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REVIEW

Cutaneous Squamous Cell Carcinoma: Defining the High-Risk Variant☆

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Abstract With a lifetime incidence of approximately 10% in the general population, cutaneous squamous cell carcinoma (CSCC) is the second most common type of nonmelanoma skin cancer.

Most CSCCs are benign and can be completely eradicated by surgery or other dermatological procedures. There is, however, a subgroup associated with an increased likelihood of lymph node metastases and, therefore, with high morbidity and mortality.

This article analyzes the various factors that define aggressive CSCC. We propose a method for defining high-risk SCC on the basis of a series of major and minor criteria. This method will allow better prognostic evaluation and enable personalized management of patients with high-risk SCC, possibly leading to improved overall survival.

PALABRAS CLAVE
Carcinoma epidermoide cutáneo;
Carcinoma epidermoide cutáneo de alto riesgo

Carcinoma epidermoide cutáneo: definendo la variante de alto riesgo

Resumen El carcinoma epidermoide cutáneo, con una incidencia en la población general de aproximadamente un 10% a lo largo de la vida, es la segunda neoplasia más frecuente dentro del grupo del cáncer cutáneo no melanoma.

La mayoría de los carcinomas epidermoides cutáneos muestran un comportamiento benigno y pueden ser completamente erradicados mediante cirugía y otros procedimientos dermatológicos. Sin embargo, existe un subgrupo de esta entidad que se asocia con una mayor capacidad de desarrollar metástasis nodal y, por tanto, con una elevada morbimortalidad.

En el presente artículo se analizan los diferentes factores que definen al carcinoma epidermoide cutáneo de comportamiento agresivo. Proponemos un método de definición del

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carcinoma epidermoide de alto riesgo basado en el establecimiento de una serie de criterios mayores y menores. Este hecho supondrá una mejor evaluación pronóstica y un manejo personalizado de este grupo de enfermos, que puede resultar en un aumento de la supervivencia global.

Introduction

Cutaneous squamous cell carcinoma (CSCC) has a lifetime incidence of between 7% and 11%. It accounts for 20% to 25% of all nonmelanoma skin cancers, and is second only to basal cell carcinoma in terms of prevalence.[1–5]

Most CSCCs are benign and can be completely eradicated by surgery and other dermatological procedures. Accordingly, 5-year survival rates after surgical excision are in excess of 90%,[5] and mortality is around 1%.[6] Nevertheless, there is a subgroup of CSCC associated with a higher frequency of lymph node metastasis and with high morbidity and mortality (Figs. 1-6).[7–10]

The main aim of this article is to accurately define this subgroup of CSCC, which is known as high-risk CSCC. A clear definition will help to refine prognosis and improve the individualized care of patients with high-risk CSCC.

Definition of High-Risk CSCC

In recent years, several authors have focused their research on analyzing differences between nonmetastatic and metastatic CSCC. The ultimate aim of such research was to predict which forms of CSCC are associated with an increased risk of locoregional and/or distant complications in order to be able to intervene promptly in patients at risk.

The findings led to the concept of high-risk CSCC, which is defined as a squamous cell carcinoma lesion, clinically staged as N0, that extends through the basement membrane and is associated with a high risk of subclinical metastasis. CSCCs that do not this meet this definition are classified as low-risk.

Defining Features of High-Risk CSCC

The factors that define high-risk CSCC can be divided into 3 subgroups: clinical factors, histologic factors, and molecular factors.

Clinical Factors

Personal History

Genetic Disorders. Patients with genetic disorders associated with an increased risk of CSCC typically develop tumors with a high risk of malignant transformation. Examples of these disorders are xeroderma pigmentosum, epidermodysplasia verruciformis, oculocutaneous albinism, dyskeratosis congenita, and recessive dystrophic epidermolysis bullosa. This last condition is associated with the highest mortality in patients with concomitant CSCC, with 5-year survival rates of just 80% after a diagnosis of the skin tumor.[4,11–13]

CSCC Arising at the Site of a Pre-existing Lesion. CSCCs that arise at the site of chronic skin damage, such as scars, slow-growing ulcers, burn sites, and chronic radiation dermatitis, have an increased risk of metastatic spread. This risk appears to be associated with a reduction in E-cadherin levels, which would favor the spread of atypical keratinocytes through the epidermis and into the dermis.[14]

Immunosuppression and Transplantation. Immune status is a predictor of prognosis in many neoplastic conditions. Immune system alterations, for example, play an important role in the development of skin cancers, such as Merkel cell carcinoma.[15]

Patients who undergo solid organ transplantation (SOT) have a 65-fold higher risk of developing CSCC than the general population; furthermore, CSCC is the most common nonmelanoma skin cancer in SOT recipients and is 3 times more common than basal cell carcinoma.[16]

Recurrence rates, locoregional metastasis, and survival in transplant recipients with CSCC vary depending on the organ transplanted. In the field of SOT, heart transplantation is considered to carry the highest risk of CSCC and its high-risk variant,[16] followed by, in decreasing order, lung, kidney, and liver transplantation. In the case of hematologic malignancies, the highest risk of both types of CSCC has been observed in patients with chronic lymphatic leukemia and small lymphocytic lymphoma.[17]

The cumulative incidence of CSCC increases progressively with the duration of immunosuppression, with observed rates of 7% after a year, 45% after 11 years, and 70% after 20 years. Furthermore, up to 66% of transplant recipients have been reported to develop a second CSCC after the first squamous cell carcinoma.[18]

CSCC recurs more frequently in immunosuppressed than in immunocompetent individuals (39% vs 15% in 5 years of follow-up),[19] and mortality is also higher (5% in transplant recipients vs 1% in immunocompetent individuals).[20] Organ transplant recipients with metastatic CSCC have a 3-year survival rate of 56%, which is similar to rates reported for patients with noncutaneous SCC of the head and neck.[20]

The fact that metastatic CSCC often has similar clinical characteristics (horizontal diameter of < 2 cm and vertical histologic thickness of < 2 mm) and favorable outcomes in immunosuppressed and immunocompetent individuals

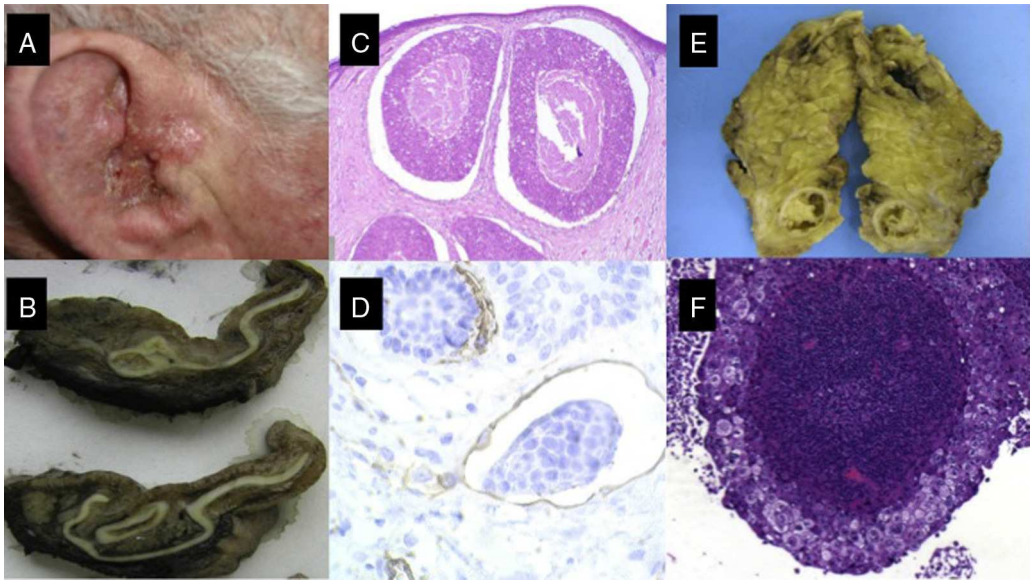

Figure 1 A, Cutaneous squamous cell carcinoma on the external ear of a 70-year-old patient. B, Fleshy tumor invading the external auditory canal. C and D, Histology showing a poorly differentiated squamous cell tumor with lymphovascular invasion (C, hematoxylin-eosin, original magnification x40; D, Immunostaining with CD31, original magnification x100); E, Parotid gland with nodular tumor. F, The parotid tumor is formed by a proliferation of poorly differentiated squamous cells in a pattern similar to that of the primary tumor (original magnification, hematoxylin-eosin, x40).

