

REVIEW ARTICLE

Cutaneous squamous cell carcinoma and human papillomavirus

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Abstract. The relationship between mucosal human papillomavirus (HPV) and cervical carcinoma or anogenital squamous cell carcinoma (SCC) is becoming increasingly evident, whereas a link between HPV and other cutaneous SCCs is less clear. Recent studies have reported links between epidermodysplasia-*verruciformis*-associated HPV and extragenital cutaneous SCC, particularly in immunosuppressed patients, although immunocompetent patients have also been affected. Mucosal HPV could also be linked to some types of Bowen disease and certain SCCs of the fingers, oropharyngeal mucosa, etc. We review the possible oncogenic mechanisms involving mucosal HPV and epidermodysplasia-*verruciformis*-associated HPV. Most SCCs could be explained by the combined action of HPV, immunosuppression, and the oncogenic and immunosuppressive effect of UV radiation. HPV might be associated with worse prognosis of SCC, with implications for clinical practice including greater risk of metastasis.

Key words: human papillomavirus, HPV, epidermodysplasia *verruciformis*, squamous cell carcinoma, skin

CARCINOMA ESPINOCELULAR CUTÁNEO Y PAPILOMAVIRUS (VPH)

Resumen. La relación entre los virus papilomas humanos (VPH) de tipo mucoso (VPH-muc) y el carcinoma de cérvix o los carcinomas espinocelulares (CE) de la región ano-genital es cada vez más evidente. Sin embargo, la relación del VPH con el resto de los CE cutáneos es más controvertida. Recientes publicaciones relacionan los VPH tipo epidermodisplasia *verruciforme* (VPH-EV) con los CE cutáneos extra-genitales, sobre todo en los pacientes inmunodeprimidos, aunque también en los inmunocompetentes. Los VPH-muc también se podrían relacionar con algunas enfermedades de Bowen y determinados CE: dedos de manos, mucosa oro-faríngea, etc.

Revisamos los posibles mecanismos oncogénicos de los VPH-muc y los VPH-EV. La mayoría de los CE podrían explicarse por la acción conjunta de los VPH, la inmunodepresión y los efectos oncogénicos e inmunosupresores de las radiaciones ultravioleta. Los VPH podrían implicar un peor pronóstico de los CE, con más posibilidades de producir metástasis, entre otras implicaciones en la práctica clínica.

Palabras clave: virus papiloma humano (VPH), epidermodisplasia *verruciforme*, carcinoma espinocelular, piel.

Introduction

Human papillomaviruses (HPV) are DNA viruses that belong to the Papovaviridae family and more than 95 different genotypes have been characterized in humans. HPV can be classified into 3 basic groups^{1,2}:

1. *Cutaneous HPV (HPV-cut)*. This group includes HPV subtypes 1, 2, 3, 4, 7, 10, 26 to 29, 41, 49, 57, 60, 63, 65, and 75 to 77, among many others.

2. *Mucosal HPV (HPV-muc)*. HPV-muc are further classified as high risk (subtypes 16 and 18), intermediate risk (subtypes 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68, 73, 82, and 83), and low risk (subtypes 6, 11, 26, 30, 32, 40, 42 to 44, 53 to 55, 62, 66, 70, 72, and 81) according to their oncogenic potential.

3. *Epidermodysplasia verruciformis-associated HPV (HPV-EV)*. This group especially includes HPV subtypes 5, 8, 9, 10, 12, 14, 15, 17, 19 to 25, 36 to 38, 47, and 50. It tends also to include certain HPV-cut which are very frequent in epidermodysplasia *verruciformis*, such as subtypes 3 and 10. New HPV-EV have been characterized thus increasing the HPV-EV list.^{3,4}

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Accepted June 5, 2007.

Some studies have correlated the histological signs attributable to the cytopathic effects of HPV with the

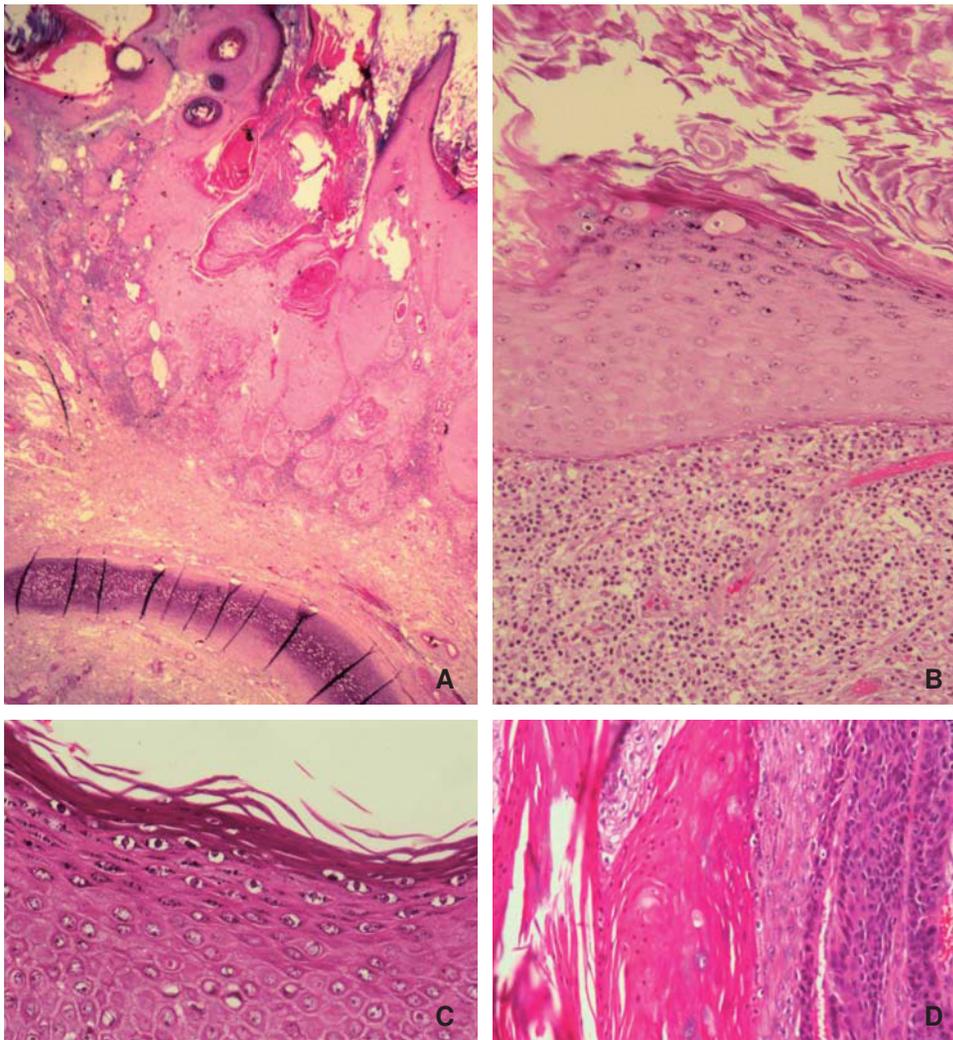


Figure 1. (A) Common wart in the surface layer and underlying squamous cell carcinoma (Hematoxylin-eosin $\times 16$). (B) Epidermodysplasia verruciformis-type cells in the granular layer of the epidermis (Hematoxylin-eosin $\times 100$). (C) Detail of the epidermis next to a squamous cell carcinoma. Increase in the size and quantity of keratohyalin granules, "bird's eyes," and other indirect signs of human papillomavirus (HPV). (Hematoxylin-eosin $\times 400$.) (D) Pseudoparakeratosis among other signs attributable to HPV (Hematoxylin-eosin $\times 200$).

different HPV subtypes (detected through polymerase chain reaction [PCR]), and thus, it would be possible to establish the HPV subtype for each lesion according to the indirect histological signs that are observed, at least approximately.⁵⁻⁸ The histological signs attributable to HPV infection include the following: the keratohyalin granules are more numerous and larger, with different shapes and sizes, and can be seen not only in the cytoplasm but also in the nucleus; koilocytes; papillomatosis; pseudoparakeratosis (rounded nuclei instead of flat ones in the stratum corneum) is occasionally observed; and cells like those seen in epidermodysplasia verruciformis (large globular cells with cytoplasmic granules)^{9,10} (Figure 1).

