Sporotrichoid Cutaneous Infection by Mycobacterium Haemophilum in an AIDS Patient

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Abstract. We report a case of primary cutaneous infection by Mycobacterium haemophilum after the bite of an aquarium fish in a severely immunodepressed AIDS patient. Clinical features consisted in nodular and ulcerative lesions that followed a sporotrichoid pattern. Histological study of nodular lesions showed a granulomatous dermatitis with numerous acid-fast bacilli. The mycobacterium was identified 3 months later by genetic hybridization from a culture in solid medium. Combined therapy with isoniazid, rifampin, clarithromycin, ethambutol, amikacin and ciprofloxacin resulted in complete resolution of the lesions. Infection by Mycobacterium haemophilum is a rare mycobacteriosis that usually affects immunodepressed patients. The most common clinical manifestations are cutaneous lesions but the development of sporotrichoid nodular lymphangitis is exceptional.

Key words: Mycobacterium haemophilum, immunosuppression, sporotrichoid, HIV, AIDS.

Case Description

We present the case of a 37-year-old man with known human immunodeficiency virus (HIV)-positive status for the past 12 years, poor compliance with antiretroviral therapy, no check-up visits in the past year, and several previous admissions due to opportunistic HIV-associated infections. He was admitted to our hospital with ascending
lymphangitis in the upper right arm that had started 2 weeks earlier and had been followed by a chain of painful, inflammatory nodular lesions that appeared along the lymphatic path and that had become ulcerative and suppurative. The patient had an aquarium and reported having been bitten by a tropical fish (blue African cichlid from Lake Tanganyika) on the left index finger 2 months earlier. He reported pain in the left knee and spread of the skin lesions to the right limbs 5 days prior to admission; he had no fever or malaise.

Clinical examination revealed several subcutaneous erythematous nodules that moved when touched; there were visible sporotrichoid lesions at the bite site and along the extensor surface and inner side of the left arm (Figure 1); some of the lesions had ulcerated and were seeping purulent exudate (Figure 2). There were similar isolated skin lesions on the right arm and on both legs, and signs of arthritis in the left knee.

The tests performed on admission revealed a CD4+ count of 26 cells/µL, a viral load of 740,000 copies/mL, and unremarkable chest radiograph findings. The joint fluid obtained by arthrocentesis showed inflammatory characteristics. Both the sputum smear and culture were negative and the radiograph of the left knee was normal. Blood cultures were also negative. The microbiological cultures of the skin specimens were negative and numerous acid-fast bacteria were observed in both specimens (Figure 5). Polymerase chain reaction results were negative for Mycobacterium tuberculosis.

Broad-spectrum empirical therapy with isoniazid, rifampicin, ethambutol, amikacin, clarithromycin, and ciprofloxacin was administered during the first days of hospitalization; antiretroviral treatment, with tenofovir, lamivudine, and efavirenz, was reinstated and administered simultaneously, although it had to be interrupted after 8 days as the patient developed a severe skin rash as a reaction to efavirenz. Although the patient showed initial clinical improvement, he developed a paradoxical inflammatory reaction on the hands in the third week of therapy,
accompanied by fever and malaise. The antibiotics were maintained and the patient improved following the administration of short-acting corticosteroids, which brought the condition into remission. Three months later, an atypical mycobacterium was detected in an iron-replete culture medium; it was identified as *M. haemophilum* by amplification of specific DNA sequences (GenoType Mycobacterium CM/AS 12). Tests performed on the strain showed it to be resistant to first-line antituberculous drugs (isoniazid, rifampicin, ethambutol, and amikacin) and susceptible to ciprofloxacin, clarithromycin, cotrimoxazole, and tetracyclines.

Five months after admission to hospital, the patient continued to receive triple antibiotic therapy with ofloxacin, clarithromycin, and minocycline; tolerance was good and the skin lesions resolved gradually.

The patient stopped coming for check-up visits when he returned to his native Germany.

### Discussion

There has been growing interest in infections caused by atypical mycobacteria, including *M. haemophilum*, in recent years due largely to the HIV pandemic and the increasing number of patients receiving immunosuppressive therapy.