suggests that as yet unknown molecular alterations have a role in the high malignant potential of CSCC in immunosuppressed individuals.

Human Immunodeficiency Virus Infection. Human immunodeficiency virus (HIV) infection, regardless of disease stage or immune status, appears to be a marker of poor prognosis in CSCC. This possibility was suggested by Nguyen et al.[21] in a retrospective study of 10 consecutively recruited HIV-positive patients aged between 31 and 54 years with high-risk CSCC. Five of the 10 patients died within 7 years of the initial diagnosis, and local recurrence, metastasis, and survival were not correlated with the number of opportunistic infections or with $CD4^+$ T cell count.

Clinical Characteristics of CSCC

Lesion Size

The size of primary lesions in CSCC has been described by many authors as being an important predictor of lymph node metastasis.[12,22–25]

Data from the first prospective study of CSCC, involving over 1000 patients, showed that horizontal tumor size was

Figure 2 A and B, Two patients, aged 67 and 69 years, respectively, treated for high-risk squamous cell carcinoma on the scalp (A) and the temple (B). The 2 patients developed invasion of the upper cervical lymph nodes in the second year of follow-up.

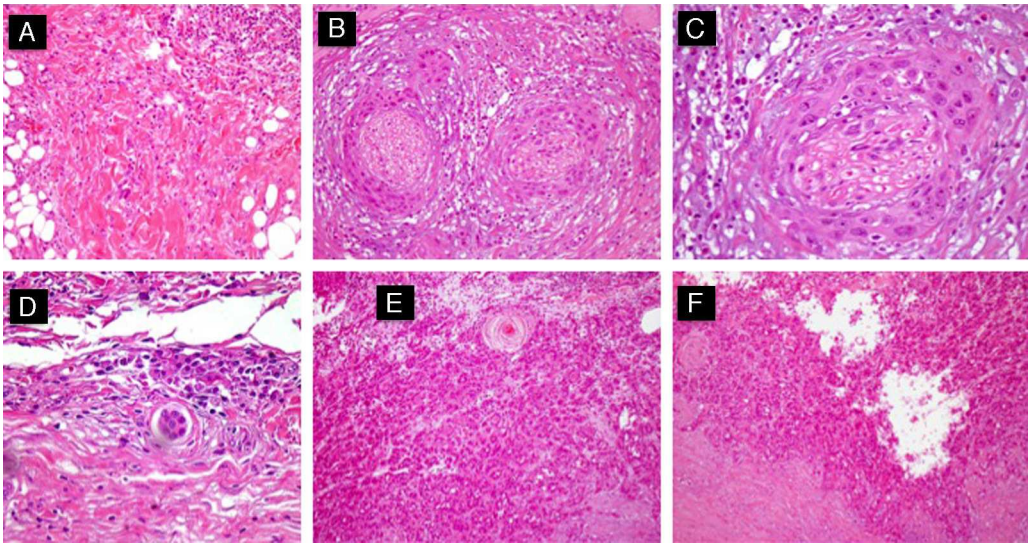

Figure 3 Different histologic subtypes associated with high-risk cutaneous squamous cell carcinoma. A, Isolated-cell pattern (hematoxylin-eosin, original magnification x40). B and C, Squamous cell carcinoma with perineural invasion (nerves > 0.1 mm) (hematoxylin-eosin, original magnification x40 [B] and x100 [C]). D, Squamous cell carcinoma with marked lymphovascular invasion (hematoxylin-eosin, original magnification x100). E, Adenoid squamous cell carcinoma (hematoxylin-eosin, original magnification x40). F, Acantholytic squamous cell carcinoma (hematoxylin-eosin, original magnification x40).

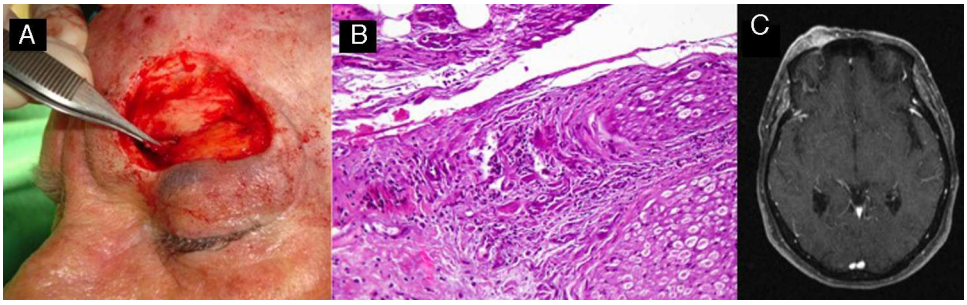

Figure 4 A, Perineural invasion detected incidentally during surgery. B, Histology showing invasion of the nerve trunk (diameter, > 0.1 mm) by atypical squamous cells (hematoxylin-eosin, original magnification x40). C, Magnetic resonance image showing intracranial invasion of the tumor along the nerve pathway.

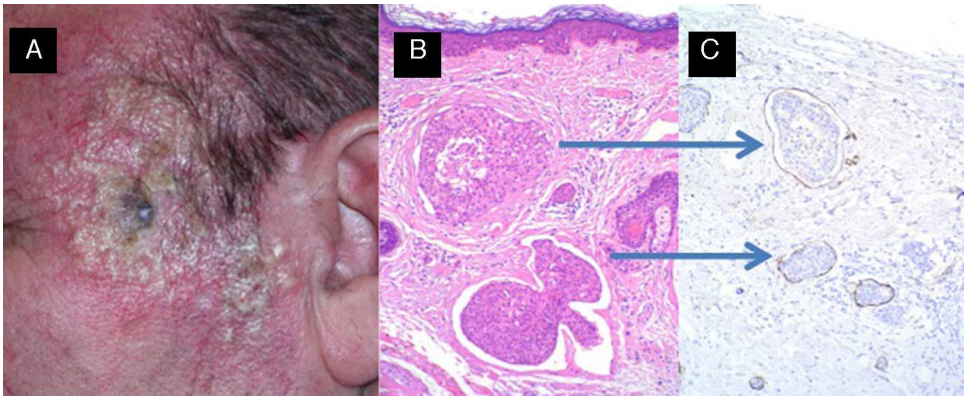

Figure 5 Carcinomatous lymphangitis secondary to squamous cell carcinoma previously excised from the left temple. A, Multiple papulous vesicles on the left temple. B and C, Invasion of lymph vessels by atypical squamous cells (B, Hematoxylin-eosin, original magnification x40; C, Immunostaining with D2-40, original magnification x40).

