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REVIEW

[Translated article] An Update on EBV-related Cutaneous Lymphoproliferative Disorders: a Systematic Review



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Received 20 February 2024; accepted 10 September 2024 Available online 10 February 2025

KEYWORDS

Systematic review; Epstein Barr Virus-Positive B-cell lymphoproliferative disorders; Epstein Barr Virus-Positive B-cell lymphomas; Epstein Barr Virus-Positive T-cell lymphoproliferative disorder Abstract Epstein Barr virus (EBV) positive lymphoproliferative disorders (LPD) with cutaneous involvement include a series of rare entities that go from indolent processes to aggressive lymphomas. B-cell EBV+ LPD mainly affect immunocompromised patients while T-cell EBV+ LPD are more prevalent in specific geographic regions such as Asia, Central America, and South America. Since the latest WHO-EORTC classification of cutaneous lymphomas in 2018, significant changes have been included in the new classifications of hematological malignancies. This systematic review summarizes the main clinical, histological, immunophenotypic and molecular characteristics of B- and T-cell EBV+ LPD that may compromise the skin at diagnosis. B-cell EBV+ LPD include primary cutaneous lymphomas such as EBV-Mucocutaneous Ulcer, as well as systemic lymphomas affecting the skin at diagnosis that may present such as lymphomatoid granulomatosis (LG), EBV diffuse large B cell lymphoma, NOS, plasmablastic lymphoma (PBL), extracavitary primary effusion lymphoma (EC-PEL) EBV+, EBV-positive polymorphic B cell LPD, and post-transplant lymphoproliferative disorders (PTLD). Regarding T-cell EBV+ LPD, most of these entities are categorized within T/NK-cell lymphoproliferative processes and lymphomas of childhood, including extranodal T/NK lymphoma, and even more exceptional forms such as EBV-positive T-cell centrofollicular lymphoma and intravascular T/NK-cell lymphoma. Diagnosis is based on integrating the clinical, histological, immunohistochemical, and genetic criteria discussed throughout this article. Differential diagnosis is a challenge for dermatologists and pathologists, so having scientific evidence available in this field is of paramount importance because overtreatment must be carefully avoided.

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DOI of original article: https://doi.org/10.1016/j.ad.2024.09.018 * Corresponding author.

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https://doi.org/10.1016/j.ad.2025.02.009

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PALABRAS CLAVE

Revisión sistemática; Procesos linfoproliferativos B virus de Epstein Barr positivo; Linfomas B virus de Epstein Barr positivo; Proceso linfoproliferativo T virus de Epstein Barr positivo

Actualización en procesos linfoproliferativos cutáneos relacionados con el virus de Epstein Barr: revisión sistemática

Resumen Los procesos linfoproliferativos (PLP) positivos para el virus de Epstein Barr (VEB) con afectación cutánea son una serie de entidades poco frecuentes que engloban desde procesos indolentes a linfomas agresivos. Los procesos linfoproliferativos B (PLP B) afectan principalmente a los pacientes inmunocomprometidos y los procesos linfoproliferativos T (PLP T) son más frecuentes en determinadas regiones geográficas como Asia, América Central y Sudamérica. Desde la última clasificación de la clasificación de consenso común de la Organización Mundial de la Salud y de la Organización Europea para la Investigación y el Tratamiento del Cáncer (WHO/EORTC) de los linfomas cutáneos en 2018 se han producido cambios significativos para estas entidades en las nuevas clasificaciones de las neoplasias hematológicas. En esta revisión sistemática se incluyen las principales características clínicas, histológicas, inmunofenotípicas y moleculares de los PLP B y T VEB⁺ que pueden afectar a la piel al diagnóstico. Entre los PLP B se incluyen linfomas primarios cutáneos como la úlcera mucocutánea positiva para el virus de Epstein Barr (UMC VEB⁺) y linfomas sistémicos que pueden presentarse con afectación cutánea como la granulomatosis linfomatoide (GL), el linfoma difuso de células grandes B asociado al virus de Epstein Barr, No especificado (LCGBD VEB⁺, NOS) el linfoma plasmablástico (LPB), el linfoma primario de cavidades con presentación extracavitaria (EC-PEL) VEB⁺, el proceso linfoproliferativo polimorfo B VEB⁺ o los PLP postrasplante (PLPPT). Dentro de los PLP T, la mayoría están englobados dentro de los PLP y linfomas de células T/NK de la infancia, así como el linfoma T/NK extranodal y formas aún más excepcionales como el linfoma T de célula centrofolicular VEB⁺ y el linfoma intravascular de célula T/NK. El diagnóstico diferencial de estas entidades es un reto para clínicos y patólogos, por lo que disponer de evidencia científica de calidad en este campo es de gran importancia para evitar el sobretratamiento.

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Results

Introduction

The classification of lymphomas has evolved significantly in recent years. Since the 2018 consensus classification by the World Health Organization (WHO) and the European Organization for Research and Treatment of Cancer (EORTC),¹ not one but two new classifications/expert consensus documents on hematolymphoid neoplasms^{2,3} have recently been published, along with the newly released WHO classification of cutaneous neoplasms.⁴ Advances in diagnostic techniques, and new discoveries on the pathogenesis and genetics of these entities, have brought substantial changes to their classification and the definition of some of them. This systematic review focuses on Epstein Barr virus (EBV)-related lymphoproliferative disorders (LPDs) and B- and T-cell lymphomas that can present with cutaneous involvement, also addressing the role of EBV in lymphomagenesis. The methodology of the review can be found in Appendix,^{5,101} as well as in Fig. 1, which provides a summary of the systematic review methodology. Additionally, Appendix includes the main clinicopathological characteristics of the entities discussed in this article. Finally, Appendix summarizes nomenclature equivalences for these processes across various current and predecessor classifications. For readability purposes, the text refers to entities by their names in the international consensus classification (ICC).

The role of Epstein Barr virus in lymphomagenesis

Over the past few decades, efforts have been made to clarify the role of certain viral infections in the emergence of cutaneous lymphomas.⁷ Epstein Barr virus (EBV) was the first human virus associated with tumors, discovered in 1964 in Burkitt's lymphoma.⁸ The EBV is a gamma herpesvirus, widely prevalent, infecting over 90% of the global population.⁹ Following primary infection, EBV persists for life in memory B cells.¹⁰ Most infected individuals control the infection through cytotoxic immune responses by NK cells or CD8⁺ T lymphocytes. Only a small subset develops chronic EBV infection and associated diseases, more commonly in the presence of immunodeficiency, genetic predisposition, or environmental factors.¹⁰ EBV exhibits various types of latent infection, largely determined by the host's immune status. These are characterized by the limited expression of viral genes, which vary across EBV-associated tumors (Table 1).

The precise mechanisms through which latent EBV infection initiates lymphoid cell transformation and neoplastic progression remain unclear, though several predisposing factors exist. For each tumor entity and location, differences are observed in gene expression profiles, tumor metabolism, signal transduction, immune evasion mechanisms, and the composition of the tumor microenvironment depending on EBV association.¹¹

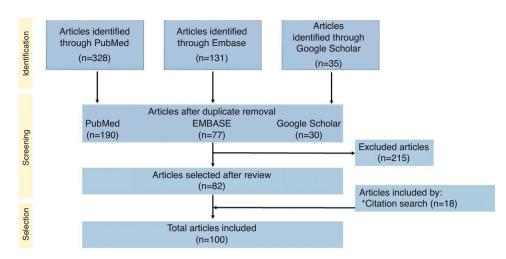


Figure 1 Flowchart of the studies included in the review. Selection process of publications following PRISMA 2020.

Table 1	EBV latencies and their relationship with EBV	⁺ B and T lymphoproliferative disorders and immune status.
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Latency	Markers	Disease	Immunocompromised
0	EBER, BARTS, miRs	PBL ^a	Yes
1	EBER,	PBL	Yes
	EBNA1	EC-PEL	Yes
П	EBER, EBNA1, LMP1, LMP2A,	EBV⁺ MCU	Yes
	LMP2B	EBV+ DLBCL, NOS	No
		LG ^a	Yes
		PBL ^a	Yes
		PTLD ^a	Yes
		HVLPD	No
		Severe mosquito bite allergy	No
		Extranodal NK/T-cell lymphoma	Yes
Ш	EBER, EBNA1, EBNA2, EBNA3A,	EBV⁺ MCU	Yes
	EBNA3B, EBNA3C, EBNA leader	LG	Yes
	protein (LP), LMP1, LMP2A,	PTLD	Yes
	LMP2B	EBV ⁺ -DLBCL	Yes

In EBV latency states, up to 6 EBV nuclear antigens (EBNA1, 2, 3A, 3B, 3C, and LP), 3 latent membrane proteins (LMP1, 2A, and 2B), and 2 EBV-encoded RNAs (EBER1 and 2) can be expressed. This table shows the relationship of these markers with latency states, associated lymphoproliferative disease, and the presence or absence of immunocompromised status.

