotic ectodermal dysplasia. *Australas J Dermatol.* 2011;52:37–42.

55. Gregoriou S, Rigopoulos D, Vergou T, Korfitis C, Menegakis G, Kontochristopoulos G. Should we consider hypohidrotic ectodermal dysplasia as a possible risk factor for malignant melanoma? *J Cutan Med Surg.* 2007;11:188–90.
56. Gaide O, Schneider P. Permanent correction of an inherited ectodermal dysplasia with recombinant EDA. *Nat Med.* 2003;9:614–8.
57. Casal ML, Lewis JR, Mauldin EA, Tardivel A, Ingold K, Favre M, et al. Significant correction of disease after postnatal administration of recombinant ectodysplasin A in canine X-linked ectodermal dysplasia. *Am J Hum Genet.* 2007;81:1050–6.
58. Mauldin EA, Gaide O, Schneider P, Casal ML. Neonatal treatment with recombinant ectodysplasin prevents respiratory disease in dogs with X-linked ectodermal dysplasia. *Am J Med Genet A.* 2009;149A:2045–9.
59. Smahi A, Courtois G, Vabres P, Yamaoka S, Heuertz S, Munich A, et al. Genomic rearrangement in NEMO impairs NF-kappaB activation and is a cause of incontinentia pigmenti. The International Incontinentia Pigmenti (IP) Consortium. *Nature.* 2000;405:466–72.
60. Smahi A, Courtois G, Rabia SH, Döffinger R, Bodemer C, Munnich A, et al. The NF-kappaB signalling pathway in human diseases: from incontinentia pigmenti to ectodermal dysplasias and immune-deficiency syndromes. *Hum Mol Genet.* 2000;11:2371–5.
61. Puel A, Picard C, Ku CL, Smahi A, Casanova JL. Inherited disorders of NF-kappaB-mediated immunity in man. *Curr Opin Immunol.* 2004;16:34–41.
62. Courtois G, Smahi A, Israël A. A NEMO/IKKgamma: linking NF-kappaB to human disease. *Trends Mol Med.* 2001;7:427–30.
63. Aradhya S, Nelson DL. NF-kappaB signaling and human disease. *Curr Opin Genet Dev.* 2001;11:300–6.
64. Rothwarf DM, Zandi E, Natoli G, Karin M. IKK-gamma is an essential regulatory subunit of the I kappa B kinase complex. *Nature.* 1998;395:297–300.
65. Bruskner AL. Incontinentia pigmenti: a window to the role of NF-kappaB function. *Semin Cutan Med Surg.* 2004;23:116–24.
66. Huang TT, Wuerzberger-Davis SM, Wu ZH, Miyamoto S. Sequential modification of NEMO/IKKgama by SUMO-1 and ubiquitin mediates NF-kappaB activation by genotoxic stress. *Cell.* 2003;115:565–76.
67. Ulvmar MH, Sur I, Mémet S, Toftgard R. Timed NF-kappaB inhibition in skin reveals dual independent effects on development of HED/EDA and chronic inflammation. *J Invest Dermatol.* 2009;129:2584–93.
68. Orange JS, Levy O, Geha RS. Human disease resulting from gene mutations that interfere with appropriate nuclear factor-kappaB activation. *Immunol Rev.* 2005;203:21–37.
69. Roberts CM, Angus JE, Leach IH, McDermott EM, Walker DA, Ravenscroft JC. A novel NEMO gene mutation causing osteopetrosis, lymphoedema, hypohidrotic ectodermal dysplasia and immunodeficiency (OL-HED-ID). *Eur J Pediatr.* 2010;169:1403–7.
70. Berlin AL, Paller AS, Chan LS. Incontinentia pigmenti: a review and update on the molecular basis of pathophysiology. *J Am Acad Dermatol.* 2002;47:169–87.
71. Jin DY, Jeang KT. Isolation of full-length cDNA and chromosomal localization of human NF-kappaB modulator NEMO to Xq28. *J Biomed Sci.* 1999;6:115–20.