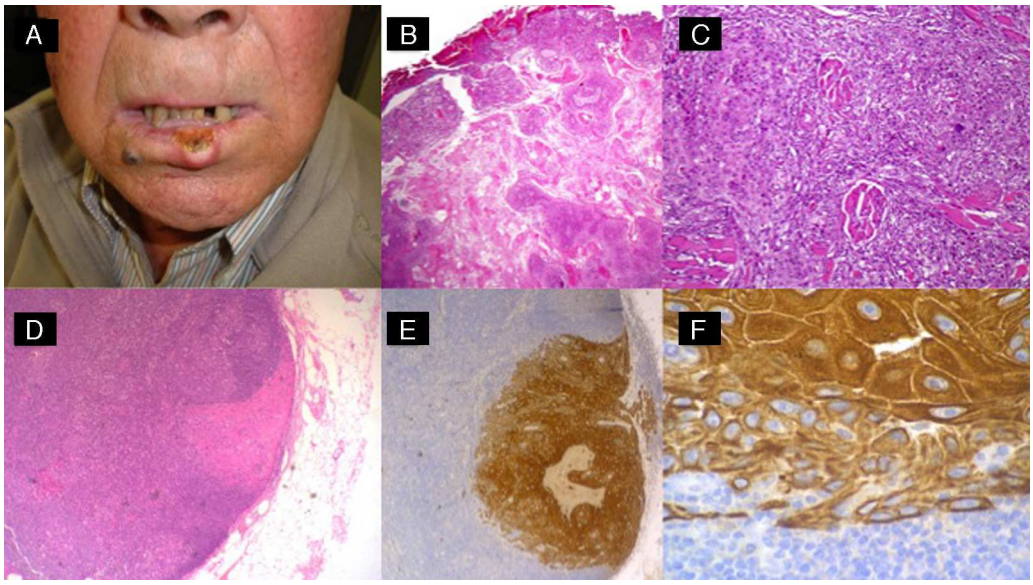

Figure 6 High-risk squamous cell carcinoma studied by sentinel lymph node biopsy. A, Squamous cell carcinoma of the lower lip in a 65-year-old man. B and C, Atypical squamous cell proliferation with acantholysis and perineural invasion. D-F, Invasion of sentinel lymph node by atypical squamous cells (hematoxylin-eosin, original magnification x40 [D] and x100 [E]; immunostaining with pankeratin, original magnification x100 [F]).

an independent risk factor for metastasis. Specifically, the risk of metastatic spread was 0.01% in lesions measuring 2 cm or less in diameter and 10% in larger lesions. In the second group, 7% of patients with a tumor size of between 2 and 5 cm developed metastasis compared with 20% of those with a tumor size of over 5 cm.[26–29]

Based on our experience and on data from the literature, we consider a horizontal size of 2 cm to be the cutoff for increased risk of lymph node metastasis in CSCC. A smaller size, by contrast, would exert a protective effect, meaning that there would not be a risk of distant metastasis in immunocompetent patients with a tumor diameter of less than 2 cm.

Lesion Site

Lesion sites with the highest incidence (20%-30%) of metastatic CSCC are the external ear (Fig. 1) and the nonglabrous lip (Fig. 6).[19,30]

Middle-risk sites include the scalp (mainly the temple), the perineal and genital areas, and acral sites (hands and feet).[6,31] It is also important to consider that areas not exposed to sunlight, such as the perineum, the sacral region, and the soles of the feet, have a proportionally higher rate of metastasis than chronically sun-exposed areas.[32]

Recurrence

Tumor recurrence tends to be associated with poor prognosis in skin cancers.

Comparative analysis of lymph node metastasis in recurrent CSCC (15%) and nonrecurrent CSCC (2%) ($P < .001$) has led to the conclusion that tumor recurrence is an important risk factor in CSCC.[33]

Clayman et al.[6] observed an association between CSCC recurrence and tumor size, and reported that large tumors were associated with a significantly higher rate of recurrence (2.4 vs 1.5 cm, $P < .0001$). They also found recurrent lesions to be associated with a higher rate of perineural invasion (PNI) (24% vs 10%), lymphovascular invasion (17% vs 8%), and subcutaneous tissue invasion (30% vs 10%).[6]

Recurrence has also been significantly associated with positive margins in surgically excised CSCC, with recurrent tumors—and consequently increased risk of metastasis—observed in up to 50% of patients with positive margins.[33]

Human Papillomavirus Infection

β-Human papillomaviruses (HPVs) are the most common type of HPVs involved in CSCC. Numerous studies have demonstrated a relationship between β-HPVs and CSCC, above all in immunosuppressed patients, although these viruses may also act as a cofactor with UV radiation in immunocompetent patients. Nevertheless, because β-HPVs appear to be involved in the etiology and pathogenesis of CSCC and not in metastatic spread, they are not considered prognostic factors.α-HPVs associated with CSCC of the genital region, the head and the neck, and acral sites might be associated with a higher risk of metastasis as they alter regulatory mechanisms, such as p53 and the retinoblastoma gene/*p16*. Most studies of *p16* in CSCC have reported loss of *p16* expression to be associated with the transformation of in situ CSCC to invasive CSCC but not with a higher risk of metastatic spread. As mentioned previously, only *p16*-positive cases associated with the presence of α-HPVs would carry an increased risk of metastasis.[34–39]

Histologic Features of CSCC

Tumor Thickness and Clark Level

Tumor Thickness. Tumor thickness is currently considered to be the most important independent predictor of metastasis in CSCC, with greater thickness associated with higher risk.[26,32]

Based on data from the largest prospective series of CSCC to date, conducted by Brantsch et al.,[26] CSCC can be divided into 3 risk groups (low-risk, middle-risk, and high-risk) based on tumor thickness. Patients in the low-risk group have tumors with a thickness of 2 mm or less, and have virtually no risk of distant metastasis. Those in the middle-risk group have a tumor thickness of between 2 and 6 mm, which is associated with a 4% increased risk of metastasis in 5 years of follow-up. Finally, those in the high-risk group have tumors with a thickness of 6 mm or more and a 16% increased risk of metastasis.

Based on data from later studies and on our own experience, we believe that a cutoff a 4 mm provides the best sensitivity for separating low-risk CSCC from CSCC with a high risk of metastatic spread. Tumors with a thickness of less than 2 mm, by contrast, would be associated with virtually no risk of distant disease.

Degree of Tumor Differentiation

Degree of tumor differentiation is another important prognostic factor in CSCC and other neoplastic diseases.

In a study of 571 patients with CSCC, a significant difference was observed for the rate of metastasis between lesions with a high degree of differentiation and other lesions (17% vs 4%, $P = .004$),[22] and in another study involving a large number of patients, high-grade CSSS was associated with a greater risk of malignant transformation than other types of CSCC (44% vs 5%, $P < .01$).[12]

Tumor differentiation is also associated with an increased risk of early recurrence. Poorly differentiated CSCCs have a 2.9-fold increased risk of distant metastasis and death compared with well-differentiated CSCCs, although well-differentiated tumors may also be associated with the development of advanced disease.[12,40]

Finally certain histologic subtypes of CSCC (acantholytic, adenoid, isolated cell pattern) should be considered as a risk factor in combination with tumor differentiation (Fig. 3).