^a Lymphoproliferative processes that may present other types of latencies and/or markers less commonly.

EBER: EBV-encoded small RNA; EBNA: EBV nuclear antigen; BARTS: BamHI fragment A rightward transcript; LMP: latent membrane protein; miRs: microRNAs; EBV⁺ MCU: EBV⁺ mucocutaneous ulcer; EBV⁺-DLBCL, NOS: EBV⁺ diffuse large B-cell lymphoma, not otherwise specified; LG: lymphomatoid granulomatosis; PBL: plasmablastic lymphoma; EC-PEL: extracavitary primary effusion lymphoma; PTLD: post-transplant lymphoproliferative disorder; HVLPD: hydroa vacciniforme-like lymphoproliferative disorder.

Notably, EBV⁺ lymphomas may develop multiple immune evasion mechanisms, making them potential candidates for immunotherapy. Treatment approaches include programmed cell death protein 1 (PD-1)/programmed deathligand 1 (PD-L1) antibodies, small molecules targeting EBV latency gene products, cellular vaccines, and even chimeric antigen receptor T (CAR-T) cell therapies targeting EBV antigens.^{11,12}

EBV-related lymphoproliferative disorders and B-cell lymphomas with cutaneous involvement

These entities comprise a clinicopathological spectrum ranging from indolent, self-limited processes to highly aggressive ones. Immunosuppression often plays a critical role in their pathogenesis.

EBV⁺ mucocutaneous ulcer

First described in 2010 in a series of 26 patients,¹³ it is the only entity classified as a primary cutaneous LPD and is now recognized as a definitive entity in the latest classifications. Specifically, the 2022 ICC classification³ specifies that the condition should involve single lesions. For patients with \geq 2 lesions, terms such as ''EBV⁺ polymorphic B-cell LPD'' or, when appropriate based on clinicopathological characteristics, EBV-related diffuse large B-cell lymphoma, Not Otherwise Specified (EBV⁺-DLBCL, NOS), are preferred.

Epidemiology and pathogenesis. This condition is more common in women,¹⁴ with a mean age of 66.4 years.¹⁵ It occurs in the context of loss of control over latent EBV infection due to various types of immunosuppression: age-related immunosenescence caused by T-cell repertoire loss and dysfunction,¹⁶ or iatrogenic immunosuppression in autoimmune diseases, solid organ transplantation, allogeneic hematopoietic stem cell transplantation (allo-HSCT), or HIV infection.¹⁷ Methotrexate is the drug most frequently associated with this LPD, followed by azathioprine, cyclosporine, imatinib, tacrolimus, tumor necrosis factoralpha inhibitors, and mycophenolate.¹⁷ Chronic mucosal inflammation or inflammatory bowel disease are additional risk factors.¹⁸

Clinical characteristics. It presents as a solitary ulcer on the skin or mucosa (Fig. 2). Clinical course is indolent, with spontaneous resolution of lesions occasionally observed.¹⁴ Patients typically exhibit very low or undetectable EBV DNA loads in peripheral blood, aiding in the differential diagnosis from EBV⁺-DLBCL, NOS.¹⁹ Rare cases with intermittent-recurrent courses without progression have been described.²⁰

Histology and molecular features. Biopsies show superficial ulcers with epidermal or mucosal acanthosis, sometimes with pseudoepitheliomatous changes.¹³ The inflammatory and tumoral infiltrate at the ulcer base is heterogeneous, with plasmacytoid Hodgkin-like apoptotic cells and necrosis, highly characteristic of these lesions¹⁴ (Fig. 2). Furthermore, CD3⁺ T-cell rings are typically found at the base. Hodgkin-like immunoblasts show variable PAX5⁺ intensity, and positivity for OCT2⁺, MUM1⁺, BOB1⁺, and CD45^{+/-}. CD20 expression is partial or absent in up to 33% of cases.¹³ These cells are typically CD30⁺ (Fig. 2), often co-expressing CD15 in a significant subset.¹⁹ EBER positivity is seen in Hodgkinlike cells, smaller lymphocytes, and occasionally adjacent epithelial cells.^{6,14} PDL1 expression has been reported in tumor cells in some series.⁶ B-cell clonality is detected in <50% of patients, while T-cell clonality or oligoclonality is common due to immunosenescence and other immune defects.14

Updates on treatment. Many cases resolve spontaneously or after discontinuing immunosuppression. In elderly patients without other immunosuppressive factors, good responses to IV rituximab monotherapy have been described.¹⁴ Other published treatments include polychemotherapy regimens, such as the combination of rituximab with cyclophosphamide, doxorubicin, vincristine sulfate, and prednisone (R-CHOP), or radiotherapy.¹⁸

Diffuse large B-cell lymphoma associated with Epstein Barr virus, not otherwise specified

This entity was first described in 2003 in a series of non-immunosuppressed patients over 60 years old, characteristically presenting with predominant extranodal involvement.²¹ These patients appeared to have a worse prognosis compared with EBV negative-DLBCL cases (EBV⁻). In the 2008 WHO classification, it was included as a provisional entity under the name ''EBV⁺-DLBCL of the elderly''.²² Over time, cases in younger patients were reported, leading to the name change to ''EBV⁺-DLBCL, NOS'' in the 2016 WHO classification.

Epidemiology and pathogenesis. Its prevalence is higher in Africa, Asia, and Latin America. It has been described across a wide age range but is more common in individuals older than 50 years and often presents with extranodal involvement (40% of cases). Unlike older patients, younger patients (<45 years) more frequently exhibit nodal disease and tend to have a better prognosis.¹⁴

Clinical features. Cutaneous lesions typically consist of plaques, nodules, or tumors, generally non-ulcerated, located on the lower extremities¹⁹ (Fig. 3). Overall, advanced disease is common at diagnosis, with high EBV viral loads in peripheral blood (PB).²³ A recent systematic review drew a comparison between 17 cases of this lymphoma with cutaneous presentation and 21 cases of primary cutaneous DLBCL, leg type (LT). It highlighted clinicopathological differences between the 3 entities, as well as a worse prognosis for EBV+ DLBCL, NOS (median overall survival, 32 months vs 88 months for LT).²⁹ Patients with EBV+ DLBCL, NOS are significantly older at diagnosis, more likely to have non-nodular lesions, and exhibit multiple skin lesions and locations.²⁹

Histology and molecular features. A spectrum of morphological presentations exists, ranging from monomorphic cases with sheets of atypical large lymphocytes and no reactive inflammatory cells, to polymorphic cases with reactive inflammatory infiltrates and scattered atypical large lymphocytes (Fig. 3). In skin, monomorphic infiltrates are the most common ones.²⁴ The prognostic significance of histology is debated. In adults, it does not appear to have prognostic implications,¹⁴ whereas in younger patients (<45 years), the polymorphic pattern suggests a better prognosis.¹⁴ Additionally, approximately 22.7% of cutaneous cases show an angiocentric pattern. Phenotypically, most atypical lymphocytes are positive for CD20, CD79a, MUM1, and CD30, with variable expression of BCL2 and BCL6.²⁴ CD30 (Fig. 3) and EBER positivity (Fig. 2F) are key immunophenotypic differences from LT.²⁴ PD-L1 and PD-L2 are frequently expressed in younger vs older patients, suggesting an immune evasion mechanism.²⁵

Unlike EBV-negative DLBCL, EBV⁺-DLBCL, NOS²⁶ often features mutations in the nuclear factor kappa beta (NF- κ B) pathway, Wingless (WNT) signaling pathway, and interleukin 6/Janus kinase-signal transducers and activators of transcription (IL6/JAK/STAT) pathway.²⁷ Similarly, integrated whole-genome sequencing and targeted amplicon sequencing clearly differentiate this tumor type from EBV⁻DLBCL due to frequent mutations in ARID1A (45%), KMT2A/KMT2D (32/30%), ANKRD11 (32%), and NOTCH2 (32%).²⁷

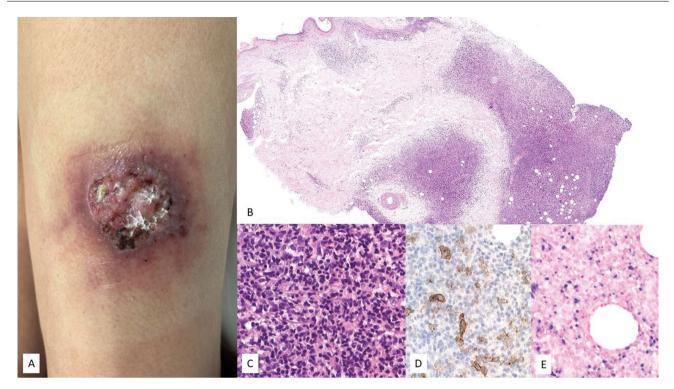


Figure 2 EBV+ mucocutaneous ulcer (EBV+-MCU). (A) Clinical image showing an isolated ulcer with necrotic areas and surrounding erythema located on the anterior aspect of the left thigh. (B) At low magnification, a dermal infiltrate with intense involvement of the deep portion and a nodular pattern is observed (H&E, $2\times$). (C) At higher magnification, the infiltrate is heterogeneous, with atypical elements of Sternberg-like habit (H&E, 20x). (D) Part of the infiltrate tests positive for CD30 ($20\times$); and (E) positive for EBER ($20\times$).