72. Parrish JE, Scheuerle AE, Lewis RA, Levy ML, Nelson DL. Selection against mutant alleles in blood leukocytes is a consistent feature in incontinentia pigmenti type 2. *Hum Mol Genet.* 1996;5:1777–83.
73. Nelson DL. NEMO, NFkappaB signaling and incontinentia pigmenti. *Curr Opin Genet Dev.* 2006;16:282–8.
74. Buinauskaite E, Buinauskaite J, Kucinskiene V, Strazdiene D, Valiukeviciene S. Incontinentia pigmenti in a male with Klinefelter syndrome: a case report and review of the literature. *Pediatr Dermatol.* 2010;27:492–5.
75. Feito-Rodríguez M, García-Macarrón J, Bravo-Burguillos ER, Vera-Casaño A, de Lucas-Laguna R. Incontinentia pigmenti: three new cases that demonstrate it is not only a matter of women. *Actas Dermosifiliogr.* 2007;98:112–5.
76. Cohen PR. Incontinentia pigmenti: clinicopathologic characteristics and differential diagnosis. *Cutis.* 1994;54:161–6.
77. Zonana J, Elder ME, Schneider LC, Orlow SJ, Moss C, Golabi M, et al. A novel X-linked disorder of immune deficiency and hypohidrotic ectodermal dysplasia is allelic to incontinentia pigmenti and due to mutations in IKK-gamma (NEMO). *Am J Hum Genet.* 2000;67:1555–62.
78. Kosaki K, Shimasaki N, Fukushima H, Hara M, Ogata T, Matsuo N. Female patient showing hypohidrotic ectodermal dysplasia and immunodeficiency (HED-ID). *Am J Hum Genet.* 2001;69:664–5.
79. Martinez-Pomar N, Munoz-Saa I, Heine-Suner D, Martin A, Smahi A, Matamoros N. A new mutation in exon 7 of NEMO gene: late skewed X-chromosome inactivation in an incontinentia pigmenti females patient with immunodeficiency. *Hum Genet.* 2005;118:458–65.
80. Orange JS, Jain A, Ballas ZK, Schneider LC, Geha RS, Bonilla FA. The presentation and natural history of immunodeficiency caused by nuclear factor kappaB essential modulator mutation. *J Allergy Clin Immunol.* 2004;113:725–33.
81. Chang TT, Behshad R, Brodell RT, Gilliam AC. A male infant with anhidrotic ectodermal dysplasia/immunodeficiency accompanied by incontinentia pigmenti and a mutation in the NEMO pathway. *J Am Acad Dermatol.* 2008;58:316–20.
82. Pachlopnik Schmid JM, Junge SA, Hossle JP, Schneider EM, Roosnek E, Seger RA, et al. Transient hemophagocytosis with deficient cellular cytotoxicity, monoclonal immunoglobulin M gammopathy, increased T-cell numbers, and hypomorphic NEMO mutations. *Pediatrics.* 2006;117:1049–56.
83. Orstavik KH, Kristiansen M, Knudsen GP, Storhaug K, Vege A, Eiklid K, et al. Novel splicing mutation in the NEMO (IKK-gamma) gene with severe immunodeficiency and heterogeneity of X-chromosome inactivation. *Am J Med Genet A.* 2006;140:31–9.
84. Mancini AJ, Lawley LP, Uzel G. X-linked ectodermal dysplasia with immunodeficiency caused by NEMO mutation: early recognition and diagnosis. *Arch Dermatol.* 2008;144:342–6.
85. Jain A, Ma CA, Brown M, Cohen J, Strober W. Specific missense mutations in NEMO result in hyper-IgM syndrome with hypohidrotic ectodermal dysplasia. *Nat Immunol.* 2001;2:223–8.
86. Orange JS, Brodeur SR, Jain A, Bonilla FA, Schneider LC, Kretschmer R, et al. Deficient natural killer cell cytotoxicity in patients with IKK-gamma/NEMO mutations. *J Clin Invest.* 2002;109:1501–9.