Histologically Positive Surgical Margins

Incomplete tumor excision—and consequently—disease persistence, is a predictor of poor prognosis in CSCC. Disease, and with it an increased risk of metastasis, recurs in up to 50% of patients with histologically positive margins following surgical excision.[33] Recurrence following tumor excision appears to be related to a risk of subclinical tumor progression, which would, in turn, favor metastasis.[41]

The decision to take a watch and wait approach with patients with incompletely excised CSCCs, i.e., with a pathology report showing the involvement of 1 or more margins, should be weighed up carefully given the high rate of lymph node disease in recurrent CSCC. Several studies have shown a history of disease recurrence in between 45% and 51% of patients with CSCC and lymph node involvement.[12,22,42,43]

Consequently, all patients with CSCC should undergo surgery until disease-free margins are achieved,[44] and if this is not possible, other treatments, mainly radiation therapy, should be considered.

Perineural Invasion

PNI occurs in approximately 5% to 10% of patients with CSCC and is usually detected as an incidental finding.[6,45] Nonetheless, histologic evidence of PNI appears to be associated with a significant increase in disease recurrence and distant metastasis rates.[6,45] A study performed at the Anderson clinic in Texas, United States, showed that compared with CSCC patients without PNI, those with PNI had a significantly increased frequency of regional metastasis (35% vs 15%, $P < .005$) and distant metastasis (15% vs 3.3%, $P < .005$).[46]PNI is important not only because of the risk of locoregional spread, but also because of disease caused by perineural spread through the cranial nerves, mostly the facial and the trigeminal nerves (Fig. 4) and because PNI is associated with worse 3-year survival in CSCC (64% in patients with PNI vs 91% in those without, $P = .002$).[47–49]

The evaluation of PNI risk in CSCC should vary depending on the thickness of the nerves affected and the presence of clinical and/or radiologic signs of invasion. Infiltration of nerves with a diameter of less than 0.1 mm, for instance, is associated with a low risk of local or distant complications, while invasion of nerves measuring more than 0.1 mm in diameter has been associated with poor short-term and long-term prognosis (CSCC-specific death of 0% in individuals with PNI of nerves < 0.1 mm vs 32% in those with PNI of nerves > 0.1 mm, $P = .003$).[50]

PNI can manifest as an incidental finding on histology, with or without accompanying symptoms. Symptoms include pain on palpation, regional paresthesia, and acute intermittent or shooting pain. Based on data from the University of Florida College of Medicine in the United States, it has been suggested that patients with asymptomatic PNI not visible on radiography have a better prognosis than those with clinical or radiological evidence of PNI (5-year local control rate of 87% vs 55%, $P = .006$).[51]

Lymphovascular Invasion

Recent studies have suggested that lymphovascular invasion may increase the risk of metastasis in CSCC. Moore et al.[43] defined lymphovascular invasion as an independent predictor of lymph node metastasis in a multivariate analysis (OR, 7.54, $P. < .00001$), and reported that 40% of patients with metastasis had lymphovascular invasion, compared with just 8% of those without. The prognostic significance of lymphovascular invasion, however, has been questioned by some authors.[12,50]

The implications of CSCC in dermal lymph vessels, which has been rarely described, are unknown, but it may increase

the risk of recurrence and explain in-transit metastasis (Fig. 5).

Other Factors

Other factors that have been proposed as possible prognostic factors in CSCC are peritumoral actinic keratosis,[37–39] Clark level,[37–39] Ki67 expression, desmoplasia,[26,38] and the presence of a tumor inflammatory response with mainly eosinophils and plasma cells. The true prognostic value, however, of these factors, is still a matter of debate and needs to be investigated in further studies.

Molecular Markers in CSCC

Seventy percent of patients with metastatic CSCC have 1 or more of the defining features of high-risk CSCC described above. However, between 20% and 30% do not (those with thin, small CSCCs), suggesting that other, as yet unknown variables, probably have an important role in the pathogenesis of high-risk CSCC.[12,51] Of relevance in this group are certain molecular alterations that appear to be associated with a subgroup of CSCCs with more aggressive behavior. Specifically, it has been suggested that mutations in genes expressing the epidermal growth factor receptor (EFGR), and to a lesser extent, *p16* and *CKS1B* mutations, are the main molecular alterations involved in high-risk CSCC; confirmation of this would have important therapeutic implications.[52–56]

Epidermal Growth Factor Receptor

Tumors that overexpress EGFR tend to be associated with more advanced disease, a greater risk of lymph node metastasis, and higher rates of early recurrence and shorter survival in various malignant disorders, including squamous cell carcinoma of the mucosa of the upper aerodigestive tract.[57–62]

Just 1 small study has analyzed the importance of *EFGR* mutations in the prognosis of CSCC. In an analysis of 15 cases of metastatic CSCC in the head and neck region, EGFR overexpression was significantly associated with metastatic potential, with strong overexpression found in 79% of patients with metastatic disease and in just 36% of those without.[62,63] Overexpression was independent of gene amplification. Alternative mechanisms that would explain the increase in EGFR expression include increased messenger RNA (mRNA) transcription, activating receptor mutations, increased levels of receptor ligands, and increased expression of heterologous receptors, such as Her-2.[62] With respect to Her-2, abnormal Her-2 expression and alterations in the gene encoding Her-2 (chromosome 17) have been analyzed in patients with EGFR overexpression.[63] While the authors did not observe Her-2 overexpression in any of the 27 cases they analyzed, they did detect Her-2 polysomy, which, like in breast cancer, was not associated with increased overexpression. The absence of overexpression leads to treatment failure with anti-Her2 drugs, such as trastuzumab.[64] Although the significance of this last finding has not yet been clarified, the detection of Her-2 polysomy may have an impact on predicting therapeutic response to tyrosine kinase inhibitors.[61,62] Nevertheless, EGFR overexpression has only been observed in up 65% to 75% of metastatic CSCCs, adding strength to the hypothesis that multiple factors are involved in the etiology of this form of CSCC.[62,63]

p16

Data from several studies support a correlation between *p16* overexpression and degree of malignancy, suggesting that *p16* expression (like *p53* expression) might be a biomarker of tumor progression.[64,65] Other authors, however, have reported a correlation between loss of *p16* expression and high-risk CSCC.[66,67] In 1 of these studies, Chang et al.[67] detected a correlation between loss of *p16* expression and the development of metastasis, suggesting that loss of this protein might be a predictor of poor prognosis.

The explanation behind why elevated levels of p16 might be associated with poor prognosis could be related to the coexistence of HPV infection. However, with the exception of epidermodysplasia verruciformis in immunocompromised individuals and CSCC in areas other than the head and neck, the role of HPV in the development of CSCC is still a matter of debate.

Another possible explanation for increased *p16* levels might be *p16* UV-induced changes. The presence of a mutated *p16*, with a long half-life but with a loss of antioncogenic function, would explain the increased risk of malignancy.