Treatment updates. These cases respond poorly to typical chemotherapy regimens like R-CHOP. Some clinical trials with anti-CD30 antibodies, such as brentuximab combined with rituximab, cyclophosphamide, doxorubicin, and prednisone (BV-R-CHP), or anti-CD79b antibodies like polatuzumab vedotin combined with R-CHP, have shown promising results. Polatuzumab R-CHP could become the treatment of choice for these patients.^{28,29}

Lymphomatoid granulomatosis

Lymphomatoid granulomatosis (LG) is a rare EBV-related B-cell LPD first described in 1972. Initially considered a peripheral T-cell lymphoma due to the predominance of accompanying T-lymphocytes,³⁰ it is now recognized as a Bcell lymphoma.³¹ This lymphoma typically affects the lungs, with less frequent involvement of the central nervous system (CNS), skin, kidneys, or liver.¹⁸

Epidemiology and pathogenesis. It shows a slight male predominance (2:1) and typically presents in the 4–6th decades of life, being very rare in children.¹⁸ Its pathogenesis is unclear, but several causes have been proposed, including the oncogenic potential of EBV. It has also been associated with chronic autoimmune diseases or immunodeficiencies, whether congenital or acquired, such as post-transplant LPDs (PTLPDs) or iatrogenic LPDs related to immunosuppressive drugs such as azathioprine, methotrexate, or imatinib.^{18,19}

Clinical features. Disease is considered extranodal, with nodal or bone marrow involvement being exceptional.

Typical presentations include cough, dyspnea, and chest pain, sometimes accompanied by systemic symptoms such as fever, myalgias, malaise, or weight loss, and peripheral neuropathy. Skin lesions may appear at any stage of the disease and have been reported as the initial sign in up to one-third of patients. Clinical and morphological variability in the described lesions exists. The most common presentation is erythematous nodules-sometimes subcutaneous-on the trunk and extremities, mimicking panniculitis. Other presentations include multiple indurated plaques, lesions resembling lichen sclerosus, or even those mimicking nasal NK/T-cell lymphoma with zone ulceration and diffuse crateriform nodules.^{32,33} Historically associated with a poor prognosis, the introduction of novel treatments has increased the life expectancy of these patients.^{19,34} Histology and molecular features. The histology of cuta-

Histology and molecular features. The histology of cutaneous lesions can differ from that seen in other organs.³² In the skin, a lymphocytic or lymphohistiocytic infiltrate with a variable presence of multinucleated giant cells is typically found, presenting as panniculitis with poorly structured granulomas. These infiltrates have a characteristic angiocentric or perivascular distribution. In other cases, cutaneous lesions resemble those found in organs such as the lungs, appearing as an angiodestructive process with fibrinoid necrosis and perivascularly arranged CD20+ and EBER+ immunoblasts. EBV is harder to detect in cutaneous lesions, and biopsying non-ulcerated lesions is preferable since necrosis hinders virus detection.³² Accompanying T lymphocytes are abundant and typically express CD8 and

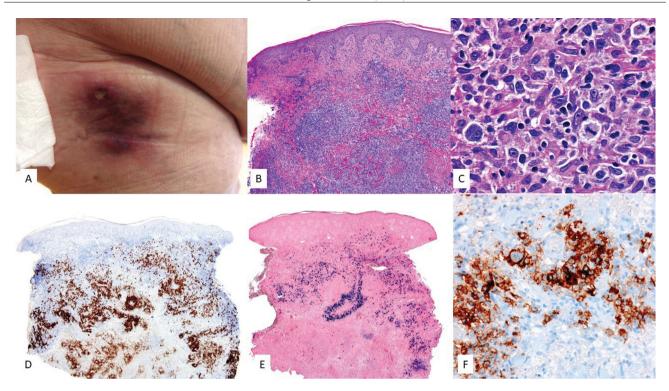


Figure 3 EBV-related diffuse large B-cell lymphoma, unspecified. (A) Clinical image showing an indurated erythematous-violaceous plaque infiltrating the upper region of the right thigh in an elderly patient. (B) At low magnification, a dermal infiltrate in the superficial and deep portions with a nodular pattern is observed (H&E, $2\times$). (C) At higher magnification, the infiltrate looks heterogeneous with the presence of large cells of atypical morphology and Sternberg-like habit (H&E, $20\times$). These cells are (D) positive for CD30 ($2\times$), (E) EBER ($2\times$), and (F) CD20 ($2\times$).

cytotoxic markers. Large cells are positive for CD30 in up to 50% of cases, while CD15 is characteristically negative.³²

A grading scheme for cases based on EBV^+ cell counts exists.³⁵ However, grading is not recommended in the skin, as different lesions may have varying numbers of EBER+ cells.

Clonal rearrangement is present in approximately 25% of cases, with variability depending on the grade (only 8% of grade 1 lesions are clonal vs 69% of grade 3 lesions).³⁶

Treatment updates. Currently, there are no consensus guidelines on the management of these patients. For iatrogenic cases associated with immunosuppression and low-grade lesions, reducing immunosuppression or discontinuing the immunosuppressive drug is recommended.¹⁸ Immunotherapies such as IFN-alpha or immunoglobulins have been used to promote the immune system antiviral action.³⁷ Grade 3 lesions are treated with polyimmunochemotherapy regimens (R-CHOP) similar to those used for EBV⁺-DLBCL, NOS. AHSCT may also be considered.³⁸

Plasmablastic lymphoma

This entity was first described under the name ''plasmablastic lymphoma'' (PBL) back in 1997^{39} and was included in the 2001 WHO classification as an aggressive and rare variant of DLBCL.⁴⁰ Primary cutaneous presentation of PBL (pcPBL) is extremely rare, with the first cases being reported between 2004 and 2005.^{41,42}

Epidemiology and pathogenesis. PBL has traditionally been considered an HIV-related disease, accounting for up to 2.6%

of HIV-associated lymphomas.^{43,44} Currently, PBL is also recognized as being associated with other immunosuppressive states.

The prognosis for this type of lymphoma is quite poor, despite the administration of chemotherapy treatments. Notably, the presence of EBV has been associated with a better prognosis compared to cases where EBV detection tested negative.^{44,45} In cases of PBL with cutaneous involvement only, prognosis is more favorable.^{46,47}

Clinical features. The most common presentation is extranodal, primarily in the oral cavity, followed by the GI tract. Less frequent are cases with nodal, pulmonary, nasal cavity, or cutaneous involvement.^{42,44,48}

At skin level, PBL typically manifests on the lower extremities as one or multiple erythematous-violaceous nodules with a tendency to ulcerate.^{47,48} Less common cutaneous presentations include recurrent scrotal abscesses,⁴⁹ enterocutaneous fistulas,⁵⁰ or perineal ulcers.⁵¹ Approximately half of the cases with cutaneous symptoms already present with systemic disease at diagnosis.⁴⁸

Histology and molecular features. Most cases are characterized by a diffuse infiltrate throughout the dermis, composed of plasmablasts.^{42,43}

Immunohistochemically, B-cell markers and CD45 are typically negative. Conversely, these cells are positive for CD138, CD38, MUM1/IRF4, and PRDM1/BLIMP1.^{42,43,48} EBER tests positive in more than 60% of cases,^{42,48} and proliferation index is usually very high (Ki67 > 90%). Occasionally,

aberrant expression of T-cell markers can occur, complicating differential diagnosis.^{42,52}