87. Döffinger R, Smahi A, Bessia C, Geissmann F, Feinberg J, Durandy A, et al. X-linked anhidrotic ectodermal dysplasia with immunodeficiency is caused by impaired NF-kappaB signalling. *Nat Genet.* 2001;27:277–85.
88. McDonald DR, Mooster JL, Reddy M, Bawle E, Secord E, Geha RS. Heterozygous N-terminal deletion of I kappa Balpha results in functional nuclear factor kappaB haploinsufficiency, ectodermal dysplasia, and immune deficiency. *J Allergy Clin Immunol.* 2007;120:900–7.
89. Courtois G, Smahi A, Reichenbach J, Döffinger R, Cancrini C, Bonnet M, et al. A hypermorphic I kappa Balpha mutation is associated with autosomal dominant anhidrotic ectodermal dysplasia and T cell immunodeficiency. *J Clin Invest.* 2003;112:1108–15.
90. Ottenhoff TH, Verreck FA, Hoeve MA, van de Vosse E. Control of human host immunity to mycobacteria. *Tuberculosis (Edinb).* 2005;85:53–64.

91. Mansour S, Woffendin H, Mitton S, Jeffery I, Jakins T, Kenwrick S, et al. Incontinentia pigmenti in a surviving male is accompanied by hypohidrotic ectodermal dysplasia and recurrent infection. *Am J Med Genet.* 2001;99:172–7.
92. Hsu H, Lacey DL, Dunstan CR, Solovyev I, Colombero A, Timms E, et al. Tumor necrosis factor receptor family member RANK mediates osteoclast differentiation and activation induced by osteoprotegerin ligand. *Proc Natl Acad Sci USA.* 1999;96:3540–5.
93. Karkkainen MJ, Ferrell RE, Lawrence EC, Kimak MA, Levinson KL, McTigue MA, et al. Missense mutations interfere with VEGFR-3 signalling in primary lymphoedema. *Nat Genet.* 2000;25:153–9.
94. Kenwrick S, Woffendin H, Jakins T, Shuttleworth SG, Mayer E, Greenhalgh L. Survival of male patients with incontinentia pigmenti carrying a lethal mutation can be explained by somatic mosaicism or Klinefelter syndrome. *Am J Hum Genet.* 2001;69:1210–7.
95. Fish JD, Duerst E, Gelfand EW, Orange JS, Bunin N. Challenges in the use of allogeneic hematopoietic SCT for ectodermal dysplasia with immune deficiency. *Bone Marrow Transplant.* 2009;43:217–21.
96. Fete M, van Bokhoven H, Clements SE, Mc Keon F, Roop DR, Koster MI, et al. International Research Symposium on Ankyloblepharon-Ectodermal Defects-Cleft Lip/Palate (AEC) syndrome. *Am J Med Genet A.* 2009;149A:1885–93.
97. King EK, Weinberg WC. P63: defining roles in morphogenesis, homeostasis and neoplasia of the epidermis. *Mol Carcinog.* 2007;46:716–24.
98. Koster MI, Roop DR. The role of p63 in development and differentiation of the epidermis. *J Dermatol Sci.* 2004;34:3–9.
99. Guazzarotti L, Caprio C, Rinne TK, Bosoni M, Pattarino G, Mauri S, et al. Limb-mammary syndrome (LMS) associated with internal female genitalia dysgenesis: a new genotype/phenotype correlation? *Am J Med Genet.* 2008;146A:2001–4.
100. Shimomura Y, Wajid M, Shapiro L, Christiano AM. P-cadherin is a p63 target gene with a crucial role in the developing human limb bud and hair follicle. *Development.* 2008;135:743–53.
101. Rinne T, Hamel B, van Bokhoven H, Brunner HG. Pattern of p63 mutations and their phenotypes-update. *Am J Med Genet A.* 2006;140A:1396–406.
102. Barrow LL, van Bokhoven H, Daac-Hirsch S, Andersen T, van Beersum SE, Gorlin R, et al. Analysis of the p63 gene in classical EEC syndrome, related syndromes, and non-syndromic orofacial clefts. *J Med Genet.* 2002;39:559–66.