CSK1B

The *CKS1B* gene encodes the cyclin-dependent kinases regulatory subunit 1. The protein binds to the catalytic subunit of cyclin-dependent kinases and play an essential role in their biologic function. *CKS1B* mRNA appears to be expressed in different patterns throughout the cell cycle of HeLa cells, indicating a specific role for the encoded protein. The peptide plays a role in cell-cycle regulation by interacting with other proteins, mainly SKP2 and CDKN1B.[68,69]

CKS1B appears to have a critical role in tumor progression in CSCC. In a study of 43 CSCCs and 26 actinic keratoses, Salgado et al.[70] analyzed CKS1B gene and protein status using fluorescence in situ hybridization and immunohistochemical analysis, and reported that chromosome 1 polysomy was a frequent event in both CSCC (30 of 43 cases) and actinic keratosis (13 of 23 cases). *CKS1B* amplification, observed in 4 cases (9.3%), was associated in all cases with aggressive tumor behavior (PNI, lymph node spread, and CSCC in transplant recipients).

In conclusion, *CKS1B* amplifications might be a marker of high-risk CSCC.

Towards a New Prognostic Classification of CSCC

CSCC is one of the most common cancers in the world and, as such, responsible for many deaths.

The ability to clearly differentiate between high-risk and low-risk CSCC would appear to be key in improving

Table 1 Comparison Between the 6th and 7th Editions of the American Joint Committee on Cancer Staging Manuals.

Factor	6th Edition	7th Edition	Comment
T category	T1: ≤ 2 cm T2: 2-5 cm T3: > 5 cm	T1: ≤ 2 cm T2: > 2 cm	No evidence regarding usefulness of cutoff of 5 cm
Histologic grade	Not included	Risk factor (poorly differentiated tumor)	Degree of differentiation described as a risk feature in CSCC
High-risk factors	Not included	Inclusion of risk factors that modify the T designation (elevated by 1 level in patients with ≥ 2 factors)	Factors include: - Histologic grade -Location on the ear or in area of the chin and lip -Thickness > 2 mm -Clark level ≥ IV -Perineural invasion
Extradermal invasion (histologic)	Used to determine T4	Removed	Lack of data showing clear prognostic value
Anatomic site	Not included	Included as high risk factor	Location of CSCC on the ear or in the area of the chin and lip associated with worse prognosis
Facial/cranial bone invasion	Included in T4 as invasion of extradermal structures	Invasion of maxilla, mandible, orbit, and temporal bone, defined as T3	Correlated with stage of SSC of the head and neck
N category	Based on presence (N1) or absence (N0) of nodal disease	Disease staged as N0-N3 based on the size and number of involved nodes	Correlated with stage of SCC of the head and neck, and with recently published data
M category	Based on presence (M1) or absence (M0) of distant metastasis	No changes	M, only TNM category not modified

survival and optimizing management of the disease according to risk.

The American Joint Committee on Cancer (AJCC) recently modified its staging system for CSCC.[71] The main changes are summarized in Table 1. One of the most significant changes was the introduction of a list of high-risk clinical and histologic features that modify the T designation, regardless of tumor size. These features include a tumor thickness of more than 2 mm, a Clark level of IV or more, location on the external ear or nonglabrous lip, PNI, bone involvement, and poor tumor differentiation.[71] Based on our experience, however, this list is not sufficient to accurately define high-risk CSCC. While anatomic location and PNI are good prognostic predictors, the situation with tumor differentiation and Clark level is less clear. Furthermore, the establishment of a cutoff of 2 mm for differentiating between low-risk and high-risk CSCC deserves special mention. A high proportion of CSCCs are thicker than 2 mm, meaning that this cutoff has high sensitivity but very poor specificity in terms of predicting risk. Finally, the new AJCC staging system does not include important factors such as immune system status, tumor recurrence, or lymphovascular invasion.

In its latest update, published in 2010, the National Comprehensive Cancer Network (NCNN) proposed that the treatment of CSCC should be guided by a series of variables. It lists a set of risk factors and considers 2 possible scenarios. First, it recommends that all CSCCs with any of the risk factors shown in Table 2 should be surgically excised with safety margins of 10 mm or with Mohs micrographic surgery. And second, it recommends that patients with 3 or more of these factors should receive special attention. In our opinion, the NCCN guidelines also have shortcomings. A high percentage of patients have at least 1 of the factors shown in Table 2, meaning that suggesting

Table 2 Factors Considered by the National Comprehensive Cancer Network for the Definition of Risk in Cutaneous Squamous Cell Carcinoma.

The presence of any of the following risk factors is sufficient to justify excision with a safety margin of 10 mm or complete assessment of all margins (Mohs micrographic surgery)
Tumor size ≥ 2 cm (or 1 cm on the head and 6 mm on the genitals, hands, and feet)
Tumor thickness ≥ 4 mm
Poorly defined borders
Recurrent tumor
Immunosuppression
Previous radiation
Chronic inflammation
Rapid growth
Perineural or vascular invasion
Moderate or poor differentiation

Special attention should be paid to patients with 3 or more of the above risk factors.

Table 3 Major and Minor Criteria That Define High-Risk Cutaneous Squamous Cell Carcinoma (CSCC).[a]

Major Criteria	Clinical Features	Histologic Features
	Personal history of: dystrophic epidermolysis bullosa epidermodysplasia verruciformis dyskeratosis congenita xeroderma pigmentosum oculocutaneous albinism	Tumor thickness > 6 mm
	Immunosuppression due to: solid organ transplantation (heart and lung) hematologic disease (chronic lymphatic leukemia, small lymphocytic lymphoma)	Perineural invasion (nerves with a diameter ≥ 0.1 mm)
	Tumor site (lip, anogenital region, external ear)	Bony involvement
	Tumor recurrence	
	Tumor diameter > 5 cm	
Minor Criteria	Clinical Features	Histologic Features
	Immunosuppression due to: solid organ transplantation (kidney and liver)	Tumor thickness 2-6 mm
	Lesion arising on preexisting lesion (scar, radiation dermatitis area)	Poorly differentiated tumor
	Tumor diameter 2-5 cm[b]	Certain CSCC variants (acantholytic, isolated cell, basosquamous)
	Infection with human immunodeficiency virus	Human papillomavirus infection in histologic section from immunosuppressed patient
		Lymphovascular invasion

[a] A CSCC is considered high risk if the tumor meets: a) 3 minor criteria, b) 2 major criteria and 2 minor criteria, and c) 1 major criterion and 4 minor criteria.
[b] > 1.5 cm on lip and ear.

treatment modifications based on this alone would appear to be quite a broad recommendation. The second scenario is even more confusing, as the NCNN considers that patients with 3 or more of the risk factors shown in Table 2 should receive special treatment. However, there is no mention of exactly how treatment or follow-up should be modified.

Proposal for Defining and Managing High-Risk CSCC

Defining High-Risk CSCC. In our opinion, the main difficulty with attempts to define high-risk CSCC to date is the fact that most studies have identified the risk factors discussed in this article in isolation. There have been no analyses of their cumulative effects.

Based on data systematically stored in a database at our hospital, a specialist skin cancer center, and data from the literature, we designed a system to identify the cumulative value of each of the prognostic factors that define high-risk CSCC. We divided risk factors into major and minor criteria and created a scoring system to differentiate between low risk and high-risk CSCC (Table 3). We then considered high-risk CSCC to be any CSCC with *a)* 3 major criteria, *b)* 2 major and 2 minor criteria, and *c)* 1 major criterion and 4 minor criteria. Our proposed definition of high-risk CSCC, which has important prognostic and therapeutic implications, needs to be corroborated in prospective studies that analyze the different prognostic factors for CSCC from a global perspective.

Optimizing the Management of High-Risk CSCC. Our provisional definition of high-risk CSCC would lead to a more aggressive treatment approach and much closer follow-up.