*M. haemophilum* is a slow-growing, aerobic, nonchromogenic acid-fast bacterium that was first discovered in 1978 by Sompolinsky et al. in a 51-year-old Israeli woman with Hodgkin disease and generalized skin ulcers. The authors reported that the bacterium had a low incubation temperature, similar to that of *Mycobacterium marinum* and *Mycobacterium ulcerans*, and that culture media had to be enriched with iron to promote growth, distinguishing it from other atypical mycobacteria. Since then, fewer than 150 cases of infection due to *M. haemophilum* have been described in the medical literature. It typically presents as a superficial infection in immunosuppressed adults and can lead to potentially serious systemic infections. The most common clinical manifestations are a broad spectrum of skin lesions ranging from localized disease to systemic disease with cutaneous dissemination. Skin lesions tend to be multiple and can present as erythematous papules, plaques, nodules, necrotic abscesses, or chronic ulcers. They generally affect the limbs, and joints in particular, although sporotrichoid spread is not characteristic of *M. haemophilum* infection. Purpuric and annular lesions have also been described. Skin lesions typically evolve from papules to asymptomatic pustules, and eventually very painful deep-seated ulcers. Less common clinical presentations include bacteremia, pneumonia, lymphangitis, arthritis, tenosynovitis, and osteomyelitis. Patients with cutaneous and articular manifestations have a more favorable prognosis than those with pulmonary involvement. Histologically, there are no differences between infections caused by *M. haemophilum* and those caused by other atypical mycobacteria. Microscopic examination of skin lesions reveals deep granulomatous dermatitis, with or without caseating necrosis; the granulomas are often poorly differentiated and frequently affect the panniculus adiposus in patients with AIDS. The histopathological pattern is strongly influenced by the time since onset, particularly in immunosuppressed patients. Early lesions include an inflammatory infiltrate that is composed mostly of neutrophils or a mixture of neutrophils with granulocytes, lymphocytes, monocytes, and occasionally, multinucleated giant cells and foamy macrophages. Subsequent lesions are characterized by a granulomatous inflammatory reaction. Acid-fast bacteria are often detected in immunosuppressed patients using specific staining techniques. Microbiological diagnosis is difficult. Generally speaking, biopsies have a greater diagnostic yield than cotton swabs or pads. Under direction examination, *M. haemophilum* turns red following Ziehl–Neelsen staining. Although this

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**Figure 4.** Granulomas without necrosis, formed by epithelial lymphocytes and histiocytes. Hematoxylin–eosin, original magnification ×100.

**Figure 5.** Acid-fast bacteria.
technique detects the presence of acid-fast bacteria, it does not distinguish between different species, meaning that a culture is always necessary. The nutritional requirements for culture of M haemophilum distinguish it from other atypical bacteria. Unlike its homologues, M haemophilum grows specifically in a standard mycobacterial culture medium (solid or liquid) that has been enriched with iron.\textsuperscript{2,13,14} An excellent low-cost alternative is the use of a culture with fresh, hemolyzed blood.\textsuperscript{15} M haemophilum requires a lower temperature (between 30ºC and 32ºC) and a longer incubation period than other atypical mycobacteria to achieve optimal growth. The bacterium is probably not detected as often as it should be because of its growth requirements and the fact that successful isolation depends greatly on the method used.\textsuperscript{2,16} Bacteria grown in culture are identified on the basis of standard biochemical tests, growth characteristics, the chromatographic analysis of membrane lipids, and in more recent years, molecular genetic studies of mycobacterial DNA. An increasing number of mycobacterial species are being identified thanks to this new technique, and recent advances in the field have provided essential information on the clinical spectrum of the diseases they cause. They have also paved the way for the detection of bacteria in biological samples or in solid or liquid media using methods such as polymerase chain reaction.\textsuperscript{3,15,17}

Since the first case of infection due to M haemophilum was published in 1978,\textsuperscript{2,18,21} there have been reports of skin lesions caused by this bacterium in patients who have undergone kidney transplants,\textsuperscript{18-21} bone marrow transplants,\textsuperscript{22} heart transplants,\textsuperscript{23} as well as in patients with lymphoma,\textsuperscript{24,25} rheumatoid arthritis,\textsuperscript{26} and Crohn disease.\textsuperscript{1,2}