103. Roelfsema NM, Cobben JM. The EEC syndrome: a literature study. *Clin Dysmorphol.* 1996;5:115–27.
104. Pierre-Louis M, Byer-Parsons T, Burkhart CN, Morrell DS. Perioral lesions in ectrodactyly, ectodermal dysplasia, clefting syndrome. *Pediatr Dermatol.* 2010;27:658–60.
105. León-Mateos A, Monteagudo B, Rodríguez L. Patient with «lobster-claw» hands and feet: ectrodactyly-ectodermal dysplasia-clefting syndrome. *Actas Dermosifiliogr.* 2008;99:822–3.
106. Brunner HG, Hamel BC, Van Bokhoven H. The p63 gene in EEC and other syndromes. *J Med Genet.* 2002;39:377–81.
107. Chuangsuwanich T, Sunsaneevithayakul P, Muangsomboon K, Limwongse C. Ectrodactyly-ectodermal dysplasia-clefting (EEC) syndrome presenting with a large nephrogenic cyst, severe oligohydramnios and hydrops fetalis: a case report and review of the literature. *Prenat Diagn.* 2005;25:210–5.
108. Dalben GS, Cursino HA, Barbosa BA, Costa B, Consolaro A. White sponge nevus in a patient with EEC syndrome. *Dermatol Online J.* 2010;16:7.
109. Johnson SE, Tatum SA, Thomson LL. Pierre Robin sequence in a patient with ectrodactyly-ectodermal dysplasia-clefting syndrome: a case report and review of the literature. *Int J Pediatr Otorrhinolaryngol.* 2002;66:309–13.
110. Hay RJ, Wells RS. The syndrome of ankyloblepharon, ectodermal defects and cleft lip and palate: An autosomal dominant condition. *Br J Dermatol.* 1976;94:277–89.
111. Julapalli MR, Scher RK, Syber VP, Siegfried EC, Bree AF. Dermatologic findings of ankyloblepharon-ectodermal defects-cleft lip/palate (AEC) syndrome. *Am J Med Genet A.* 2009;149A:1900–6.
112. Dishop M, Bree A, Hicks M. Pathologic features of skin and hair in ankyloblepharon-ectodermal defects-cleft lip/palate (AEC) syndrome. *Am J Med Genet A.* 2009;149A:1935–41.
113. Hofman D, Moore K, Cooper R, Eagle M, Cooper S. Use of topical steroids on chronic leg ulcers. *J Wound Care.* 2007;16:227–30.
114. Shwayder TA, Lane AT, Miller ME. Hay-Wells syndrome. *Pediatr Dermatol.* 1986;3:399–402.
115. Siegfried E, Bree A, Fete M, Sybert VP. Skin erosions and wound healing in ankyloblepharon-ectodermal defect-cleft lip and/or palate. *Arch Dermatol.* 2005;141:1591–4.
116. Avitan-Hersh E, Indelman M, Bergman R, Sprecher E. ADULT syndrome caused by a mutation previously associated with EEC syndrome. *Pediatr Dermatol.* 2010;27:643–5.
117. Reisler TT, Patton MA, Meagher PP. Further phenotypic and genetic variation in ADULT syndrome. *Am J Med Genet A.* 2006;140A:2495–500.
118. Amiel J, Bougeard G, Francannet C, Ranclin V, Munnich A, Lyonnet S, et al. TP63 mutation in ADULT syndrome. *Eur J Hum Genet.* 2001;9:642–5.
119. Van Bokhoven H, Hamel BC, Bamshad M, Sangiorgi E, Gurrieri F, Duijff PH, et al. P63 gene mutations in EEC syndrome, limb-mammary syndrome, and isolated split hand-foot malformation suggest a genotype-phenotype correlation. *Am H Hum Genet.* 2001;69:481–92.
120. Van Bokhoven H, Jung M, Smits AP, van Beersum S, Rüschen-dorf F, van Steensel M, et al. Limb mammary syndrome: a new genetic disorder with mammary hypoplasia, ectrodactyly, and other hand/foot anomalies maps to human chromosome 3q27. *Am J Hum Genet.* 1999;64:538–46.