Treating High-Risk CSCC. The treatment of choice for high-risk CSCC is surgical excision. In conventional excision, safety margins should range from 4 mm for tumors measuring 2 cm or less to 6 mm for larger tumors. Mohs micrographic surgery is the treatment of choice for tumors in locations with a risk of cosmetic or functional sequelae and for recurrent tumors[42,71–75] (Fig. 7).

Radiation therapy, which is considered by several authors to be a first-line treatment for high-risk CSCC, produces poorer outcomes than surgical excision and is associated with a high percentage of earlier and more aggressive recurrence and considerable direct and indirect costs. Furthermore, it can cause iatrogenic carcinogenesis in the irradiated area. Radiation therapy, thus, should be reserved for patients who are not candidates for surgery, either because of poor general health status or the inability to achieve disease-free margins by surgical techniques.[42,71–73]

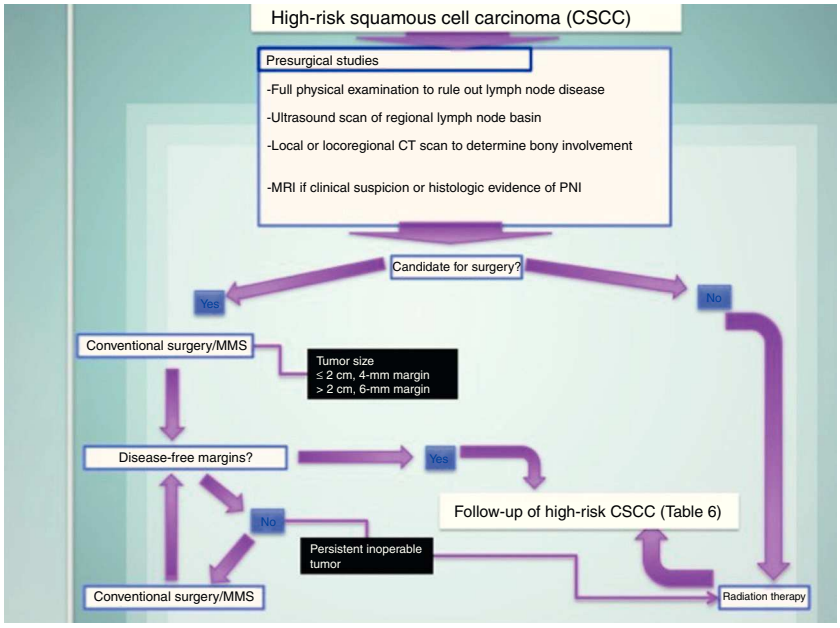

Figure 7 Treatment algorithm for high-risk squamous cell carcinoma; CT indicates computed tomography; MMS, Mohs micrographic surgery; MRI, magnetic resonance imaging; PNI, perineural invasion.

Table 4 Follow-up Protocol for Squamous Cell Carcinoma Based on Risk.

Period[a]	Check-up Frequency	Physical Examination	Additional Tests
Years 1 and 2 CT/MRI every 6 mo[c]	Every 3 mo	Yes	Ultrasound every 3 mo[b]
Years 3-5 Annual CT/MRI[c]	Every 6 mo	Yes	Ultrasound every 6 mo[b]
After fifth year	Annually	Yes	No

Abbreviations: CT; computed tomography; MRI, magnetic resonance imaging.
[a] Ninety-five percent of recurrences and metastasis are observed in the first 5 years.
[b] Ultrasound of lymph node regions in the area of the tumor.
[c] In immunosuppressed patients or patients with perineural invasion of the main nerve trunk.

Additional Strategies for the Management of High-Risk CSCC

Additional management strategies are necessary in patients with high-risk CSCC given the high associated risk of lymph node invasion and mortality.

Sentinel Lymph Node Biopsy

In 2006, Ross et al.[74] concluded that sentinel lymph node biopsy (SLNB) was associated with a reliable diagnosis of locoregional invasion and with low morbidity in CSCC provided that the surgeon had sufficient experience. This is also the case with cutaneous melanoma.

Indeed, SLNB provides better results in CSCC than in melanoma as the early detection of lymph node involvement in high-risk CSCC leads to a significant reduction in mortality.

We therefore believe that SLNB is justified in CSCCs defined as high-risk according to our provisional definition (Table 3 and Fig. 6).

Follow-up in Patients With a History of High-Risk CSCC

High-risk CSCC is associated with a higher risk of recurrence and lymph node metastasis than low-risk CSCC in the first 5 years after treatment. The early detection of recurrence and lymph node metastasis is of supreme importance, as has been shown in multiple studies. Of particular relevance in this respect is a study by Ebrahimi et al.[75] that showed that in patients with CSCC and lymph node involvement, a single diseased lymph node measuring 3 cm or less in diameter without extracapsular nodal spread had a low risk of distant metastasis and mortality.

We therefore propose a follow-up approach based on risk (Table 4), whereby patients with high-risk CSCC should be followed very closely during the first 5 years after surgery. Close monitoring is particularly important during the first 24 months, which is when the risk of lymph node invasion is highest.

Conclusions

The incidence of CSCC has increased considerably in recent years. CSCC is the second most common nonmelanocytic skin tumor in the general population and causes a similar number of deaths as melanoma.

A clear definition of epidemiological, clinical, and histologic factors of CSCCs with high rates of systemic spread (high-risk CSCC) will lead to a different management approach when dealing with this subgroup of patients. Such an approach should include more exhaustive staging at the time of diagnosis, more aggressive treatment, including SLNB, and closer follow-up. Individualized care will help to reduce the mortality associated with this malignant tumor.

In the not-so-distant future, the characterization of the molecular biology of high-risk CSCC and the analysis of genetic differences with respect to low-risk CSCC will probably help to define the malignant potential of this high-risk variant and allow the optimization of treatment via drugs that act on key molecular targets in the pathogenesis of squamous cell carcinoma.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