Cutaneous HPV

HPV-cut is normally found in benign lesions such as common warts, flat warts, plantar warts, some

epidermal cysts, etc.¹¹ The high prevalence of common, flat, and plantar warts in the healthy population (especially during childhood and adolescence) and the mechanisms of HPV infection (by personal contact and following minor injuries or skin damage) are epidemiological data suggesting that HPV may be present asymptotically, and is probably symbiotic on the mucocutaneous surfaces of most immunocompetent adults and children.^{11,12}

In benign lesions, HPV DNA is located in the cell nucleus, although outside the chromosomes. In contrast, in serious dysplasias and in carcinomas caused by HPV, viral DNA is fully integrated in the cellular genome of the keratinocytes.¹³ On the other hand, cases of squamous cell carcinoma (SCC) arising from common warts¹⁴⁻¹⁹ or plantar warts¹¹ have been described, suggesting that nononcogenic HPV, such as HPV-cut, could favor the development of malignant processes when occurring with other factors, such as immunosuppression.

Mucosal HPV

HPV-muc is normally detected in genital warts and in premalignant and malignant lesions in the anogenital region, and also in some patients (eg, those infected by the human immunodeficiency virus [HIV]) who apparently do not present clinical lesions.²⁰ HPV-muc has also been detected in lesions caused by genital lichen sclerosus, although the pathogenic role played by HPV in this process remains unclear.²¹

The oncogenic role of HPV-muc (especially some subtypes such as 16, 18, 31, 33, and 35) has been well characterized in uterine cervical carcinoma; similarly, the possible oncogenic role of these HPV-muc subtypes in cutaneous SCC of the anogenital region (including SCC of the vulva, penis, anus, or perineum) is increasingly clear.²²⁻²⁴ On the other hand, HPV-muc has also been associated with the following:

1. Benign lesions such as oral papillomas^{25,26} (especially subtypes 6 and 11) or some types of seborrheic keratosis.²⁷
2. Some studies have observed classic oncogenic HPV-muc (such as subtype 16), with almost symbiotic behavior on the skin and mucous membranes of children and immunocompetent adults.^{12,28}
3. Extragenital Bowen disease. Clavel et al²⁹ detected HPV-muc in 83% of patients with Bowen disease (78 cases out of 94). While some studies have found similar percentages of HPV-muc in Bowen disease of the hands,^{30,31} others have found much lower percentages³²⁻³⁸ or a predominance of HPV-EV.^{3,39,40} Some recent publications support a possible relationship between HPV-muc and some cases of Bowen disease.^{41,42}
4. SCC of the area around the nails or of the fingers⁴³⁻⁴⁵ (a possible explanation would be autoinoculation or hand-genital transmission).
5. SCC of the oropharyngeal mucous membrane, by acting synergistically or potentiating other oncogenic factors such as tobacco.^{25,26,46-48} According to some recent studies, HPV-muc could also be involved in carcinomas of the larynx, esophagus, tonsils, etc.⁴⁹⁻⁵⁴

In conclusion, HPV-muc could be involved in SCC of the anogenital region,²²⁻²⁴ SCC of the fingers,⁴³⁻⁴⁵ in some cases of Bowen disease,²⁹⁻⁴² and in carcinomas of the oropharyngeal mucosa^{25,26,46-48} (also in carcinomas of the larynx, esophagus, tonsils, etc, as reported in several studies⁴⁹⁻⁵⁴).

Epidermodysplasia Verruciformis-Associated HPV

Epidermodysplasia verruciformis-associated HPV could be an important oncogenic factor in most cases of cutaneous

SCC, especially in immunocompromised but also in immunocompetent patients.⁵⁵

Cutaneous SCC and HPV-EV in Immunocompromised Patients

The increased incidence of SCC in immunocompromised patients (HIV-infected patients, transplant patients treated with immunosuppressive drugs, patients with cancer, etc) has been confirmed in numerous studies.⁵⁶⁻⁶⁰ Some have found HPV-EV in approximately 80% of cases of SCC in immunocompromised patients,^{4,61} an observation which would support a possible oncogenic relationship between HPV-EV and SCC in these patients. It should be pointed out that these figures are similar to those reported in cervical carcinoma, where HPV (albeit mucosal) is found in approximately 90% of cases.^{23,62,63}

Some authors have pointed out striking parallels between immunocompromised patients and those affected by HPV-EV.⁵⁵ Immunocompromised patients often develop multiple actinic keratoses, Bowen disease, and SCC, especially in sunlight-exposed areas,⁵⁷ similar to the situation in patients with epidermodysplasia verruciformis.⁶⁴ Furthermore, immunocompromised patients have been described with lesions which are clinically and histologically indistinguishable from those in epidermodysplasia verruciformis.⁶⁵⁻⁶⁸ Finally, some subtypes of HPV-EV are often detected in immunocompromised patients, such as HPV 5 and 8, which are usually found in tumors of patients with epidermodysplasia verruciformis.^{55,56,66,69,70}

Thus, HPV-EV could play an important oncogenic role in immunocompromised patients, by acting synergistically and potentiating other oncogenic factors such as UV radiation.^{55,70,71}

Cutaneous SCC and HPV-EV in Immunocompetent Patients

The possible oncogenic role of HPV-EV in immunocompetent patients is more debatable. HPV (predominantly HPV-EV) has been found in approximately 35%-45% of cases,^{29,34,56,61,72-74} which are similar percentages to those in the healthy population; based on these data, some authors consider that HPV-EV are normal symbiotes of the skin.^{12,75}

Thus, there may not be many differences between immunocompromised and immunocompetent patients. We consider that almost all patients with SCC are immunocompromised based on the fact that several studies discuss local or systemic immunosuppression caused by chronic exposure to UV radiation.^{11,76-78} This would trigger

Table. Immunosuppressive Effects of UV Radiation

↓ Presentation of antigens (↓ number and function of Langerhans cells, among others)
↑ Immunosuppressive mediators and ↑ Th2 lymphocyte activity (inducing skin tolerance and lack of cellular response for antigens): <ul style="list-style-type: none"> • Increase in IL-10, IL-6, IL-15, TNF-α
↓ Th1 lymphocyte activity (directing cell-mediated immune activation): <ul style="list-style-type: none"> • Decrease in IL-12, IL-1, IL-7, IFN-γ
↓ Number and function of natural-killer cells
Induction of T lymphocyte apoptosis
Other immunosuppressive effects that inhibit the cytotoxic and immune response: <ul style="list-style-type: none"> • Trans-urocanic acid changes to cis-urocanic acid • Free radical formation • Alterations in keratinocyte DNA • Alterations in surface receptors and adhesion molecules

Abbreviations: IFN, interferon; IL, interleukin; TNF, tumor necrosis factor; Th, T helper.

a cascade of interrelated biochemical and cellular alterations that would result in immunosuppressive effects^{11,76-78} (Table). Another study has shown that old age itself is a strong immunocompromising factor.⁷⁹ If we take into account the immunosuppressive effects of UV radiation and old age, and given that most patients with SCC have also undergone chronic exposure to solar radiation, are older, or both, these patients could be considered immunocompromised, at least with local or cutaneous immunosuppression.