121. Clements SE, Techanukul T, Holden ST, Mellerio JE, Dorkins H, Escande F, et al. Rapp-Hodgkin and Hay-Wells ectodermal dysplasia syndromes represent a variable spectrum of the same genetic disorder. *Br J Dermatol.* 2010;163:624–9.
122. Kannu P, Savarirayan R, Ozoemena L, White SM, McGrath JA. Rapp-Hodgkin ectodermal dysplasia syndrome: the clinical and molecular overlap with Hay-Wells syndrome. *Am J Med Genet A.* 2006;140:887–91.
123. Bertola DR, Kim CA, Albano LM, Scheffer H, Meijer R, van Bokhoven H. Molecular evidence that AEC syndrome and Rapp-Hodgkin syndrome are variable expression of a single genetic disorder. *Clin Genet.* 2004;66:79–80.
124. Dianzani I, Garelli E, Gustavsson P, Carando A, Gustafsson B, Dahl N, et al. Rapp-Hodgkin and AEC syndromes due to a new frameshift mutation in the TP63 gene. *J Med Genet.* 2003;40:e133.
125. Steele JA, Hansen H, Arn P, Kwong PC. Spectrum of phenotypic manifestations from a single point mutation of the p63 gene, including new cutaneous and immunologic findings. *Pediatr Dermatol.* 2005;22:415–9.
126. Cambiaghi S, Tadini G, Barbareschi M, Menni S, Caputo R. Rapp-Hodgkin and AEC syndrome: are they the same entity? *Br J Dermatol.* 1994;130:97–101.
127. Prontera P, Escande F, Cocchi G, Danti E, Martini A, Sensi A. An intermediate phenotype between Hay-Wells and Rapp-Hodgkin syndromes in a patient with a novel P63 mutation: confirmation of a variable phenotypic spectrum with a common aetiology. *Genet Couns.* 2008;19:397–402.

128. Price JA, Bowden DW, Wright JT, Pettenati MJ, Hart TC. Identification of a mutation in DLX3 associated with tricho-dento-osseous (TDO) syndrome. *Hum Molec Genet.* 1998;7:563–9.
129. Wright JT, Hong SP, Simmons D, Daly B, Uebelhart D, Luder HU. DLX3 c.561_562delCT mutation causes attenuated phenotype of tricho-dento-osseous syndrome. *Am J Med Genet.* 2008;146A:343–9.
130. Wright JT, Kula K, Hall K, Simmons JH, Hart TC. Analysis of the tricho-dento-osseous syndrome genotype and phenotype. *Am J Med Genet.* 1997;72:197–204.
131. Jumlongras D, Bei M, Stimson JM, Wang WF, DePalma SR, Seidman CE, et al. A nonsense mutation in MSX1 causes Witkop syndrome. *Am J Hum Genet.* 2001;69:67–74.
132. Memarpour M, Shafiei F. Witkop tooth and nail syndrome: a report of three cases in a family. *Pediatr Dermatol.* 2011;28:281–5.
133. Ye X, Song G, Fan M, Shi L, Jabs EW, Huang S, et al. A novel heterozygous deletion in the EVC2 gene causes Weyers acrofacial dysostosis. *Hum Genet.* 2006;119:199–205.
134. Ruiz-Perez VL, Ide SE, Strom TM, Lorenz B, Wilson D, Woods K, et al. Mutations in a new gene in Ellis-van Creveld syndrome and Weyers acrofacial dysostosis. *Nat Genet.* 2000;24:283–6.
135. Ruiz-Perez VL, Tompson SW, Blair HJ, Espinoza-Valdez C, Lapunzina P, Silva EO, et al. Mutations in two nonhomologous genes in a head-to-head configuration cause Ellis-van Creveld syndrome. *Am J Hum Genet.* 2003;72:728–32.
136. Tompson SW, Ruiz-Pérez VL, Blair HJ, Barton S, Navarro V, Robson JL, et al. Sequencing EVC and EVC2 identifies mutation in two-thirds of Ellis-van Creveld syndrome patients. *Hum Genet.* 2007;120:663–70.