1. Ramirez CC, Federman DG, Kirsner RS. Skin cancer as an occupational disease: the effect of ultraviolet and other forms of radiation. Int J Dermatol. 2005;44:95–100.
2. de Vries E, de Poll-Franse LV, Louwman WJ, de Gruijl FR, Coebergh JW. Predictions of skin cancer incidence in the Netherlands up to 2015. Brit J Dermatol. 2005;152: 481–8.
3. Kwa RE, Campana K, Moy RL. Biology of cutaneous squamous cell carcinoma. J Am Acad Dermatol. 1992;26: 1–26.
4. Johnson TM, Rowe DE, Nelson BR, Swanson NA. Squamous cell carcinoma of the skin (excluding lip and oral mucosa). J Am Acad Dermatol. 1992;26:467–84.
5. Yoong C, De'Ambrosis B. Cutaneous invasive squamous cell carcinoma: 10-year experience and recommendations for follow up. Australas J Dermatol. 2009;50:261–5.
6. Clayman GL, Lee JL, Holsinger C, Zhou X, Duvic M, El-Naggar AK, et al. Mortality risk from squamous cell skin cancer. J Clin Oncol. 2005;22:759.
7. Moore B, Weber RS, Prieto V, Zhou X, Duvic M, El-Naggar AK, et al. Lymph node metastases from cutaneous squamous cell carcinoma of the head and neck. Laryngoscope. 2006;115: 1561.
8. Veness MJ. Defining patients with high-risk cutaneous squamous cell carcinoma. Australas J Dermatol. 2006;47: 28–33.
9. Veness MJ, Morgan GJ, Palme C, Gebski V. Cutaneous head and neck squamous cell carcinoma metastatic to cervical lymph nodes: surgery and adjuvant radiotherapy should be considered best practice. Laryngoscope. 2005;44:870–5.
10. Audet N, Palme CE, Gullane PJ, Gilbert RW, Brown DH, Irish J, et al. Cutaneous metastatic squamous cell carcinoma to the parotid gland: analysis and outcome. Head Neck. 2004;26:727–32.
11. Burnworth B, Popp S, Stark H-J, Steinkraus V, Bröcker EB, Hartschuh W, et al. Gain of 11q/cyclin D1 overexpression is an essential early step in skin cancer development and causes abnormal tissue organization and differentiation. Oncogene. 2006;25:4399.
12. Cherpelis B, Marcusen C, Lang PG. Prognostic factors for metastasis in squamous cell carcinoma of the skin. Dermatol Surg. 2002;28:268.
13. Fine JD, Johnson LB, Weiner M, Li KP, Suchindran C. Epidermolysis bullosa and the risk of life-threatening cancers: the National EB registry experience, 1986-2006. J Am Acad Dermatol. 2009;60:203–11.
14. Lyakhovitsky A, Barzilai A, Fogel M, Trau H, Huszar M. Expression of E-cadherin and beta catenin in cutaneous squamous cell carcinoma and its precursors. Am J Dermatopathol. 2004;26:372.
15. Heath M, Jaimes N, Lemos B, Mostaghimi A, Wang LC, Peñas PF, et al. Clinical characteristics of Merkel cell carcinoma at diagnosis in 195 patients: the AEIOU features. J Am Acad Dermatol. 2008;58:375–81.
16. Veness MJ, Quinn DI, Ong CS, Keogh AM, Macdonald PS, Cooper SG, et al. Aggressive cutaneous malignancies following cardiothoracic transplantation: the Australian experience. Cancer. 1999;85:1758–64.
17. Mehrany K, Weenig RH, Lee KK, Pittelkow MR, Otley CC. Increased metastasis and mortality from cutaneous squamous cell carcinoma in patients with chronic lymphocytic leukemia. J Am Acad Dermatol. 2005;53:1067–71.
18. Euvrard S, Kanitakis J, Decullier E, Butnaru AC, Lefrançois N, Boissonnat P, et al. Subsequent skin cancers in kidney and heart transplant recipients after the first squamous cell carcinoma. Transplantation. 2006;81:1093–100.
19. Veness MJ, Palme CE, Morgan GJ. High-risk cutaneous squamous cell carcinoma of the head and neck: results from 266 treated patients with metastatic lymph node disease. Cancer. 2006;106:2389–96.
20. Martinez JC, Clark CO, Stasko T, Euvrard S, Brown C, Schanbacher CF, et al. Defining the clinical course of metastatic skin cancer in organ transplant recipients. Arch Dermatol. 2003;139:301–6.
21. Nguyen P, Vin-Christian K, Ming ME, Berger T. Aggressive squamous cell carcinomas in persons infected with the human immunodeficiency virus. Arch Dermatol. 2002;138: 758–63.
22. Rowe DE, Carroll RJ, Day CD. Prognostic factors for local recurrence, metastasis, and survival rates in squamous cell carcinoma of the skin, ear and lip. J Am Acad Dermatol. 1992;26:976–90.
23. Rodolico V, Barresi E, Di Lorenzo R, Leonardi V, Napoli P, Rappa F, et al. Lymph node metastasis in lower lip squamous cell carcinoma in relation to tumour size, histologic variables and p27kip1 protein expression. Oral Oncol. 2004;40:92–8.
24. Mullen JT, Feng L, Xing Y, Mansfield PF, Gershenwald JE, Lee JE, et al. Invasive squamous cell carcinoma of the skin: defining a high-risk group. Ann Surg Oncol. 2006;13:902–9.
25. Kraus DH, Carew JF, Horrison LB. Regional lymph node metastasis from cutaneous squamous cell carcinoma. Arch Otolaryngol. 1998;124:582–7.
26. Brantsch KD, Meisner C, Schönfisch B, Trilling B, Wehner-Caroli J, Röcken M, et al. Analysis of risk factors determining prognosis of cutaneous squamous-cell carcinoma: a prospective study. Lancet Oncol. 2008;9:713–20.
27. Lee D, Nash M, Har-El G. Regional spread of auricular and preauricular cutaneous malignancies. Laryngoscope. 1996;106:998–1001.
28. Tavin E, Persky M. Metastatic cutaneous squamous cell carcinoma of the head and neck region. Laryngoscope. 1996;106:156–8.
29. Yoon M, Chougule P, Dufresne R, Wanebo HJ. Localized carcinoma of the external ear is an unrecognized aggressive disease