On the other hand, several studies have reported an increase in the quantity of HPV-EV residing in hair follicles compared to the rest of the skin,^{80,81} in sunlight-exposed areas,^{3,70,75} in patients with psoriasis receiving psoralen-UV-A (PUVA) therapy,^{70,82-85} and in elderly patients,^{70,75} all of which may give an indication of what occurs in the general population after chronic exposure to the sun.

Despite these studies, several authors have expressed doubt regarding the possible involvement of HPV in extragenital SCC, especially in immunocompetent patients.^{12,75} There could be several reasons for this:

1. As discussed later, the oncogenic mechanisms of HPV-EV are not as well characterized as those of HPV-muc. HPV-EV has low oncogenic potential *in vitro* and requires the synergistic action of solar radiation to induce oncogenesis. Unlike in cervical carcinoma (where HPV could be the main oncogenic factor),^{23,63} in skin cancers HPV would require other oncogenic factors, such as UV radiation or immunosuppression.⁷¹

2. In many cases it is not possible to demonstrate the presence of HPV through *in situ* hybridization or PCR. If we review recent publications, there is wide variability regarding the prevalence of HPV or which HPV subtypes predominate (cutaneous, EV or mucosal),⁵⁵ especially in studies conducted with immunocompetent patients, both those investigating SCC^{29,33-35,56,61,72-74,86} and those studying actinic keratosis or Bowen disease.^{33,35-40,56,72,73,86} Some have investigated the same lesions using different PCR techniques (with the aim of comparing them) and have obtained totally different results depending on the PCR technique used.⁸⁷⁻⁸⁹ Frequently, larger or smaller quantities of HPV are found depending on the PCR technique used.^{56,89} On the other hand, some studies have only investigated HPV-muc or some specific HPV serotypes.^{35,36,38,40,86} Furthermore, viral DNA could be degraded and not detected when the sample lesions have been in paraffin for a long period or if they are in formalin for more than 24 hours, which is usually the case in most skin lesions.^{90,91} A modified *in situ* hybridization method has recently been introduced that may have greater sensitivity and specificity.^{92,93}
3. Finally, HPV may be eliminated (thus making its DNA undetectable) once the process of oncogenesis is initiated; that is, HPV could disappear having left its fingerprint.^{94,95}

Association Between Cutaneous SCC and HPV-EV

Several relatively recent studies have found a statistically significant relationship between the presence of HPV-EV and greater risk of acquiring SCC in immunocompromised and immunocompetent patients:

1. Stark et al⁷⁰ found a statistically significant association between positive serological findings for HPV-8 and the presence of nonmelanoma skin cancer (NMSC) in immunocompetent patients. Serological findings were positive for HPV-8 in 45.6% of immunocompetent patients with NMSC (36 out of 79), whereas in people without NMSC this figure was lower: 7.6% (16 out of 210) in immunocompetent patients and 21% (39 of 185) in immunocompromised patients.
2. More recently, Masini et al⁹⁶ reported a similar statistically significant association between positive serological findings for HPV-8 and the presence of SCC in immunocompetent patients.
3. Feltkamp et al⁹⁷ also found a relationship between positive serological findings for HPV-EV (especially for HPV-8 and HPV-38) and a greater risk of acquiring SCC.
4. In an Australian study, Boxman et al³² reported that the presence of HPV-EV in hair follicles was related to the

risk of acquiring NMSC (statistically nonsignificant association).

5. Harwood et al⁹⁸ studied a group with SCC and one without SCC using PCR and found a statistically significant relationship between the presence of HPV-EV in normal skin and the risk of acquiring SCC (in immunocompetent and immunocompromised patients).
6. Karagas et al⁹⁹ reached similar conclusions, finding a relationship between positive serological findings for HPV-EV, particularly for HPV-5, and a greater risk of SCC.
7. With respect to the precursor lesions, Bouwes Bavinck et al¹⁰⁰ found a relationship between positive serological findings for HPV-8 and the presence of actinic keratosis. Boxman et al¹⁰¹ also found an association between the presence of HPV-EV in hair follicles and the presence of actinic keratosis, although this association was significant only in men.

In conclusion, HPV-EV is involved in cutaneous SCC in immunocompromised and immunocompetent patients,^{55,96,98,102} as shown by Purdie et al.¹⁰²

Oncogenesis and HPV

1. The tumor suppressor protein p16 inhibits tumor progression in cells whose DNA has been damaged through inhibition of the *retinoblastoma* (*Rb*) gene.^{103,104} Greater expression of p16 induced by UV radiation¹⁰⁵⁻¹⁰⁷ and oncogenic HPV has been reported.^{104,108} Specific oncogenic HPV induce alterations in the *Rb* gene through E7 protein and thus trigger the suppressive mechanism of p16 protein.^{104,108,109} Furthermore, these oncogenic HPV, through E6 protein, induce alterations in the *p53* gene,^{55,104} and this would at least partly explain the alterations found in *p53* in areas not exposed to sunlight.¹¹⁰
2. The oncogenic mechanisms of HPV-muc in cutaneous SCC (for example, in some SCC of the fingers⁴⁴) are comparable to those found in cervical and anogenital carcinomas. Mucosal HPV may produce alterations in the *p53* gene and *Rb* gene through proteins E6 and E7, respectively, inducing cellular immortality or tumor progression.^{55,104} In addition, *p53* gene polymorphism should be taken into account; specific individuals with amino acid changes (arginine instead of proline) in codon 72 of the *p53* gene, especially homozygotes, would be more susceptible to the oncogenic action of this HPV-muc E6 protein.¹¹¹⁻¹¹³
3. The oncogenic mechanisms of HPV-EV are less clear. On the one hand, HPV-EV E6 and E7 proteins, unlike those of HPV-muc, do not have the same affinity for the *p53* and *Rb* genes, respectively.^{55,114} In addition, the in vitro oncogenic potential of these HPV-EV is low¹¹⁵

and would require the combined action of UV radiation to be oncogenic, as occurs in epidermodysplasia verruciformis patients.¹¹⁴ A possible oncogenic mechanism of these HPV-EV could be proapoptotic Bak protein inhibition.^{114,116} UV radiation induces Bak protein activation, which then induces apoptosis, regardless of the p53 mechanism. The E6 protein of these HPV-EV (and of other HPV-cut such as HPV-10 and HPV-77) induces Bak protein degradation, thus inhibiting the apoptosis induced by UV radiation, independently of the state of p53.^{114,116} Jackson et al¹¹⁶ demonstrated that SCC without HPV had high levels of apoptotic Bak protein, whereas this protein was practically undetectable in SCC with HPV. In an in vitro model, Akgul et al¹¹⁷ demonstrated how the E7 protein of HPV-EV type 8 could induce keratinocyte proliferation and, more importantly, favor the migration and invasion of these keratinocytes into the dermis. Nevertheless, more in vivo studies are needed to support this finding.

4. Furthermore, HPV uses immunosuppressive-type mechanisms to evade the immune system; these mechanisms could indirectly facilitate the escape of tumor cells.^{94,118,119}

Taking all the above into account, HPV could play an important role as an initiator and promoter of SCC oncogenesis (Figure 2).