Treatment updates. There is no consensus for its treatment. Various chemotherapeutic regimens are commonly used, with CHOP being one of the most frequently employed regimens.⁵³ Other combinations have also been tested, including regimens based on etoposide phosphate, prednisone, vincristine sulfate, cyclophosphamide, and doxorubicin hydrochloride (EPOCH); regimens based on cyclophosphamide, vincristine sulfate, doxorubicin hydrochloride, dexamethasone, methotrexate, and cytarabine (HyperC-VAD); or regimens alternating high-dose cyclophosphamide, vincristine, doxorubicin, and methotrexate with ifosfamide, etoposide, and cytarabine (CODOX-M/IVAC),^{43,53} with variable response. Currently, clinical trials are being conducted with bortezomib, ganciclovir, and CAR-T therapies.⁴³

In the case of primary pcPBL, surgical excision along with adjuvant radiotherapy can be considered as a more conservative treatment, or chemotherapy in the presence of multiple lesions.⁴⁵

Primary cavity lymphoma of extracavitary presentation, EBV⁺

This entity was first recognized in the WHO classification of 2001^{54,55} as a rare and aggressive type of non-Hodgkin B-cell lymphoma defined by the presence of human herpesvirus type 8 (HHV8). Coinfection with EBV is relatively common.⁵⁵ *Epidemiology and pathogenesis*. This lymphoma occurs more frequently in people with HIV but can also arise in other states of immunocompromise, such as elderly immunosenes-cent patients or solid organ transplant recipients.^{2,56} HIV⁺ or elderly patients are typically HHV8⁺ and EBV⁺.⁵⁶

Clinical features. Classically, this lymphoma affects body cavity effusions such as pleura, peritoneum, and pericardium. However, there is a solid extracavitary (EC) variant involving nodal and extranodal sites, mainly the GI tract and skin.⁵⁶ In the skin, with fewer than 15 published cases, it shows as subcutaneous nodules,⁵⁷ panniculitis-like lesions,⁵⁸ or Kaposi sarcoma-like lesions (Fig. 4).⁵⁹

Histology and molecular features. It presents diffuse infiltrates of large, pleomorphic cells with immunoblastic or plasmablastic features (Fig. 4). By definition it is HHV8⁺ and expresses latency-associated nuclear antigen (LANA). Although these tumor cells lack B-cell markers and germinal center markers such as CD10 and BCL6, they test positive for plasma cell markers such as MUM1, BLIMP1, CD38, and CD138⁵⁶ (Fig. 4). MYC oncogene mutations are rare, ⁵⁶ unlike in PBL. EBV may be present.

Treatment updates. Polychemotherapy regimens like CHOP are used, along with antiretroviral therapy (ART), which is critically important, as poorer prognosis is observed in patients who do not use ART.⁵⁶

Other LPDs with cutaneous involvement

Post-transplant lymphoproliferative disorder with cuta-

neous involvement. From a biological standpoint, nearly all EBV-related LPDs can occur during the state of immunosuppression after transplantation.⁶⁰ The risk of developing these correlates with the level and duration of immunosuppression required for transplantation, as well as the recipient's age and EBV serostatus.¹⁹ EBV-naïve recipients who acquire the infection post-transplant are at the highest risk of developing PTLPD. Approximately 22% of PTLPD patients may have cutaneous involvement, most widely described after renal transplantation.⁶¹ Polymorphic PTLDs are, by definition, B-cell in origin and nearly all EBV⁺, while the term monomorphic can refer to various B- or T-cell lymphomas, which may or may not be EBV⁺. In the literature, cases of cutaneous PTLPDs present clinically as maculopapular lesions, nodules, or tumors, with or without associated ulceration.⁶² An unusual form of these cutaneous B-cell PTLPDs is EBV⁺ marginal zone lymphoma.⁶³

Polymorphic EBV⁺ **B-lymphoproliferative disorders, unspec***ified.* Introduced in the 2022 ICC classification,³ this term is proposed for B-lymphoid proliferations with or without known immunodeficiency that do not fit into recognized entities. It can also be used when diagnostic certainty is hampered by small or low-quality biopsy samples.

EBV-related T-lymphoproliferative disorders and lymphomas with cutaneous involvement

In the skin, EBV⁺ T- or NK-cell LPDs are very rare, collectively accounting for <2% of primary cutaneous lymphomas,¹ with higher incidence rates in Asian and Latin American populations, suggesting a possible genetic predisposition. Unlike EBV⁺ B-cell lymphomas, immunosuppression does not appear to play a significant role in the development of these lymphomas.⁶⁴

Childhood EBV-related T/NK-lymphoproliferative disorders and lymphomas

According to the 5th edition of the WHO classification² and the ICC of mature lymphoid neoplasms,³ 4 main groups are included within this family: hydroa vacciniforme-like lymphoproliferative disorder (HVLPD), severe mosquito bite allergy, chronic active EBV systemic disease (CAEBV), and systemic childhood EBV⁺ T-cell lymphoma. Notably, the first 3 have a risk of progressing to systemic lymphomas and/or hemophagocytic syndrome, making early diagnosis crucial. EBV viral load follow-up doesn't seem to distinguish between clinical forms or prognostic outcomes.⁶⁵ There is no standardized therapeutic approach for these entities. Sections below discuss those with cutaneous involvement.

Hydroa vacciniforme-like lymphoproliferative disorder. The concept and definition of HVLPD, introduced in the most recent classifications,^{2,3} have changed dramatically since this disease was first described in 1862.⁶⁶ Initially considered a rare idiopathic and photosensitive skin disorder, it is now recognized as an EBV⁺ T- and NK-cell lymphoproliferative disorder with a wide spectrum of clinical aggression and course.

Epidemiology and pathogenesis. While it is more prevalent among Asian or Latin American children and adolescents, isolated cases in adults and Caucasians have also been described.⁶⁷ Clonal rearrangements of the T-cell receptor (TCR) are frequent, though they have no prognostic impact. On the other hand, targeted genetic studies to identify driver mutations in this entity have been limited, but recent reports indicate mutations in STAT3, IKBKB, ELB, CHD7,

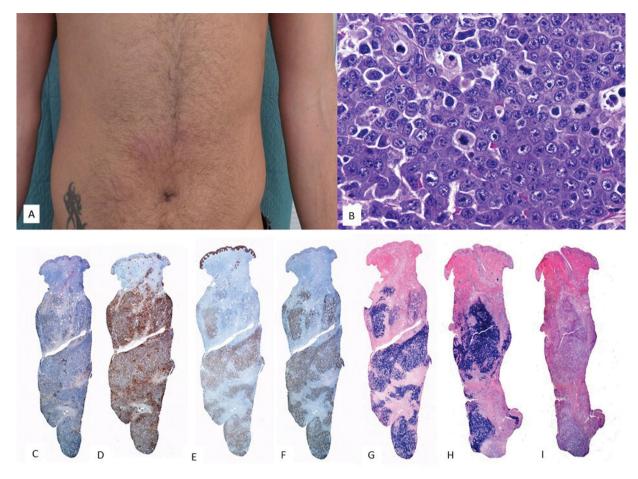


Figure 4 EBV⁺ extracavitary PEL. (A) Clinical image showing a subcutaneous mass with erythema on the periumbilical skin. (B) At higher magnification, the tumor infiltrate consists of large elements with a plasmablastic habit (H&E, $20 \times$). The immunohistochemical study reveals that these cells are (C) negative for CD20 (2×) and (D) negative for CD3 (2×). Conversely, they show positivity for (E) CD138 (2×), (F) HHV8 (2×), and (G) EBER (2×), and (H) monotypic expression for kappa light chains (2×). (I) Lambda light chains (2×).

and KMT2D.⁶⁸ These findings require validation in larger cohorts. Another case has been associated with a DOCK8 gene mutation.⁶⁹

Clinical features. Two clinical forms are recognized, different in disease course and prognosis.¹⁴ The classic form behaves as a benign, self-limiting disease that spontaneously remits during adolescence. It predominantly affects white patients and is characterized by the absence of systemic symptoms. Typical papulovesicular eruptions occur on sun-exposed skin and resolve with photoprotection.⁷⁰ The systemic form, initially described as "angiocentric Tcell lymphoma of childhood",⁷¹ is more common among Asians and Hispanics. Patients may develop lesions on both sun-exposed and non-exposed skin, with a more prolonged disease course and more severe skin lesions vs the classic form. These can be accompanied by systemic symptoms and progression to lymphoma. In one of the earliest published series, 4 male children aged 3-12 years presented with persistent facial edema, necrosis, and valioliform scars (Fig. 5).⁷² Subsequent series have identified a broader spectrum of clinical presentations.72-78 Atypical presentations, such as periorbital or ocular involvement with swelling, marked edema, and conjunctival congestion have been associated with very poor outcomes. 75,79,80 Another rare sign is oral mucosal involvement. 81