137. Ruiz-Pérez VL, Goodship JA. Ellis van-Creveld Syndrome and Weyers acrofacial dysostosis are caused by cilia-mediated diminished response to hedgehog ligands. *Am J Med Genet C Semin Med Genet.* 2009;151C:341–51.
138. Cahuana A, Palma C, Gonzáles W, Geán E. Oral manifestations in Ellis-van Creveld syndrome: report of five cases. *Pediatr Dent.* 2004;26:277–82.
139. Lamartine J, Muñoz Essenfelder G, Kibar Z, Lanneluc I, Calouet E, Laoudj D, et al. Mutations in GJB6 cause hidrotic ectodermal dysplasia. *Nat Genet.* 2000;2:142–4.
140. Jan AY, Amin S, Ratajczak P, Richard G, Sybert P. Genetic heterogeneity of KID syndrome: identification of a Cx30 gene (GJB6) mutation in a patient with KID syndrome and congenital atrichia. *J Invest Dermatol.* 2004;122:1108–13.
141. Smith FJ, Morley SM, McLean WH. A novel connexin 30 mutation in Clouston syndrome. *J Invest Dermatol.* 2002;118:530–2.
142. Essenfelder GM, Bruzzone R, Lamartine J, Charollais A, Blanchet-Bardon C, Barbe MT, et al. Connexin30 mutations responsible for hidrotic ectodermal dysplasia cause abnormal hemichannel activity. *Hum Mol Genet.* 2004;13:1703–14.
143. Wilkey WD, Stevenson GH. Family with inherited ectodermal dystrophy. *Can Med Assoc J.* 1945;53:226–30.
144. Sriver CR, Solomons CC, Davies E, Williams M, Bolton J. A molecular abnormality of keratin in ectodermal dysplasia. *J Pediatr.* 1965;67:946.
145. Gagnon CA, Berg SZ, Moeschler JB. Clouston hydrotic ectodermal dysplasia: report of a large New England family. *Am J Hum Genet.* 1989;45:121.
146. Van Steensel MA, Jonkman MF, van Geel M, Steiljen PM, McLean WH, Smith FJ. Clouston syndrome can mimic pachyonychia congenital. *J Invest Dermatol.* 2003;121:1035–8.
147. Tan E, Tay YK. What syndrome is this? Hidrotic ectodermal dysplasia (Clouston syndrome). *Pediatr Dermatol.* 2000;17:65–7.
148. Mascaro JM. Considérations sur les tumeurs fibro-épithéliales: le syringofibroadénome eccrine. *Ann Dermatol Syphil.* 1963;90:146–53.
149. Poonawalla T, Xia L, Patten S, Stratman EJ. Clouston syndrome and eccrine syringofibroadenomas. *Am J Dermatopathol.* 2009;31:157–61.
150. Jain K, Jain VK, Aggarwal K, Bansal A. Clouston syndrome associated with severe congenital pseudo-ainhum. *Pediatr Dermatol.* 2007;24:342–4.
151. Zlotogora J, Zilberman Y, Tenenbaum A, Wexler MR. Cleft lip and palate, pili torti, malformed ear, partial syndactyly of fingers and toes, and mental retardation: a new syndrome? *J Med Genet.* 1987;24:291–3.
152. Brancati F, Fortugno P, Bottillo I, Lopez M, Josselin E, Boudghene-Stambouli O, et al. Mutations in PVRL4 encoding cell adhesion molecule nectin-4, cause ectodermal dysplasia-syndactyly syndrome. *Am J Hum Genet.* 2010;87:265–73.
153. Matsushima H, Utani A, Endo H, Matsuura H, Kakuta M, Nakamura Y, et al. The expression of nectin-1alpha in normal human skin and various skin tumours. *Br J Dermatol.* 2003;148:755–62.
154. Sözen MA, Suzuki K, Tolarova MM, Bustos T, Fernández Iglesias JE, Spritz RA. Mutation of PVRL1 is associated with sporadic, non-syndromic cleft lip/palate in northern Venezuela. *Nature Genet.* 2001;29:141–2.