In short, in SCC oncogenesis, the main pathogenic factor seems to be solar radiation, especially chronic or continuous exposure.^{120,121} Other individual factors, such as skin phototype, age, or changes in the *p53* suppressor gene, could increase individual susceptibility to SCC.⁵⁵

Certain HPV, such as HPV-EV, could also be another synergistic or potentiating pathogenic factor in epithelial oncogenesis, especially in immunocompromised patients, but also in immunocompetent patients.^{55,96,98} The significant immunosuppressive effects caused by UV radiation,⁷⁶⁻⁷⁸ age,⁷⁹ or HPV itself^{94,118,119} would induce immunosuppression (local, systemic, or both) in immunocompetent patients and would aggravate immunosuppression in immunocompromised patients. Alterations in the immune system (especially in its control functions and the elimination of cells with damaged DNA) could play as important a role as UV radiation.⁷¹

The interaction of these 3 pathogenic factors (UV radiation, alterations in the immune system, and HPV), together with other factors that may have an effect on these 3 (age, skin phototype, individual predisposition, etc) could explain most cases of SCC oncogenesis (Figure 3).

Practical and Clinical Implications

The possible role of HPV in SCC oncogenesis could have a strong impact on clinical practice and dermatological treatment:

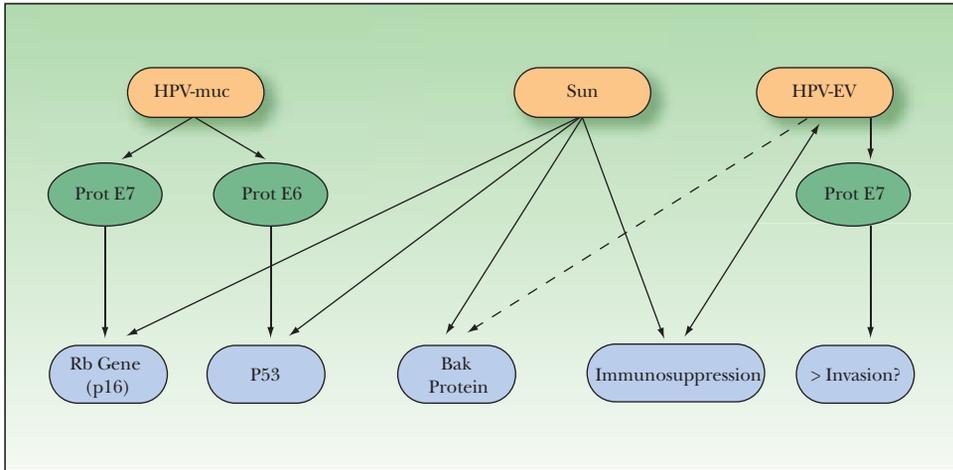


Figure 2. Oncogenic mechanisms of mucous-type human papillomavirus (HPV-muc) and epidermodysplasia verruciformis-type HPV (HPV-EV).

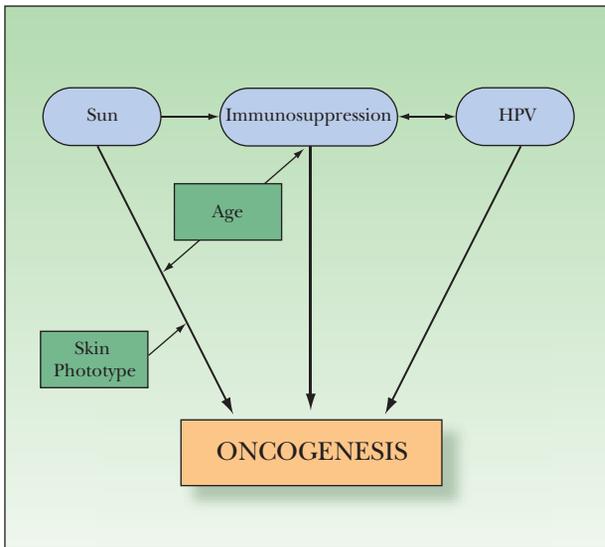


Figure 3. Summary of squamous cell carcinoma oncogenesis. HPV indicates human papillomavirus.

1. For example, HPV-muc vaccine^{122,123} has recently been shown to have a strong prophylactic effect against cervical carcinoma, but also against SCC of the anogenital region, certain types of skin SCC, some cases of Bowen disease, and even other SCC such as those of the oropharyngeal region. These vaccines may also prove effective in men and so their use should not be restricted to young women. It may be possible to develop a vaccine against HPV-EV in the future (this would be very useful in immunocompromised patients, as well as in patients with a fair skin phototype or those who have undergone intense chronic exposure to the sun, etc).

2. The employment of early prevention campaigns similar to those against cervical carcinoma (some countries conduct these for SCC of the anus in patients at risk).^{124,125}
3. Treatment with topical imiquimod for acuminata condylomata¹²⁶ could be indirectly useful as a prophylactic against certain SCC (for example, in the anogenital region).
4. Several publications have recently appeared stating that the presence of HPV in SCC would indicate worse prognosis with, for example, a greater probability of dissemination and a greater incidence of lymph node metastases.¹²⁷⁻¹³⁰ Some authors attribute this to the changes caused by HPV-muc in the regulator gene *p53* through E6 protein, among other effects.¹³¹⁻¹³³ Another possible explanation is that E7 protein could induce greater invasiveness toward the dermis¹¹⁷ (especially in the case of HPV-muc, since HPV-EV has only been studied *in vitro*¹¹⁷), thus encouraging the development of metastasis or greater aggressiveness in these tumors. Nevertheless, many more studies are needed to confirm or reject these hypotheses.
5. Some authors have shown that *Polypodium leucotomos* is useful in preventing SCC developing in patients receiving PUVA treatment.¹³⁴ Given that the pathogenesis of SCC in these patients is very similar to that in immunocompromised patients, this treatment could also be very useful in preventing SCC in the latter (we are aware that a multicenter study on this issue is currently taking place).
6. In any case, as occurs in epidermodysplasia verruciformis, HPV-EV requires the synergistic action of UV radiation to induce oncogenesis⁵⁵ (through apoptotic Bak protein inhibition,¹¹⁶ for example). Thus, the best way to prevent cutaneous SCC continues

to be reduction of exposure to the sun or the use of topical or systemic sun filters.

Conflicts of Interest

The authors declare no conflicts of interest.