Histology. Various histological patterns have been described.⁷³ Some cases exhibit intraepidermal spongiotic vesiculation with necrosis of the epidermis. Others display periadnexal infiltrates or a combination of both (Fig. 5). Angiodestructive patterns are not always present and cannot be considered a sine qua non diagnostic feature.^{72,74} Neural infiltration by tumor cells has also been reported.⁷¹ Subcutaneous tissue involvement in some cases may lead to differential diagnosis with subcutaneous panniculitis-like T-cell lymphoma.⁶⁷ Cytomorphological features include infiltrates that may be monomorphic, composed of smallto-medium sized lymphocytes, or heterogeneous, with atypical lymphocytes intermixed with reactive elements such as histiocytes, plasma cells, and eosinophils. Immunohistochemically, neoplastic cells exhibit a cytotoxic T/NK phenotype, expressing EBER, TIA-1, granzyme B, and perforin (Fig. 5). Most cells are CD8+, though CD4+, CD4/CD8+, or NK phenotypes have been reported as well. The latter may mimic panniculitis-like T-cell lymphoma.¹⁴ CD30 is frequently expressed, and some authors suggest its expression may correlate with a more aggressive clinical course.^{71,82}

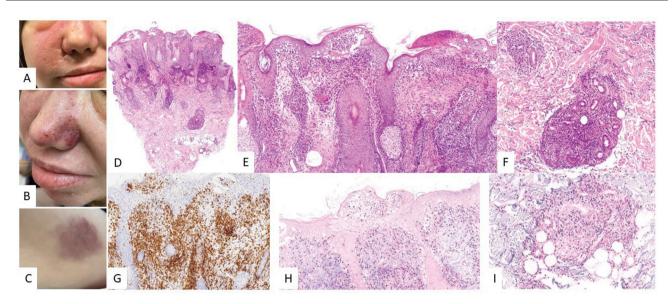


Figure 5 Hydroa vacciniforme-like lymphoproliferative disorder. (A-C) Clinical images of a patient's face showing significant malar edema and lesions in the form of erythematous-violaceous plaques, along with small varioliform scars from previous lesions. (D-F) H&E, $2 \times$ and $10 \times$. At low magnification, a skin punch shows superficial and deep lymphoid infiltration with epidermotropism. At higher magnification, spongiotic vesicles with keratinocyte necrosis are observed (E), and perianexial infiltrates (F). Tumor population is positive for CD2 (G, $10 \times$) and EBER (H and I, $10 \times$).

Expression of PD-L1 has been poorly studied, with a few positive cells among the infiltrates, mainly corresponding to small reactive lymphocytes.⁶⁷

Treatment updates. In mild forms, some patients have responded to antiviral therapy.^{83,84} In advanced cases, immunotherapy or chemotherapy may play a role, although AHSCT remains the only curative treatment.

Severe mosquito bite allergy. The concept and definition of severe mosquito bite allergy have not changed in recent classification updates.

Epidemiology and pathogenesis. This is a very rare NK-cell EBV⁺ LPD. As it happens with HVLPD, the etiology is unknown. At molecular level, no specific changes have been identified in this condition, as no targeted studies have been conducted to this date.

Clinical features. It is characterized by high fever and local skin symptoms following a mosquito bite, presenting as erythema, blisters, ulcers, or necrosis, which leave deep scars.⁸⁵ Patients show elevated serum IgE levels, high EBV DNA titers, and increased NK cells in peripheral blood. Although it is typically self-limiting, there is an increased risk of developing hemophagocytic syndrome and/or progressing to systemic NK/T-cell lymphoma or aggressive NK-cell leukemia.

Histology. Microscopically, skin lesions resemble HVLPD, with more extensive local necrosis and a higher frequency of angiodestruction. The infiltrate is more polymorphic, with lymphocytes of varying sizes—some of them atypical—along with histiocytes and abundant eosinophils. Cells have an NK-cell phenotype, expressing CD3 ε , CD56, TIA-1, granzyme B, and perforin.

Treatment updates. There is no standard treatment. Omalizumab has shown efficacy in preventing anaphylactic episodes in a patient with severe mosquito bite allergy.⁸⁶ *Chronic active EBV disease.* This entity has been renamed in the latest classifications of lymphoid neoplasms, replacing "infection" with "disease" (WHO 5th edition and ICC 2022),^{2,3} considering that only a small proportion of individuals with chronic active EBV infection develop the disease (Appendix).

Epidemiology and pathogenesis. Its incidence is also higher among Asian populations and individuals native to Central and South America. At molecular level, recurrent somatic mutations have been identified in DDX3X, KMT2D, BCOR/BCORL1, KDM6A, and TET2. In the largest study conducted on 83 cases of CAEBV, at least 1 of these mutations was present in 58% of cases. Of particular interest are mutations in DDX3X, also identified in other lymphomas, suggesting that acquiring mutations in this gene may initiate lymphomagenesis. Another group has reported the presence of intragenic deletions in BamHI as well as in other genes needed to produce viral particles. These deletions are hypothesized to be related to the reactivation of the lytic cycle, preventing viral production and cell lysis. Finally, some patients have shown minor defects in cellular immunity that may alter the role of EBV-infected T/NK cells in recognizing exogenous antigens.⁸⁷

Clinical features. This is a condition persisting for 3 or 4 months, during which patients show elevated levels of EBV DNA in peripheral blood and tissue infiltration by EBV-infected T/NK lymphocytes in the absence of immunodeficiency. Approximately half of the patients exhibit symptoms similar to infectious mononucleosis, such as fever, lymphadenopathy, and hepatosplenomegaly. Clinical course is variable but prolonged, and in most cases, disease progresses. Hemophagocytic lymphohistiocytosis is a complication with a poor prognosis.

Histology. Skin involvement is variable, ranging from presentations resembling severe mosquito bite allergy to HV-like signs, making histological studies of the lesions nonspecific. Cases in the form of panniculitis have also been reported.^{88,89}

Treatment updates. Prognosis for patients with CAEBV is poor, and advanced age at presentation seems to be an adverse prognostic factor. Treatment must address both the inflammatory and tumoral processes and should be initiated before disease progresses to lymphoma or hemophagocytic syndrome. The only curative treatment is hematopoietic stem cell transplantation, and the role of chemotherapy is limited to reducing the disease burden prior to transplantation. Given the activation of STAT3 in this disease, the efficacy profile of ruxolitinib has been tested in a clinical trial with promising results.⁹⁰

Extranodal NK/T-cell lymphoma

In the latest classifications of lymphoid neoplasms (WHO 5th edition and ICC 2022), the ''nasal type'' designation has been removed from this entity (Appendix).

Epidemiology and pathogenesis. Unlike pediatric NK/Tcell lymphomas and LPDs, this lymphoma almost exclusively affects adults, with a mean age of 44-54 years and a maleto-female ratio of 2-3:1. Although the exact role of EBV in the pathogenesis of the disease is unknown, EBV positivity is considered essential for diagnosis. On the other hand, multiple genetic alterations have been described in this type of lymphoma, with 6q21-25 deletion being the most common one. This region harbors various tumor suppressor genes, such as PRDM1, PTPRK, FOXO3, and HACE1. Other recurrent alterations include gains in 1g21-g44, 2g, and 7g, and losses in 17p15-22. Additionally, gene expression studies have also highlighted dysregulation in various oncogenic pathways (cell cycle/apoptosis, NF-κB, NOTCH, and JAK/STAT) and alterations in individual genes (MYC, RUNX3, and EZH2). Recently, mutations in epigenetic regulators, such as PRDM1, BCOR, DDX3X, STAT3, and TP53, have been identified.87,91

Clinical features. Involvement of the nasal cavity, nasopharynx, or the upper aerodigestive tract is characteristic, presenting as ulcerative, destructive lesions with bone erosion. Extranasal forms are much less common, with the most frequently involved organs being the skin, the GI tract, testes, and soft tissues. Skin lesions are usually found in the lower extremities as multiple nodules with necrotic ulcerative centers (Fig. 5A). Occasionally, they mimic panniculitis-like lesions. Prognosis is poor, and the disease is typically diagnosed in advanced stages.

Histology. In the skin, dermal involvement can be observed, with infiltration being interstitial or nodular (Fig. 6). Tumor cells are medium to large in size, distributed around blood vessels, infiltrating and destroying vessel walls. This is associated with fibrinoid necrosis, elastic lamina fragmentation, and thrombosis (Fig. 6). These infiltrates may extend into subcutaneous tissue, mimicking panniculitis-like inflammatory processes.