The first report of an infection due to M haemophilum in a patient with AIDS was published in 1987, when Males et al.\textsuperscript{29} reported isolating the bacterium from the joint fluid of a 32-year-old man with septic arthritis and skin lesions. An additional 2 cases involving disseminated cutaneous infection in patients with AIDS were published a year later.\textsuperscript{3,28} By 2001, 90 cases of M haemophilum infection had been reported worldwide. The majority of the patients had developed skin and soft tissue manifestations; 72 were adults and they all were all immunosuppressed due to AIDS or immunosuppressive therapy received prior to organ transplantation.\textsuperscript{2,22,25,29-31} Nonetheless, several cases of cervicofacial lymphadenitis due to M haemophilum have been reported in immunocompetent children,\textsuperscript{3,13,22} with a clinical picture similar to that produced by other mycobacteria (Mycobacterium avium complex, M tuberculosis, and Mycobacterium scrofulaceum). There have also been anecdotal reports of infection in otherwise healthy adults with skin lesions.\textsuperscript{7,13}

Because relatively few cases of the disease are seen and the bacterium is difficult to isolate in the laboratory, the true epidemiological impact of M haemophilum infection is unknown.\textsuperscript{1,3,13}

The environment is probably the reservoir of infection, as is the case with other atypical mycobacteria, but little is known about the natural habitat of the bacterium or the mechanism by which it infects humans. One recent study reported a series of cases in cities near large expanses of water (lakes and oceans) although it has not been possible to isolate the bacterium from aquatic environments.\textsuperscript{31} Nosocomial human-to-human transmission was also suggested as a possible cause of infection following an outbreak of cases in several hospitals in the metropolitan areas of New York between 1989 and 1991. The only epidemiological link detected between the patients, however, was that they were all immunosuppressed.\textsuperscript{1,2,23,31,32} Of the few series that have been published to date, none has been able to establish a link between a specific exposure scenario and transmission.\textsuperscript{1,2,30,31}

M haemophilum has a very different antimicrobial susceptibility pattern to that of other atypical mycobacteria. The studies published to date have all used different susceptibility analysis methods\textsuperscript{3,13,22,29,30,31,32} and there are no established antibiotic protocols for treating infected immunosuppressed patients. M haemophilum is resistant to first-line antituberculous drugs (isoniazid pyrazinamide and ethambutol) and has a susceptibility of just 56% to rifampicin.\textsuperscript{10}

The bacterium is, however, susceptible to other antibiotics such as ciprofloxacin, clarithromycin, and rifabutin, all of which are drugs used to treat infections caused by other atypical mycobacteria.\textsuperscript{1,2,30,31} It occasionally shows susceptibility to tetracyclines and cotrimoxazole.\textsuperscript{1} In a series published by Bernard et al.\textsuperscript{36} involving 17 isolates from 12 immunosuppressed patients, the most effective antibiotics against M haemophilum were rifamycins, macrolides, quinolones, clofazimine, and amikacin. The authors also noted that 2 patients with progressive or recurrent disease developed resistance to rifamycins during treatment. There have been occasional reports of a lack of correlation between in vitro susceptibility and clinical response to treatment.\textsuperscript{37}

Although there are no standard protocols for susceptibility studies designed to determine minimum inhibitory concentrations of antimicrobial agents against M haemophilum, clinical experience indicates that antibiotic therapy in association with clarithromycin, ciprofloxacin, and rifampicin or rifabutin is a reasonably adequate empirical treatment option.\textsuperscript{2,38} In patients with more serious disease, including lung disease, the addition of amikacin, possibly in combination with doxycycline or minocycline, is recommended.\textsuperscript{2} Clinical response to treatment is varied, even when the above antibiotics are used. Prolonged maintenance therapy lasting months or even years with several drugs is generally necessary, particularly in patients with sustained immunosuppression.\textsuperscript{10}

Given that M haemophilum is not normally pathogenic in immunocompetent individuals, the most efficient
treatment in some cases seems to be reconstitution of the immune system in hosts with potentially reversible immunosuppression.18

We have presented a case of sporotrichoid cutaneous infection due to Mycobacterium haemophilum in an immunosuppressed patient. In our review of the literature we found no reports of sporotrichoid nodular lymphangitis secondary to M. haemophilum. While it is true that this clinical form is well documented for a number of infections caused by other atypical bacteria such as M. marinum, and Mycobacterium chelonae,19 to the best of our knowledge, this is the first time that it has been described for M. haemophilum.

Conflicts of Interest
The authors declare no conflicts of interest.

References