155. Rodini ES, Richieri-Costa A. Autosomal recessive ectodermal dysplasia, cleft lip/palate, mental retardation, and syndactyly: the Zlotogora-Ogur syndrome. *Am J Med Genet.* 1990;36:473–6.
156. Zlotogora J. Syndactyly, ectodermal dysplasia, and cleft lip/palate. *J Med Genet.* 1994;31:957–9.
157. Fine JD, Eady RA, Bauer EA, Bauer JW, Bruckner-Tuderman L, Heagerty A, et al. The classification of inherited epidermolysis bullosa (EB): Report of the Third International Consensus Meeting on Diagnosis and Classification of EB. *J Am Acad Dermatol.* 2008;58:931–50.
158. Ersoy-Evans S, Erkin G, Fassih H, Chan I, Paller AS, Sürücü S, et al. Ectodermal dysplasia-skin fragility syndrome resulting from a new homozygous mutation, 888delC, in the desmosomal protein plakophilin 1. *J Am Acad Dermatol.* 2006;55:157–61.
159. Bergman R, Sprecher E. Histopathological and ultrastructural study of ectodermal dysplasia/skin fragility syndrome. *Am J Dermatopathol.* 2005;27:333–8.
160. Albrechtsen B, Svendsen IB. Hypotrichosis, syndactyly, and retinal degeneration in two siblings. *Acta Derm Venereol.* 1956;36:96–101.
161. Kjaer KW, Hansen L, Schwabe GC, Marques-de-Faria AP, Eiberg H, Mundlos S, et al. Distinct CDH3 mutations cause ectodermal dysplasia, ectrodactyly, macular dystrophy (EEM syndrome). *J Med Genet.* 2005;42:292–8.
162. Senecky Y, Halpern GJ, Invar D, Attias J, Shohat M. Ectodermal dysplasia, ectrodactyly and macular dystrophy (EEM syndrome) in siblings. *Am J Med Genet.* 2001;101:195–7.
163. Adaimy L, Chouery E, Megarbane H, Mrouech S, Delague V, Nicolas E, et al. Mutations in WNT10A is associated with an autosomal recessive ectodermal dysplasia: the odonto-onycho-dermal dysplasia. *Am J Hum Genet.* 2007;81:821–8.
164. Nawaz S, Klar J, Wajid M, Aslam M, Tariq M, Schuster J, et al. WNT10A missense mutation associated with a complete odonto-onycho-dermal dysplasia syndrome. *Eur J Hum Genet.* 2009;17:1600–5.
165. Kantaputra P, Sripathomsawat W. WNT10A and isolated hypodontia. *Am J Med Genet A.* 2011;15:1119–22.
166. Zirbel GM, Ruttum MS, Post AC, Esterly NB. Odonto-onycho-dermal dysplasia. *Br J Dermatol.* 1995;133:797–800.
167. Adams BB. Odonto-onycho-dermal dysplasia syndrome. *J Am Acad Dermatol.* 2007;57:732–3.
168. Fadhil M, Ghabra TA, Deeb M, Der Kaloustian VM. Odontoonycho-dermal dysplasia: a previously apparently undescribed ectodermal dysplasia. *Am J Med Genet.* 1983;14:335–46.

169. Arnold WP, Merkx MAW, Steijlen PM. Variant of odontoonycho-dermal dysplasia? *Am J Med Genet.* 1995;59:242–4.
170. Mégarbané H, Haddad M, Delague V, Renoux J, Boehm N, Mégabarné A. Further delineation of the odonto-onycho-dermal dysplasia syndrome. *Am J Med Genet.* 2004;129A: 193–7.
171. Bohring A, Stamm T, Spaich C, Haase C, Spree K, Hehr U, et al. WNT10A mutations are a frequent cause of a broad spectrum of ectodermal dysplasias with sex-biased manifestation pattern in heterozygotes. *Am J Hum Genet.* 2009;85: 97–105.