References

- De Villiers EM. Taxonomic classification of papillomaviruses. *Papillomavirus Report*. 2001; 12: 57-63.
- Chan SY, Delius H, Halpern AL, Bernard HU. Analysis of genomic sequences of 95 papillomavirus types: uniting typing, phylogeny and taxonomy. *J Virol*. 1995; 69: 3074-83.
- de Jong-Tieben LM, Berkhout RJ, ter Schegget J, Vermeer BJ, de Fijter JW, Bruijn JA, et al. The prevalence of human papillomavirus DNA in benign keratotic skin lesions of renal transplant recipients with and without a history of skin cancer is equally high: a clinical study to assess risk factors for keratotic skin lesions and skin cancer. *Transplantation*. 2000;69:44-9.
- Berkhout RJ, Bouwes Bavinck JN, ter Schegget J. Persistence of human papillomavirus DNA in benign and (pre)malignant skin lesions from renal transplant recipients. *J Clin Microbiol*. 2000;38:2087-96.
- Egawa K. New types of human papillomaviruses and intracytoplasmic inclusion bodies: a classification of inclusion warts according to clinical features, histology and associated HPV types. *Br J Dermatol*. 1994;130:158-66.
- Gross G, Pfister H, Hagedorn M, Gissmann L. Correlation between human papillomavirus (HPV) type and histology of warts. *J Invest Dermatol*. 1982;78:160-4.
- Jablonska S, Orth G, Obalek S, Croissant O. Cutaneous warts. Clinical, histologic and virologic correlations. *Clin Dermatol*. 1985;3:71-82.
- Androphy EJ. Molecular biology of human papillomavirus infection and oncogenesis. *J Invest Dermatol*. 1994;103:248-56.
- Weedon D. Viral diseases. In: Weedon D, editor. *Skin Pathology*. New York: Churchill Livingstone Ed; 1998. p.583-603.
- Grayson W, Calonje E, McKee PhH. Infections diseases of the skin. In: McKee PhH, Calonje E, Granter S, editors. *Pathology of the skin with clinical correlations*. 3rd ed. Philadelphia: Elsevier Mosby Ed; 2005. p.838-50.
- Villarrubia VG, Costa LA, Pérez M, Vidal S, Jaen P. Epidemiología e inmunopatogenia del cáncer cutáneo no melanoma. El papel iniciador y promotor del VPH. *Piel*. 2001;16:428-38.
- Astori G, Lavergne D, Benton C, Hockmayr B, Egawa K, Garbe C, et al. Human papillomaviruses are commonly found in normal skin of immunocompetent hosts. *J Invest Dermatol*. 1998;110:752-5.
- Howley PM. The role of papillomaviruses in human cancer. In: De Vita V, Hellman S, Rosenberg SA, editors. *Important advances in oncology*. Philadelphia: Lippincott Ed; 1987. p.55-73.
- Shelley WB, Wood MG. Transformation of the common wart into squamous cell carcinoma in a patient with primary lymphedema. *Cancer*. 1981;48:820-4.
- Noel JC, Peny MO, Goldschmidt D, Verhest A, Heenen M, de Dobbeleer G. Human papillomavirus type 1 DNA in verrucous carcinoma of the leg. *J Am Acad Dermatol*. 1993;29:1036-8.
- Kopelson PL, Nguyen QH, Moy RL. Verruca vulgaris and radiation exposure are associated with squamous cell carcinoma of the finger. *J Dermatol Surg Oncol*. 1994;38-41.
- Echt AF, Hurwitz RM, Davis TE. Warts: benign or malignant? *Indiana Med*. 1991;84:476-9.
- Milburn PB, Brandsma JL, Goldsman CI, Teplitz ED, Heilman EI. Disseminated warts and evolving squamous cell carcinoma in a patient with acquired immunodeficiency syndrome. *J Am Acad Dermatol*. 1988;19:401-5.
- Philips ME, Ackerman AB. «Benign» and «malignant» neoplasms associated with verrucae vulgares. *Am J Dermatopathol*. 1982;4:61-84.
- Castaño-Suárez E, Guerra-Tapia A, Iglesias-Díez L, Llamas-Martín R, Picazo-Garza JJ, Postigo-Llorente C, et al. Detección y tipado de Papillomavirus humano en raspados de pacientes infectados por el virus de la inmunodeficiencia humana. *Actas DermoSifiliogr*. 2003;94:17-23.
- Acosta-Dibarráz G, González-Domínguez V, Paciel-Vaz J, Vignale-Peirano R. Presencia del virus del papiloma humano en lesiones de liquen escleroso y atrofico vulvar. Estudio por inmunohistoquímica e hibridización in situ. *Actas DermoSifiliogr*. 2002;93:389-92.
- Bjorge T, Engeland A, Luostarinen T, Mork J, Gislefoss RE, Jellum E, et al. Human papillomavirus infection as a risk factor for anal and perianal skin cancer in a prospective study. *Br J Cancer*. 2002;87:61-4.
- Bosch FX, Lorincz A, Muñoz N, Meijer CJ, Shah KV. The causal relation between human papillomavirus and cervical cancer. *J Clin Pathol*. 2002;55:244-65.
- Ahmed AM, Madkan V, Tying SK. Human papillomaviruses and genital disease. *Dermatol Clin*. 2006;24:157-65.
- Kreimer AR, Clifford GM, Boyle P, Franceschi S. Human papillomavirus types in head and neck squamous cell carcinomas worldwide: a systematic review. *Cancer Epidemiol Biomarkers Prev*. 2005;14:467-75.
- Major T, Szarka K, Sziklai I, Gergely L, Czeglédy J. The characteristics of human papillomavirus DNA in head and neck cancers and papillomas. *J Clin Pathol*. 2005;58: 51-5.
- Gushi A, Kanekura T, Kanzaki T, Eizuru Y. Detection and sequences of human papillomavirus DNA in nongenital seborrheic keratosis of immunocompetent individuals. *J Dermatol Sci*. 2003;31:143-9.
- Rice PS, Mant C, Cason J, Bible JM, Muir P, Kell B, et al. High prevalence of human papillomavirus type 16 infection among children. *J Med Virol*. 2000;61:70-5.
- Clavel CE, Pham-Huu V, Durlach AP, Birembaut PL, Bernard PM, Derancourt CG. Mucosal oncogenic human papillomaviruses and extragenital Bowen disease. *Cancer*. 1999;86:282-7.
- Pham-Huu V, Derancourt C, Clavel C, Durlach A, Birembaut P, Bernard P. Oncogenic mucosal human papillomaviruses in Bowen's disease of the hands. *Ann Dermatol Venereol*. 1999;126:808-12.
- Mitsuishi T, Sata T, Matsukura T, Iwasaki T, Kawashima M. The presence of mucosal human papillomavirus in Bowen's disease of the hands. *Cancer* 1997;79:1911-7.