Tumor cells exhibit a T/NK-cell phenotype, with positivity for CD56, CD3 ε , and CD2, as well as variable positivity for FAS, FASL, CD25, CD38, and CD30. Conversely, these cells lack surface CD3, CD4, and CD5 expression. In a small percentage of cases, the tumor population shows a cytotoxic CD8+ T-cell phenotype with monoclonal TCR rearrangements.

Treatment updates. L-asparaginase-based chemotherapy is the gold standard here; however, response is generally poor. Other therapeutic targets in the pipeline include PD-1/PD-L1 inhibitors and drugs modulating the JAK/STAT and NF- κ B pathways.⁸⁷

Primary cutaneous peripheral T-cell lymphoma with a follicular center phenotype

This is a poorly characterized entity with little correspondence to previously described conditions and recently included as a separate entity in the 5th WHO classification (Appendix). According to the largest series, these cases seem to share biological features with nodal Tcell lymphomas of centrofollicular phenotype.⁹² Very few published cases are EBV⁺,⁹³ and they seem to have a worse prognosis and a more aggressive clinical course. Despite the limited case reports, they are mentioned in this review to inform readers and highlight their importance in the differential diagnosis with cutaneous involvement of angioimmunoblastic-type T-follicular helper (TFH) cell lymphoma.

Epidemiology and pathogenesis. A higher prevalence in men has been reported, with a median age at presentation of 67 years. At molecular level, mutations in RHOA and TET2 are the most common alterations. In most cases, clonal rearrangement is detected.

Clinical features. On the skin, multiple nodules or papules appear—often as isolated disease—but with the potential to progress into systemic lymphoma.

Histology. Various patterns have been described, ranging from dense and deep infiltrates to superficial band-like involvement or even a perivascular pattern. Cytologically, infiltrates consist of intermediate to large lymphocytes with irregular nuclei. Phenotypically, tumor cells are T-cells, expressing CD4 and variably showing positivity for Bcl6, CD10, CXCL13,and PD1. EBER-positive cases can be easily distinguished from other entities described in this manuscript based on the phenotype of tumor cells and their clinical presentation. In this case, the main differential diagnosis is with cutaneous involvement by angioimmunoblastic-type T-follicular helper (TFH) cell lymphoma.

Treatment updates. Clinical management varies depending on the case. Most patients receive chemotherapy. Cases with molecular alterations similar to those of angioimmunoblastic-type T-follicular helper (TFH) cell lymphomamay benefit from new targeted therapies, such as histone deacetylase inhibitors and hypomethylating agents.

Intravascular NK/T-cell lymphoma

Since its initial description in 1959 as angioendotheliomatosis proliferans systemisata, less than 30 cases have been published.^{91,94–99} Unlike intravascular B-cell lymphoma, it is not currently considered a distinct entity, and it remains controversial whether it represents a form of aggressive NK leukemia or extranodal NK/T-cell lymphoma.⁹¹

Epidemiology and pathogenesis. Described molecular alterations suggest a multifactorial etiopathogenesis involving genes related to epigenetic regulation⁹¹: histone

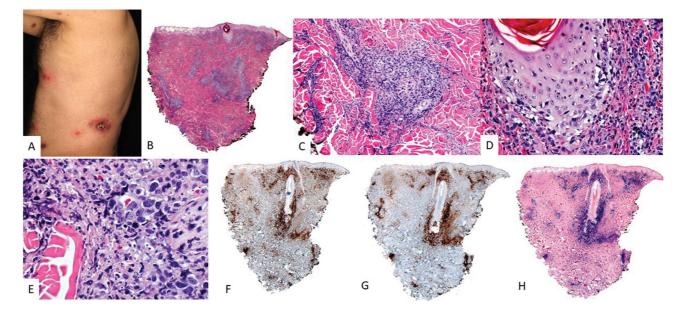


Figure 6 Extranodal NK/T-cell lymphoma. (A) Clinical image shows multiple nodules with necrotic centers and erythematous halos distributed on the trunk. (B) (H&E, $2\times$): At low magnification, a dermal infiltrate is observed in the superficial and deep portions, related to vessels (C, $4\times$) and adnexa with epidermotropism (D, $10\times$). At higher magnification (E, $20\times$), the infiltrate is composed of large atypical elements. These cells are positive for CD3 (F, $2\times$), CD56 (G, $2\times$), and EBER (H, $2\times$).

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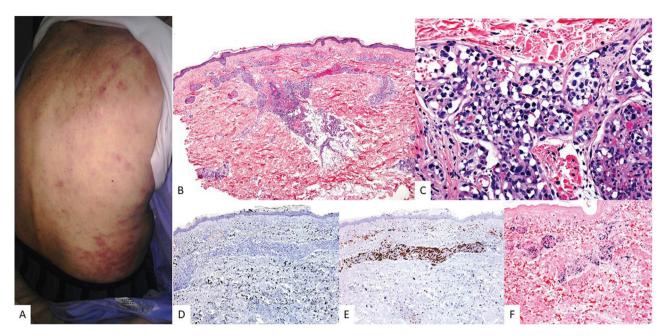


Figure 7 Cutaneous intravascular NK/T-cell lymphoma. (A) Clinical image shows multiple ecchymotic macules and plaques distributed on the trunk, some with a linear or grouped distribution. (B) (H&E, $2 \times$): At low magnification, a dermal infiltrate involving the superficial and deep vessel plexuses is observed. At higher magnification (C, $10 \times$), the infiltrate is located within the vessels and consists of large atypical elements with marked nuclear hyperchromatism. Although these cells are negative for CD20 (D, $4 \times$), they are positive for CD3 (E, $4 \times$) and EBER (F, $4 \times$).

genes (HIST1H2AN, HIST1H2BE, HIST1H2BN, H3F3A) and methylation-related genes (TET2and DNMT1). Additionally, some data suggest involvement in the alternative splicing process (HRAS, MDM2, VEGFA). Finally, some cases have demonstrated strong PD1 expression, which may be related to EBV infection.¹⁰⁰

Clinical features. In addition to the CNS, the skin is one of the most widely affected organs. While it has been suggested that cases with exclusive cutaneous involvement may have a better prognosis than those with multiple organ involvement,⁹⁶ the differences are not statistically significant, and it is still considered an aggressive disease with poor response to chemotherapy. On the skin, it presents non-specifically as erythematous-violaceous patches or plaques on the trunk and extremities (Fig. 7A).

Histology. It is characterized by proliferation—confined to the lumens of vessels—of intermediate to large lymphoid cells of T/NK phenotype, with EBV presence (EBER+) (Fig. 7B-F).

Treatment updates. Currently, there is no standardized chemotherapy regimen, although it seems clear that traditional CHOP regimens are insufficient.

Conflicts of interest

Author Lucía Prieto has participated in training provided by Kiowa and Takeda. The remaining authors declared no conflicts of interest whatsoever.

Acknowledgments

We wish to thank Dr. Socorro María Rodríguez Pinilla for her contribution to the images in Figs. 2, 3, 5 and 6.

Appendix. Supplementary data

Supplementary data associated with this article can be found in the online version available at https://doi.org/ 10.1016/j.ad.2025.02.009.

References

- 1. Willemze R, Cerroni L, Kempf W, Berti E, Facchetti F, Swerdlow SH, et al. The 2018 update of the WHO-EORTC classification for primary cutaneous lymphomas. Blood. 2019;133:1703–14, http://dx.doi.org/10.1182/blood-2018-11-881268.
- Alaggio R, Amador C, Anagnostopoulos I, Attygalle AD, Barreto de Oliveira Araujo I, Berti E, et al. The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours: lymphoid neoplasms. Leukemia. 2022;36:1720–48, http://dx.doi.org/10.1038/s41375-022-01620-2.
- Campo E, Jaffe ES, Cook JR, Quintanilla-Martinez L, Swerdlow SH, Anderson KC, et al. The International Consensus Classification of mature lymphoid neoplasms: a report from the clinical advisory committee. Blood. 2022;140:1229–53, http://dx.doi.org/10.1182/blood.2022015851.
- 4. WHO Classification of Tumours Editorial Board. Haematolymphoid tumors [Internet; beta version ahead of print]. (WHO classification of tumours series, 5th ed.; vol.11). WHO classification of tumours series. 5th ed. Lyon (France): International Agency for Research on Cancer2022. Available from: http://tumourclassification.iarc.who.int/chapters/63 [accessed 2.5.24].