with a high propensity for local regional recurrence. Am J Surg. 1992;164:574–7.
30. Lai SY, Weinstein GS, Chalian AA, Rosenthal DI, Weber RS. Parotidectomy in the treatment of aggressive cutaneous malignancies. Arch Otolaryngol Head Neck Surg. 2002;128: 521–6.
31. Alam M, Ratner D. Cutaneous squamous cell carcinoma. N Engl J Med. 2001;344:975.
32. Motley R, Kersey P, Lawrence C. Multiprofessional guidelines for the management of the patient with primary cutaneous squamous cell carcinoma. Br J Dermatol. 2002:18–25.
33. Huang C, Boyce SM. Surgical margins of excision for basal cell carcinoma and squamous cell carcinoma. Semin Cutan Med Surg. 2004;23:167–73.
34. Plasmeijer EI, Neale RE, de Koning MN, Quint WG, McBride P, Feltkamp MC, et al. Persistence of betapapillomavirus infections as a risk factor for actinic keratoses, precursor to cutaneous squamous cell carcinoma. Cancer Res. 2009;69:8926–31.
35. Proby CM, Harwood CA, Neale RE, Green AC, Euvrard S, Naldi L, et al., EPI-HPV-UV-CA group. A case-control study of betapapillomavirus infection and cutaneous squamous cell carcinoma in organ transplant recipients. Am J Transplant. 2011;11:1498–508.
36. Harwood CA, Surentehran T, McGregor JM, Spink PJ, Leigh IM, Breuer J, et al. Human papillomavirus infection and non-melanoma skin cancer in immunosuppressed and immunocompetent individual. J Med Virol. 2000;61:289–97.
37. Forslund O, Lindelof B, Hradil E, Nordin P, Stenquist B, Kirnbauer R, et al. High prevalence of cutaneous human papillomavirus DNA on the top of skin tumors but not in «Stripped» biopsies from the same tumors. J Invest Dermatol. 2004;123:388–94.
38. Breuninger H, Schaumburg-Lever G, Holzschuh J, Horny HP. Desmoplastic squamous cell carcinoma of skin and vermilion surface. A highly malignant subtype of skin cancer. Cancer. 1997;79:915–9.
39. Turner SJ, Morgan GJ, Palme CE, Veness MJ. Metastatic cutaneous squamous cell carcinoma of the external ear: a high risk cutaneous subsite. J Laringol Otol. 2010;124:26–31.
40. Jensen V, Prasad AR, Smith A, Raju M, Wendel CS, Schmelz M, et al. Prognostic criteria for squamous cell cancer of the skin. J Surg Res. 2010;159:509–16.
41. De Visscher JG, Gooris PJJ, Vermey A, Roodenburg JLN. Surgical margins for resection of squamous cell carcinoma of the lower lip. Int J Oral Maxillofac Surg. 2002;31:154–7.
42. Brodland DG, Zitelli JA. Surgical margins for excision of primary cutaneous squamous cell carcinoma. J Am Acad Dermatol. 1992;27:241–8.
43. Moore B, Weber RS, Prieto V, El-Naggar A, Holsinger FC, Zhou X, et al. Lymph node metastases from cutaneous squamous cell carcinoma of the head and neck. Laryngoscope. 2006;115:1561.
44. Veness MJ. High-risk cutaneous squamous cell carcinoma of the head and neck. J Biomed Biotechnol. 2007;3:1–6.
45. Nijsten TE, Stern RS. Oral retinoid use reduces cutaneous squamous cell carcinoma risk in patients with psoriasis treated with psoralen-UVA: a nested cohort study. J Am Acad Dermatol. 2003;49:644–50.
46. Goepfert H, Dichtel WJ, Medina JE, Lindberg RD, Luna MD. Perineural invasion in squamous cell skin carcinoma of the head and neck. Am J Surg. 1984;148:542–7.
47. Frierson HF, Cooper PH. Prognostic factors in squamous cell carcinoma of the lower lip. Hum Pathol. 1986;17:346–54.
48. Garcia-Serra A, Hinerman RW, Mendenhall WM, Amdur RJ, Morris CG, Williams LS, et al. Carcinoma of the skin with perineural invasion. Head Neck. 2003;25:1027–33.
49. McCord MW, Mendenhall WM, Parsons JT, Flowers FP. Skin cancer of the head and neck with incidental microscopic perineural invasion. Int J Radiat Oncol Biol Phys. 1999;43:591–5.
50. Ross AS, Whalllen FM, Elenitsas R, Xu X, Troxel A, Schmults CD. Diameter of involved nerves predicts outcomes in cutaneous squamous cell carcinoma with perineural invasion: an investigator-blinded retrospective cohort study. Dermatol Surg. 2009;35:1859–66.
51. Nolan RC, Chan MTL, Heenan PJ. A clinicopathologic review of lethal nonmelanoma skin cancers in Western Australia. J Am Acad Dermatol. 2005;52:101–8.
52. Veness MJ. Treatment recommendations in patients diagnosed with high-risk cutaneous squamous cell carcinoma. Australas Radiol. 2005;49:365–76.
53. Maubec E, Duvillard P, Velasco V, Crickx B, Avril MF. Immunohistochemical analysis of EGFR and HER-2 patients with metastatic squamous cell carcinoma of the skin. Anticancer Res. 2005;25:1205–10.
54. Shimizu T, Izumi H, Oga A, Furumoto H, Murakami T, Ofuji R, et al. Epidermal growth factor receptor overexpression and genetic aberrations inmetastatic squamous-cell carcinoma of the skin. Dermatology. 2001;202:203–6.
55. Orth G. Human papillomavirus associated with epidermodysplasia verruciformis in nonmelanoma skin cancers: guilty or innocent? Invest Dermatol. 2005;125:12–3.
56. Fallon JH, Seroogy KB, Loughlin SE, Morrison RS, Bradshaw RA, Knaver DJ, et al. Epidermal growth factor immunoreactive material in the central nervous system: location and development. Science. 1984;224:1107–9.
57. Jensen P, Hansen S, Moller B, Leivestad T, Pfeffer P, Geiran O, et al. Skin cancer in kidney and heart transplant recipients and different long-term immunosuppressive therapy regimens. J Am Acad Dermatol. 1999;40:177–86.
58. Hartevelt MM, Bavinck JN, Kootte AMM, Vermeer BJ, Van denbroucke JP. Incidence of skin cancer after renal transplantation in The Netherlands. Transplantation. 1990;49: 506–9.
59. Lindelof B, Sigurgeirsson B, Gäbel H, Stern RS. Incidence of skin cancer in 5,356 patients following organ transplantation. Br J Dermatol. 2000;143:513–9.
60. Cooper JZ, Brown MD. Special concern about squamous cell carcinoma of the scalp in organ transplant recipients. Arch Dermatol. 2006;142:755–8.
61. Sobin LH, Wittekind C, editors. TNM classification of malignant tumours: International Union Against Cancer. 6th ed. New York: John Wiley & Sons; 2002.
62. Chang S, Low I, Ng D, Brasch H, Sullivan M, Davis P, et al. Epidermal growth factor receptor: a novel biomarker for aggressive head and neck cutaneous squamous cell carcinoma. Hum Pathol. 2008;39:34.
63. Bauman JE, Eaton KD, Martins RG. Treatment of recurrent squamous cell carcinoma of the skin with cetuximab. Arch Dermatol. 2007;143:889–92.
64. Corbalan-Velez R, Oviedo-Ramirez I, Ruiz-Macia JA, Conesa-Zamora P, Sánchez-Hernández M, Martínez-Barba E, et al. Immunohistochemical staining of p16 in squamous cell carcinomas of the genital and extragenital area. Actas Dermosifiliogr. 2011;102:439–47.
65. Blokx WA, de Jong EM, de Wilde PC, Bulten J, Link MM, Ruiter DJ, et al. P16 and p53 expression in (pre)malignant epidermal tumors of renal transplant recipients and immunocompetent individuals. Mod Pathol. 2003;16:869–78.
66. Salama ME, Mahmood MN, Qureshi HS, Ma C, Zarbo RJ, Ormsby AH. P16INK4a expression in actinic keratosis and Bowen's disease. Br J Dermatol. 2003;149:1006–12.
67. Chang TG, Wang J, Chen LW, Hsu CY, Chang HW, Chen JS, et al. Loss of expression of the p16 gene is frequent in malignant skin tumors. Biochem Biophys Res Commun. 1997;230: 85–8.
68. Richardson HE, Stueland CS, Thomas J, Russell P, Reed SI. Human cDNAs encoding homologs of the small p34Cdc28/

Cdc2-associated protein of Saccharomyces cerevisiae and Schizosaccharomyces pombe. Genes Dev. 1990;4:1332–44.
69. Pines J. Cell cycle: reaching for a role for the Cks proteins. Curr Biol. 1996;6:1399–402.
70. Salgado R, Toll A, Alameda F, Baro T, Martín-Ezquerra G, Sanmartín O, et al. CKS1B amplification is a frequent event in cutaneous squamous cell carcinoma with aggressive clinical behaviour. Genes Chromosomes Cancer. 2010;49:1054–61.
71. Cutaneous squamous cell carcinoma and other cutaneous carcinomas. AJCC Cancer staging handbook. Edge, S.B. 7th edition. Springer; USA, 2010. p. 359-75.
72. Pugliano-Mauro M, Goldman G. Mohs surgery is effective for high-risk cutaneous squamous cell carcinoma. Dermatol Surg. 2010;36:1544–53.
73. Jennings L, Schmults CD. Management of high-risk cutaneous squamous cell carcinoma. J Clin Aesthet Dermatol. 2010;3:39–48.
74. Ross AS, Schmults CD. Sentinel lymph node biopsy in cutaneous squamous cell carcinoma: a systematic review of the English literature. Dermatol Surg. 2006;32:1309.
75. Ebrahimi A, Clark JR, Lorincz BB, Milross CG, Veness MJ. Metastatic head and neck cutaneous squamous cell carcinoma: Defining a low-risk patient. Head Neck. 2012;34:365–70.