32. Boxman IL, Russell A, Mulder LH, Bavinck JN, Schegget JT, Green A. Case-control study in subtropical Australian population to assess the relation between non-melanoma skin cancer and epidermodysplasia verruciformis human papillomavirus DNA in plucked eyebrow hairs. *Int J Cancer*. 2000;86:118-21.
33. Meyer T, Arndt R, Christophers E, Nindl I, Stockfleth E. Importance of human papillomaviruses for the development of skin cancer. *Cancer Detect Prev*. 2001;25:533-47.
34. Meyer T, Arndt R, Christophers E, Stockfleth E. Frequency and distribution of HPV types detected in cutaneous squamous-cell carcinomas depends upon the HPV detection system: a comparison of four PCR assays. *Dermatology*. 2000;201:204-11.
35. Iftner A, Klug SJ, Garbe, Blum A, Stancu A, Wilczynski SP, et al. The prevalence of human papillomavirus genotypes in non-melanoma skin cancers of non-immunosuppressed individuals identifies high-risk genital types as possible risk factors. *Cancer Res*. 2003;63:7515-9.
36. Lampert A, Pauwels C, Dubouché C, Morel G, Poveda JD, Perie G. Détection de papillomavirus humains dans la maladie de Bowen cutanée extragénitale chez le sujet immunocompétent. *Ann Dermatol Vénereol*. 2000;127:40-5.
37. Hama N, Ohtsuka T, Yamazaki S. Detection of mucosal human papilloma virus DNA in bowenoid papulosis, Bowen's disease and squamous cell carcinoma of the skin. *J Dermatol*. 2006;33:331-7.
38. Derancourt C, Mouglin C, Chopard-Lallier M, Coumes-Marquet S, Drobacheff C, Laurent R. Oncogenic human papillomaviruses in extra-genital Bowen disease revealed by in situ hybridization. *Ann Dermatol Vénereol*. 2001;128:715-8.
39. Mitsuishi T, Kawana S, Kato T, Kawashima M. Human papillomavirus infection in actinic keratosis and Bowen's disease: comparative study with expression of cell-cycle regulatory proteins p21 (waf1/cip1), p53, PCNA, Ki-67, and Bcl-2 in positive and negative lesions. *Human Pathol*. 2003;34:886-92.
40. Quereux G, N'Guyen JM, Dreno B. Human papillomavirus and extragenital in situ carcinoma. *Dermatology*. 2004;209:40-5.
41. Sato T, Morimoto A, Ishida Y, Matsuo I. Human papillomavirus associated with Bowen's disease of the finger. *J Dermatol*. 2004;31:927-30.
42. Zheng S, Adachi A, Shimizu M, Shibata SI, Yasue S, Sakakibara A, et al. Human papillomaviruses of the mucosal type are present in some cases of extragenital Bowen's disease. *Br J Dermatol*. 2005;152:1243-7.
43. Forslund O, Nordin P, Hansson BG. Mucosal human papillomavirus types in squamous cell carcinomas of the uterine cervix and subsequently on the fingers. *Br J Dermatol*. 2000;142:1148-53.
44. Alam M, Caldwell JB, Eliezri YD. Human papillomavirus-associated digital squamous cell carcinoma: Literature review and report of 21 new cases. *J Am Acad Dermatol*. 2003;48:385-93.
45. Zabawski EJ jr, Washak RV, Cohen JB, Cockerell CJ, Brown SM. Squamous cell carcinoma of the nail bed: is finger predominance a clue to etiology? A report of 5 cases. *Cutis*. 2001;67:59-64.
46. Gillison ML. Human papillomavirus-associated head and neck cancer is a distinct epidemiologic, clinical, and molecular entity. *Semin Oncol*. 2004;31:744-54.
47. Koppikar P, de Villiers EM, Mulherkar R. Identification of human papillomaviruses in tumors of the oral cavity in an Indian community. *Int J Cancer*. 2005;113:946-50.
48. Syrjänen S. Human papillomavirus (HPV) in head and neck cancer. *J Clin Virol*. 2005;32Suppl1:S59-66.
49. Castillo A, Aguayo F, Koriyama C, Torres M, Carrascal E, Corvalán A, et al. Human papillomavirus in esophageal squamous cell carcinoma in Colombia and Chile. *World J Gastroenterol*. 2006;12:6188-92.
50. Farhadi M, Tahmasebi Z, Merat S, Kamangar F, Nasrollahzadeh D, Malekzadeh R. Human papillomavirus in squamous cell carcinoma of esophagus in a high-risk population. *World J Gastroenterol*. 2005;11:1200-3.
51. Brouchet L, Valmary S, Dahan M, Didier A, Galateau-Salle F, Brousset P, et al. Detection of oncogenic virus genomes and gene products in lung carcinoma. *Br J Cancer*. 2005;92:743-6.
52. Coissard CJ, Besson G, Polette MC, Monteau M, Birembaut PL, Clavel CE. Prevalence of human papillomaviruses in lung carcinomas: a study of 218 cases. *Mod Pathol*. 2005;18:1606-9.
53. Torrente MC, Ampuero S, Abud M, Ojeda JM. Molecular detection and typing of human papillomavirus in laryngeal carcinoma specimens. *Acta Otolaryngol*. 2005;125:888-93.
54. Dahlstrand HM, Dalianis T. Presence and influence of human papillomaviruses (HPV) in tonsillar cancer. *Adv Cancer Res*. 2005;93:59-89.
55. Harwood CA, Proby CM. Human papillomaviruses and non-melanoma skin cancer. *Curr Opin Infect Dis*. 2002;15:101-14.
56. Harwood CA, Surentheran T, McGregor JM, Spink PJ, Leigh IM, Breuer J, et al. Human papillomavirus infection and non-melanoma skin cancer in immunosuppressed and immunocompetent individuals. *J Med Virol*. 2000;61:289-97.
57. Berg D, Otley CC. Skin cancer in organ transplant recipients: Epidemiology, pathogenesis, and management. *J Am Acad Dermatol*. 2002;47:1-17.
58. Fuente MJ, Sabat M, Roca J, Lauzurica R, Fernández-Figueras MT, Ferrandiz C. A prospective study of the incidence of skin cancer and its risk factors in a Spanish Mediterranean population of kidney transplant recipients. *Br J Dermatol*. 2003;149:1221-6.
59. Fortina AB, Piaserico S, Caforio AL, Abeni D, Alaibac M, Angelini A, et al. Immunosuppressive level and other risk factors for basal cell carcinoma and squamous cell carcinoma in heart transplant recipients. *Arch Dermatol*. 2004;140:1079-85.
60. Lindelof B, Sigurgeirsson B, Gabel H, Stern RS. Incidence of skin cancer in 5356 patients following organ transplantation. *Br J Dermatol*. 2000;143:614-8.
61. O'Connor DP, Kay EW, Leader M, Murphy GM, Atkins GJ, Mabruk MJ. Altered p53 expression in benign and malignant skin lesions from renal transplant recipients and immunocompetent patients with skin cancer: correlation with human papillomaviruses? *Diagn Mol Pathol*. 2001;10:190-9.