- 5. Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, et al. The 2016 revision of the World Health Organization classification of lymphoid neoplasms. Blood. 2016;127:2375–90, http://dx.doi.org/10.1182/blood-2016-01-643569.
- Prieto-Torres L, Erana I, Gil-Redondo R, Gómez de la Riva I, Manso R, Pajares R, et al. The spectrum of EBVpositive mucocutaneous ulcer: a study of 9 cases. Am J Surg Pathol. 2019;43:201-10, http://dx.doi.org/10.1097/pas. 000000000001186.
- Nagore E, Ledesma E, Collado C, Oliver V, Pérez-Pérez A, Aliaga A. Detection of Epstein Barr virus and human herpesvirus 7 and 8 genomes in primary cutaneous T- and B-cell lymphomas. Br J Dermatol. 2000;143:320–3, http://dx.doi.org/10.1046/ j.1365-2133.2000.03657.x.
- Epstein MA, Achong BG, Barr YM. Virus particles in cultured lymphoblasts from Burkitt's lymphoma. Lancet. 1964;1:702–3, http://dx.doi.org/10.1016/s0140-6736(64)91524-7.
- 9. Dunmire SK, Verghese PS, Balfour HH Jr. Primary Epstein Barr virus infection. J Clin Virol. 2018;102:84–92, http://dx. doi.org/10.1016/j.jcv.2018.03.001.
- Damania B, Münz C. Immunodeficiencies that predispose to pathologies by human oncogenic γ-herpesviruses. FEMS Microbiol Rev. 2019;43:181–92, http://dx.doi.org/ 10.1093/femsre/fuy044.
- Bauer M, Jasinski-Bergner S, Mandelboim O, Wickenhauser C, Seliger B. Epstein Barr virus-associated malignancies and immune escape: the role of the tumor microenvironment and tumor cell evasion strategies. Cancers (Basel). 2021;13, http://dx.doi.org/10.3390/cancers13205189.
- 12. Münz C. Immune escape by non-coding RNAs of the Epstein Barr virus. Front Microbiol. 2021;12:657387, http://dx. doi.org/10.3389/fmicb.2021.657387.
- Dojcinov SD, Venkataraman G, Raffeld M, Pittaluga S, Jaffe ES. EBV positive mucocutaneous ulcer – a study of 26 cases associated with various sources of immunosuppression. Am J Surg Pathol. 2010;34:405–17, http://dx.doi.org/10.1097/ PAS.0b013e3181cf8622.
- 14. Quintanilla-Martinez L, Swerdlow SH, Tousseyn T, Barrionuevo C, Nakamura S, Jaffe ES. New concepts in EBV-associated B, T, and NK cell lymphoproliferative disorders. Virchows Arch. 2023;482:227–44, http://dx.doi.org/10.1007/s00428-022-03414-4.
- Ikeda T, Gion Y, Yoshino T, Sato Y. A review of EBVpositive mucocutaneous ulcers focusing on clinical and pathological aspects. J Clin Exp Hematop. 2019;59:64–71, http://dx.doi.org/10.3960/jslrt.18039.
- Dojcinov SD, Venkataraman G, Pittaluga S, Wlodarska I, Schrager JA, Raffeld M, et al. Age-related EBV-associated lymphoproliferative disorders in the Western population: a spectrum of reactive lymphoid hyperplasia and lymphoma. Blood. 2011;117:4726-35, http://dx.doi.org/10.1182/ blood-2010-12-323238.
- Dojcinov SD, Fend F, Quintanilla-Martinez L. EBV-positive lymphoproliferations of B- T- and NK-cell derivation in non-immunocompromised hosts. Pathogens. 2018;7:28, http://dx.doi.org/10.3390/pathogens7010028.
- Plaza JA, Gru AA, Sangueza OP, Lourenco SV, Puccio FB, Sanches JA, et al. An update on viral-induced cutaneous lymphoproliferative disorders. CME. Part I. J Am Acad Dermatol. 2023;88:965–80, http://dx.doi.org/10.1016/ j.jaad.2021.11.068.
- Gru AA, Jaffe ES. Cutaneous EBV-related lymphoproliferative disorders. Semin Diagn Pathol. 2017;34:60–75, http://dx.doi.org/10.1053/j.semdp.2016.11.003.
- Natkunam Y, Gratzinger D, Chadburn A, Goodlad JR, Chan JK, Said J, et al. Immunodeficiency-associated lymphoproliferative disorders: time for reappraisal? Blood. 2018;132:1871–8.

- Oyama T, Ichimura K, Suzuki R, Suzumiya J, Ohshima K, Yatabe Y, et al. Senile EBV+ B-cell lymphoproliferative disorders: a clinicopathologic study of 22 patients. Am J Surg Pathol. 2003;27:16–26, http://dx.doi.org/10.1097/ 00000478-200301000-00003.
- 22. Bosman F, Jaffe E, Lakhani S, Ohgaki H. World Health Organization classification of tumours. In: Swerdlow SCE, Harris N, editors. WHO classification of tumours of haematopoietic and lymphoid tissues. Lyon, France: IARC; 2008.
- Okamoto A, Yanada M, Miura H, Inaguma Y, Tokuda M, Morishima S, et al. Prognostic significance of Epstein Barr virus DNA detection in pretreatment serum in diffuse large B-cell lymphoma. Cancer Sci. 2015;106:1576–81, http://dx.doi.org/10.1111/cas.12812.
- 24. Jung JM, Na HM, Won CH, Chang SE, Lee MW, Choi JH, et al. Cutaneous Epstein Barr virus-positive diffuse large B-cell lymphoma, not otherwise specified: a systematic review and comparative analysis with Epstein Barr virusnegative, leg type. J Am Acad Dermatol. 2022;86:221–5, http://dx.doi.org/10.1016/j.jaad.2021.01.088.
- 25. Nicolae A, Pittaluga S, Abdullah S, Steinberg SM, Pham TA, Davies-Hill T, et al. EBV-positive large B-cell lymphomas in young patients: a nodal lymphoma with evidence for a tolerogenic immune environment. Blood. 2015;126:863–72, http://dx.doi.org/10.1182/blood-2015-02-630632.
- 26. Vermaat JS, Somers SF, de Wreede LC, Kraan W, de Groen RAL, Schrader AMR, et al. MYD88 mutations identify a molecular subgroup of diffuse large B-cell lymphoma with an unfavorable prognosis. Haematologica. 2020;105:424–34, http://dx.doi.org/10.3324/haematol.2018.214122.
- 27. Gebauer N, Künstner A, Ketzer J, Witte HM, Rausch T, Benes V, et al. Genomic insights into the pathogenesis of Epstein Barr virus-associated diffuse large B-cell lymphoma by whole-genome and targeted amplicon sequencing. Blood Cancer J. 2021;11:102, http://dx.doi.org/10.1038/ s41408-021-00493-5.
- Malpica L, Marques-Piubelli ML, Beltran BE, Chavez JC, Miranda RN, Castillo JJ. EBV-positive diffuse large B-cell lymphoma, not otherwise specified: 2022 update on diagnosis, risk-stratification, and management. Am J Hematol. 2022;97:951–65, http://dx.doi.org/10.1002/ajh.26579.
- Tilly H, Morschhauser F, Sehn LH, Friedberg JW, Trněný M, Sharman JP, et al. Polatuzumab vedotin in previously untreated diffuse large B-cell lymphoma. N Engl J Med. 2022;386:351–63, http://dx.doi.org/10.1056/NEJMoa2115304.
- Liebow AA, Carrington CR, Friedman PJ. Lymphomatoid granulomatosis. Hum Pathol. 1972;3:457–558, http://dx.doi.org/10.1016/s0046-8177(72)80005-4.
- Wilson WH, Kingma DW, Raffeld M, Wittes RE, Jaffe ES. Association of lymphomatoid granulomatosis with Epstein Barr viral infection of B lymphocytes and response to interferon-alpha 2b. Blood. 1996;87:4531–7.
- Beaty MW, Toro J, Sorbara L, Stern JB, Pittaluga S, Raffeld M, et al. Cutaneous lymphomatoid granulomatosis: correlation of clinical and biologic features. Am J Surg Pathol. 2001;25:1111–20, http://dx.doi.org/10.1097/ 00000478-200109000-00001.
- Pollack K, Guffey D, Gru AA. Necrotic plaque on the distal nose with diffuse crateriform nodules. JAMA Dermatol. 2019;155:113-4, http://dx.doi.org/10.1001/jamadermatol. 2018.2552.
- Dunleavy K, Roschewski M, Wilson WH. Lymphomatoid granulomatosis and other Epstein Barr virus associated lymphoproliferative disorders. Curr Hematol Malig Rep. 2012;7:208–15, http://dx.doi.org/10.1007/s11899-012-0132-3.
- **35.** Lipford EH Jr, Margolick JB, Longo DL, Fauci AS, Jaffe ES. Angiocentric immunoproliferative lesions: a clinicopatho-

logic spectrum of post-thymic T-cell proliferations. Blood. 1988;72:1674-81.