62. Melo A, Roa I, Montenegro S, Capurro I, Roa JC. Detection of human papillomavirus in cytologic samples or biopsies of the cervix. *Rev Med Chil.* 2005;133:639-44.
63. Bertelsen BI, Kugarajh K, Skar R, Laerum OD. HPV subtypes in cervical cancer biopsies between 1930 and 2004: detection using general primer pair PCR and sequencing. *Virchows Arch.* 2006;449:141-7.
64. Sánchez-García VP, Sanz A, Eloy C, Vera A, Martín T, Sánchez-Fajardo F. Epidermodisplasia verruciforme. *Actas DermoSifiliogr.* 2005;96:531-3.
65. Berger TG, Sawchuk WS, Leonardi C, Langenberg A, Tappero J, Leboit PE. Epidermodysplasia verruciformis-associated papillomavirus infection complicating human immunodeficiency virus disease. *Br J Dermatol.* 1991;124:79-83.
66. Tieben LM, Berkhout RJ, Smits HL, Bouwes Bavinck JN, Vermeer BJ, Bruijn JA, et al. Detection of epidermodysplasia verruciformis-like human papillomavirus types in malignant and premalignant skin lesions of renal transplant recipients. *Br J Dermatol.* 1994;131:226-30.
67. Slawsky LD, Gilson RT, Hockley AJ, Libow LF. Epidermodysplasia verruciformis associated with severe immunodeficiency, lymphoma, and disseminated molluscum contagiosum. *J Am Acad Dermatol.* 1992;27:448-50.
68. Obalek S, Favre M, Szymanczyk J, Misiewicz J, Jablonska S, Orth G. Human papillomavirus (HPV) types specific of epidermodysplasia verruciformis detected in warts induced by HPV3 or HPV3-related types in immunosuppressed patients. *J Invest Dermatol.* 1992;98:936-41.
69. Lutzner MA, Orth G, Dutronquay V, Ducasse MF, Kreis H, Crosnier J. Detection of human papillomavirus type 5 DNA in skin cancers of an immunosuppressed renal allograft recipient. *Lancet.* 1983;2:422-4.
70. Stark S, Petridis AK, Ghim SJ, Jenson AB, Bouwes Bavinck JN, Gross G, et al. Prevalence of antibodies against viruslike particles of epidermodysplasia verruciformis-associated HPV 8 in patients at risk of skin cancer. *J Invest Dermatol.* 1998;111:696-701.
71. Aubin F, Laurent R. Human papillomavirus-associated cutaneous lesions. *Rev Prat.* 2006;56:1905-13.
72. Pfister H, Fuchs PG, Majewski S, Jablonska S, Pniewska I, Malejczyk M. High prevalence of epidermodysplasia verruciformis-associated human papillomavirus DNA in actinic keratoses of the immunocompetent population. *Arch Dermatol Res.* 2003;295:273-9.
73. Forslund O, Lindelof B, Hradil E, Nordin P, Stenquist B, Kirnbauer R, et al. High prevalence of cutaneous human papillomavirus DNA on the top of skin tumors but not in «Stripped» biopsies from the same tumors. *J Invest Dermatol.* 2004;123:388-94.
74. Stockfleth E, Nindl I, Sterry W, Ulrich C, Schmook T, Meyer T. Human papillomaviruses in transplant-associated skin cancers. *Dermatol Surg.* 2004;30:604-9.
75. Antonsson A, Forslund O, Ekberg H, Sterner G, Hansson BG. The ubiquity and impressive genomic diversity of human skin papillomaviruses suggest a commensal nature of these viruses. *J Virol.* 2000;74:11636-41.
76. Aubin F. Mechanism involved in ultraviolet light-induced immunosuppression. *Eur J Dermatol.* 2003;13:515-23.
77. Carrascosa JM. Efectos de la radiación ultravioleta sobre el sistema inmunitario. Implicaciones terapéuticas. *Piel.* 2004;19:303-12.
78. Hanneman KK, Cooper KD, Baron ED. Ultraviolet immunosuppression: mechanisms and consequences. *Dermatol Clin.* 2006;24:19-25.
79. Fagnoni FF, Vescovini R, Passeri G, Bologna G, Pedrazzoni M, Lavagetto G, et al. Shortage of circulating naive CD8+T cells provides new insights on immunodeficiency in aging. *Blood.* 2000;95:2860-8.
80. Bouwes Bavinck JN, Feltkamp M, Struijk L, ter Schegget J. Human papillomavirus infection and skin cancer risk in organ transplant recipient. *J Invest Dermatol Symp Proc.* 2001;6:207-11.
81. Jenson AB, Geyer S, Sundberg JP, Ghim S. Human papillomavirus and skin cancer. *J Invest Dermatol Symp Proc.* 2001;6:203-6.
82. Weissenborn SJ, Hopfl R, Weber F, Smola H, Pfister HJ, Fuchs PG. High prevalence of a variety of epidermodysplasia verruciformis-associated human papillomaviruses in psoriatic skin of patients treated or not treated with PUVA. *J Invest Dermatol.* 1999;113:122-6.
83. Wolf P, Seidl H, Bäck B, Binder B, Hofler G, Quehenberger F, et al. Increased prevalence of human papillomavirus in hairs plucked from patients with psoriasis treated with psoralen-UVA. *Arch Dermatol.* 2004;140:317-24.
84. Harwood CA, Spink PJ, Suretheran T, Leigh IM, Hawke JL, Proby CM, et al. Detection of human papillomavirus DNA in PUVA-associated non-melanoma skin cancers. *J Invest Dermatol.* 1998;111:123-7.
85. Zumtobel U, Schwarze HP, Favre M, Taieb A, Delaunay M. Widespread cutaneous carcinomas associated with human papillomavirus 5, 14 and 20 after introduction of methotrexate in two long-term PUVA-treated patients. *Dermatology.* 2001;202:127-30.
86. Biliris KA, Koumantakis E, Dokianakis DN, Sourvinos G, Spandidos DA. Human papillomavirus infection of non-melanoma skin cancers in immunocompetent hosts. *Cancer Lett.* 2000;161:83-8.
87. Harwood CA, Spink PJ, Suretheran T, Leigh IM, de Villiers EM, Mc Gregor JM, et al. Degenerate and nested PCR: a highly sensitive and specific method for detection of human papillomavirus infection in cutaneous warts. *J Clin Microbiol.* 1999;37:3545-55.
88. Forslund O, Antonsson A, Nordin P, Stenquist B, Hansson BG. A broad range of human papillomavirus types detected with a general PCR method suitable for analysis of cutaneous tumours and normal skin. *J Gen Virol.* 1999;80:2437-43.
89. Forslund O, Ly H, Higgins G. Improved detection of cutaneous human papillomavirus DNA by single tube nested «hanging droplet» PCR. *J Virol Methods.* 2003;110:129-36.
90. Manos MM, Ting Y, Wright DK. Use of polymerase chain reaction amplification for the detection of genital human papillomavirus. *Cancer Cel.* 1989;7:209-14.
91. Martínez A, Nás R, LaCruz C, Hellín T, Tercero JC, Valverde E, et al. Detección y tipado de papilomavirus humano por amplificación genómica en biopsias, frotis y orina. *Acta ginecologica.* 1995;52:51-6.
92. Dabic MM, Hlupic L, Babic D, Jukic S, Seiwerth S. Comparison of polymerase chain reaction and catalyzed signal amplification in situ hybridization methods for human papillomavirus detection in paraffin-embedded cervical preneoplastic and neoplastic lesions. *Arch Med Res.* 2004;35:511-6.

93. Biedermann K, Dandachi N, Trattner M, Vogl G, Doppelmayer H, More E, et al. Comparison of real-time PCR signal-amplified in situ hybridization and conventional PCR for detection and quantification of human papillomavirus in archival cervical cancer tissue. *J Clin Microbiol.* 2004;42:3758-65.
94. Aubin F, Humbey O, Guérrini JS, Mougin C, Laurent R. Cancers cutanés non mélaniques et papillomavirus humains. *Ann Dermatol Venerol.* 2003;130:1131-8.
95. Zur Hausen H. Papillomavirus infections: a major cause of human cancers. *Biochem Biophys Acta.* 1996;1288:55-78.
96. Masini C, Fuchs PG, Gabrielli F, Stark S, Sera F, Ploner M, et al. Evidence for the association of human papillomavirus infection and cutaneous squamous cell carcinoma in immunocompetent individuals. *Arch Dermatol.* 2003;139:890-4.
97. Feltkamp MC, Broer R, di Summa FM, Struijk L, van der Meijden E, Verlaan BP, et al. Seroreactivity to epidermodysplasia verruciformis-related human papillomavirus types is associated with nonmelanoma skin cancer. *Cancer Res.* 2003;63:2695-700.
98. Harwood CA, Suretheran T, Sasieni P, Proby CM, Bordea C, Leigh IM, et al. Increased risk of skin cancer associated with the presence of epidermodysplasia verruciformis human papillomavirus types in normal skin. *Br J Dermatol.* 2004;150:949-57.
99. Karagas MR, Nelson HH, Sehr P, Waterboer T, Strukel TA, Andrew A, et al. Human papillomavirus infection and incidence of squamous cell and basal cell carcinomas of the skin. *J Natl Cancer Inst.* 2006;98:389-95.
100. Bouwes Bavinck JN, Stark S, Petridis AK, Maruqq ME, ter Schegget J, Westendorp RG, et al. The presence of antibodies against virus-like particles of epidermodysplasia verruciformis-associated human papillomavirus type 8 in patients with actinic keratoses. *Br J Dermatol.* 2000;42:103-9.
101. Boxman IL, Russell A, Mulder LH, Bavinck JN, ter Schegget J, Green A, et al. Association between epidermodysplasia verruciformis-associated human papillomavirus DNA in plucked eyebrow hair and solar keratoses. *J Invest Dermatol.* 2001;117:1108-12.
102. Purdie KJ, Suretheran T, Sterling JC, Bell L, McGregor JM, Proby CM, et al. Human papillomavirus gene expression in cutaneous squamous cell carcinomas from immunosuppressed and immunocompetent individuals. *J Invest Dermatol.* 2005;125:98-107.
103. Hodges A, Smoller BR. Immunohistochemical comparison of p16 expression in actinic keratoses and squamous cell carcinomas of the skin. *Mod Pathol.* 2002;15:1121-5.
104. Ruas M, Peters G. The p16INK4a/CDKN2A tumor suppressor and its relatives (review). *Biochim Biophys Acta.* 1998;1378:115-77.
105. Pavey S, Conroy S, Russel T, Gabrielli B. Ultraviolet radiation induces p16 expression in human skin. *Cancer Res.* 1999; 59:4185-9.
106. Soufir N, Moles JP, Vilmer C, Moch C, Verola O, Rivet J, et al. P16 UV mutations in human skin epithelial tumors. *Oncogene.* 1999;18:5477-81.
107. Conscience I, Jovenin N, Coissard C, Lorenzato M, Durlach A, Grange F, et al. P16 is overexpressed in cutaneous carcinomas located on sun-exposed areas. *Eur J Dermatol.* 2006;16:518-22.
108. Nuovo GJ, Plaia TW, Belinsky SA, Baylin SB, Herman JG. In situ detection of the hypermethylation induced inactivation of the p16 gene as an early event in oncogenesis. *Proc Natl Acad Sci USA.* 1999;96:12754-9.
109. Masumoto N, Fujii T, Ishikawa M, Saito M, Iwata T, Fukuchi T, et al. P16 overexpression and human papillomavirus infection in small cell carcinoma of the uterine cervix. *Human Pathol.* 2003;34:778-83.
110. Hayashi M, Tamura G, Kato N, Ansai S, Kondo S, Motoyama T. Genetic analysis of cutaneous squamous cell carcinomas arising from different areas. *Pathol Int.* 2003;53:602-7.
111. Dokianakis DN, Koumantaki E, Billiri K, Spandidos DA. P53 codon 72 polymorphism as a risk factor in the development of HPV-associated non-melanoma skin cancers in immunocompetent host. *Int J Mol Med.* 2000;5:405-9.
112. Quenneville LA, Trotter MJ, Maeda T, Tron VA. P53-dependent regulation of heat shock protein 72. *Br J Dermatol.* 2002;146:786-91.
113. Padlewska K, Ramoz N, Cassonet P, Riou G, Barrois M, Majewski S, et al. Mutation and abnormal expression of the p53 gene in the viral skin carcinogenesis of epidermodysplasia verruciformis. *J Invest Dermatol.* 2001;117:935-42.
114. Jackson S, Storey A. E6 proteins from diverse cutaneous HPV types inhibit apoptosis in response to UV damage. *Oncogene.* 2000;19:592-8.
115. Pfister H, ter Schegget J. Role of HPV in cutaneous premalignant and malignant tumours. *Clin Dermatol.* 1997;15:335-47.
116. Jackson S, Harwood CA, Thomas M, Banks L, Storey A. Role of Bak in UV-induced apoptosis in skin cancer and abrogation by HPV E6 proteins. *Genes Dev.* 2000;14:3065-73.
117. Akgul B, García-Escudero R, Ghali L, Pfister HJ, Fuchs PG, Navsaria H, et al. The E7 protein of cutaneous human papillomavirus type 8 causes invasion of human keratinocytes into the dermis in organotypic cultures of skin. *Cancer Res.* 2005;65:2216-23.
118. Frazer IH, Thomas R, Zhou J, Leggatt GR, Dunn L, McMillan N, et al. Potential strategies utilised by papillomavirus to evade host immunity. *Immunol Rev.* 1999;168:131-42.
119. Zur Hausen H. Papillomaviruses causing cancer: evasion from host-cell control in early events in carcinogenesis. *J Natl Cancer Inst.* 2000;92:690-8.
120. Zanetti R, Rosso S, Martínez C, Navarro C, Schraub S, Sancho-Garnier H, et al. The multicentre south European study «Helios» I: skin characteristics and sunburns in basal cell and squamous cell carcinomas of the skin. *Br J Cancer.* 1996;73:1440-6.
121. Rosso S, Zanetti R, Martínez C, Torno MJ, Schraub S, Sancho-Garnier H, et al. The multicentre south European study «Helios» II: different sun exposure patterns in the aetiology of basal cell and squamous cell carcinomas of the skin. *Br J Cancer.* 1996;73:1447-54.
122. Rodríguez-Cerdeira C, Alba Menéndez A, Vilata Corell JJ. Desarrollo de nuevas vacunas frente al virus del papiloma humano. *Piel.* 2007;22:51-3.
123. Villa LL, Costa RL, Petta CA, Andrade RP, Aula KA, Giuliano AR, et al. Prophylactic quadrivalent human papillomavirus (types 6, 11, 16, and 18) L1 virus-like particle

- vaccine in young women: a randomised double-blind placebo-controlled multicentre phase II efficacy trial. *Lancet Oncol.* 2005;6:271-8.
124. Papaconstantinou HT, Lee AJ, Simmang CL, Ashfaq R, Gokaslan ST, Sokol S, et al. Screening methods for high-grade dysplasia in patients with anal condyloma. *J Surg Res.* 2005;127:8-13.
 125. Chin-Hong PV, Vittinghoff E, Cranston RD, Browne L, Buchbinder S, Colfax G, et al. Age-related prevalence of anal cancer precursors in homosexual men: the explore study. *J Natl Cancer Inst.* 2005;97:896-905.
 126. Arican O, Guneri F, Bilgic K, Karaoglu A. Topical imiquimod 5% cream in external anogenital warts: a randomized, double-blind, placebo-controlled study. *J Dermatol.* 2004;31:627-31.
 127. Umudum H, Rezanko T, Dag F, Dogruluk T. Human papillomavirus genome detection by in situ hybridization in fine-needle aspirates of metastatic lesions from head and neck squamous cell carcinomas. *Cancer.* 2005;105:171-7.
 128. Hoffmann M, Gottschlich S, Gorogh T, Lohrey C, Schwarz E, Ambrosch P, et al. Human papillomaviruses in lymph node neck metastases of head and neck cancers. *Acta Otolaryngol.* 2005;125:415-21.
 129. Licitra L, Perrone F, Bossi P, Suardi S, Mariani L, Artusi R, et al. High-risk human papillomavirus affects prognosis in patients with surgically treated oropharyngeal squamous cell carcinoma. *J Clin Oncol.* 2006;24:5623-5.
 130. Reimers N, Kasper HU, Weissenborn SJ, Stulzer H, Preuss SF, Hoffmann TK, et al. Combined analysis of HPV-DNA, p16 and EGFR expression to predict prognosis in oropharyngeal cancer. *Int J Cancer.* 2007;120:1731-8.
 131. Bahnassy AA, Zekri AR, Abdallah S, El-Shehaby AM, Sherif GM. Human papillomavirus infection in Egyptian esophageal carcinoma: correlation with p53, p21, mdm2, C-erbB2 and impact on survival. *Pathol Int.* 2005;55:53-62.
 132. Schlecht NF. Prognostic value of human papillomavirus in the survival of head and neck cancer patients: An overview of the evidence. *Oncol Rep.* 2005;14:1239-47.
 133. Kozomara R, Jovic N, Magic Z, Brankovic-Magic M, Minic V. p53 mutations and human papillomavirus infection in oral squamous cell carcinomas: correlation with overall survival. *J Craniomaxillofac Surg.* 2005;33:342-8.
 134. Middelkamp-Hup MA, Pathak MA, Parrado C, Garcia-Caballero T, Rius-Diaz F, Fitzpatrick TB, et al. Orally administered *Polypodium leucotomos* extract decreases psoralen-UVA-induced phototoxicity, pigmentation, and damage of human skin. *J Am Acad Dermatol.* 2004;50:41-9.