- 36. Song JY, Pittaluga S, Dunleavy K, Grant N, White T, Jiang L, et al. Lymphomatoid granulomatosis a single institute experience: pathologic findings and clinical correlations. Am J Surg Pathol. 2015;39:141–56, http://dx.doi.org/10.1097/pas. 000000000000328.
- 37. Shapiro RS, Chauvenet A, McGuire W, Pearson A, Craft AW, McGlave P, et al. Treatment of B-cell lymphoproliferative disorders with interferon alfa and intravenous gamma globulin. N Engl J Med. 1988;318:1334, http://dx.doi.org/10.1056/ nejm198805193182013.
- 38. Siegloch K, Schmitz N, Wu HS, Friedrichs B, van Imhoff GW, Montoto S, et al. Hematopoietic stem cell transplantation in patients with lymphomatoid granulomatosis: a European group for blood and marrow transplantation report. Biol Blood Marrow Transplant. 2013;19:1522–5, http://dx.doi.org/10.1016/j.bbmt.2013.07.023.
- **39.** Delecluse HJ, Anagnostopoulos I, Dallenbach F, Hummel M, Marafioti T, Schneider U, et al. Plasmablastic lymphomas of the oral cavity: a new entity associated with the human immunodeficiency virus infection. Blood. **1997;89**: 1413–20.
- 40. Jaffe ES, Harris NL, Stein H, Vardiman JW. World Health Organisation Classification of Tumours. Pathology and genetics: tumours of the haematopoietic and lymphoid tissues. In: Kc G, Ra W, editors. Diffuse large B-cell lymphoma. Lyon, France: International Agency for Research on Cancer (IARC); 2001. p. 171–6.
- **41.** Jordan LB, Lessells AM, Goodlad JR. Plasmablastic lymphoma arising at a cutaneous site. Histopathology. 2005;46:113–5.
- 42. Zanelli M, Palicelli A, Sanguedolce F, Zizzo M, Filosa A, Ricci L, et al. Cutaneous involvement in diseases with plasma cell differentiation: diagnostic approach. Curr Oncol. 2022;29:3026–43, http://dx.doi.org/10.3390/curroncol 29050246.
- Castillo JJ, Bibas M, Miranda RN. The biology and treatment of plasmablastic lymphoma. Blood. 2015;125:2323-30, http://dx.doi.org/10.1182/blood-2014-10-567479.
- 44. Morscio J, Dierickx D, Nijs J, Verhoef G, Bittoun E, Vanoeteren X, et al. Clinicopathologic comparison of plasmablastic lymphoma in HIV-positive, immunocompetent, and posttransplant patients: single-center series of 25 cases and meta-analysis of 277 reported cases. Am J Surg Pathol. 2014;38:875–86, http://dx.doi.org/10.1097/pas.0000000000234.
- 45. Roberts SJ, McNally B, Rosser JA, Willard N, Golitz L, Wisell J. Diverse clinical and histopathologic features of cutaneous post-transplant lymphoproliferative disorders: a presentation of two cases. Aust J Dermatol. 2019;60:e317–21, http://dx.doi.org/10.1111/ajd.13076.
- **46.** Costello CM, Maly CJ, Snider S, Severson KJ, DiCaudo DJ, Rosenthal AC, et al. Immunosuppression-associated primary cutaneous plasmablastic lymphoma secondary to romidepsin. JAAD Case Rep. 2019;6:19–22.
- 47. Black CL, Foster-Smith E, Lewis ID, Faull RJ, Sidhu SK. Post-transplant plasmablastic lymphoma of the skin. Aust J Dermatol. 2013;54:277–82, http://dx.doi.org/10.1111/j. 1440-0960.2012.00939.x.
- Zanelli M, Sanguedolce F, Zizzo M, Fragliasso V, Broggi G, Palicelli A, et al. Skin involvement by hematological neoplasms with blastic morphology: lymphoblastic lymphoma, blastoid variant of mantle cell lymphoma and differential diagnoses. Cancers (Basel). 2023;15, http://dx.doi.org/10.3390/ cancers15153928.
- Wiemer L, Quan JR, Omman R. An atypical presentation of an uncommon malignancy: plasmablastic lymphoma presenting as recurrent scrotal abscesses. Cureus. 2023;15:e38879, http://dx.doi.org/10.7759/cureus.38879.

- 50. Sato S, Nakahara M, Kato K, Moriyama T, Utsumi S, Sasaki K, et al. Plasmablastic lymphoma occurring in the vicinity of enterocutaneous fistula in Crohn's disease. J Dermatol. 2020;47:e442–3, http://dx.doi.org/10.1111/ 1346-8138.15600.
- **51.** Behera B, Kumari R, Chandrashekar L, Thappa DM, Kar R, Rajesh NG. Primary cutaneous plasmablastic lymphoma presenting as perineal ulcero-proliferative growth in a human immunodeficiency virus-seropositive patient. Indian J Dermatol Venereol Leprol. 2017;83:83–6.
- Varricchio S, Pagliuca F, Travaglino A, Gallo L, Villa MR, Mascolo M. Cutaneous localization of plasmablastic multiple myeloma with heterotopic expression of CD3 and CD4: Skin involvement revealing systemic disease. J Cutan Pathol. 2019;46:619–22, http://dx.doi.org/10.1111/cup.13486.
- 53. Chikeka I, Grossman M, Deng C, Jacob AT, Husain S. Plasmablastic lymphoma in an HIV patient with cutaneous presentation: a case of remarkable remission in a typically refractory disease. JAAD Case Rep. 2020;3:161–5.
- Banks P, Warnke R. WHO Classification of Tumours. Pathology and genetics of haematopoietic and lymphoid tissues. Primary effusion lymphoma. Lyon, France: IARC; 2001. p. 179–80.
- 55. Patel S, Xiao P. Primary effusion lymphoma. Arch Pathol Lab Med. 2013;137:1152–4, http://dx.doi.org/10.5858/ arpa.2012-0294-RS.
- Cesarman E, Chadburn A, Rubinstein PG. KSHV/HHV8mediated hematologic diseases. Blood. 2022;139:1013–25.
- 57. Crane GM, Ambinder RF, Shirley CM, Fishman EK, Kasamon YL, Taube JM, et al. HHV-8-positive and EBV-positive intravascular lymphoma: an unusual presentation of extracavitary primary effusion lymphoma. Am J Surg Pathol. 2014;38:426-32, http://dx.doi.org/10.1097/pas.00000000000128.
- Saggini A, Di Prete M, Facchetti S, Rapisarda VM, Anemona L. Panniculitis-like presentation of extracavitary primary effusion lymphoma. Am J Dermatopathol. 2020;42:446–51, http://dx.doi.org/10.1097/dad.00000000001539.
- 59. Crane GM, Xian RR, Burns KH, Borowitz MJ, Duffield AS, Taube JM. Primary effusion lymphoma presenting as a cutaneous intravascular lymphoma. J Cutan Pathol. 2014;41:928–35, http://dx.doi.org/10.1111/cup.12405.
- **60.** Burns DM, Chaganti S. Epstein Barr virus-associated lymphoproliferative disorders in immunosuppressed patients. Hum Pathol. 2021;38:1293–304.
- 61. Salama S, Todd S, Cina DP, Margetts P. Cutaneous presentation of post-renal transplant lymphoproliferative disorder: a series of four cases. J Cutan Pathol. 2010;37:641–53, http://dx.doi.org/10.1111/j.1600-0560.2009.01449.x.
- 62. Seçkin D, Barete S, Euvrard S, Francès C, Kanitakis J, Geusau A, et al. Primary cutaneous posttransplant lymphoproliferative disorders in solid organ transplant recipients: a multicenter European case series. Am J Transplant. 2013;13:2146–53, http://dx.doi.org/10.1111/ajt.12281.
- 63. Gibson SE, Swerdlow SH, Craig FE, Surti U, Cook JR, Nalesnik MA, et al. EBV-positive extranodal marginal zone lymphoma of mucosa-associated lymphoid tissue in the posttransplant setting: a distinct type of posttransplant lymphoproliferative disorder? Am J Surg Pathol. 2011;35:807–15, http://dx.doi.org/10.1097/PAS.0b013e3182190999.
- 64. Gratzinger D, de Jong D, Jaffe ES, Chadburn A, Chan JK, Goodlad JR, et al. T- and NK-cell lymphomas and systemic lymphoproliferative disorders and the immunodeficiency setting: 2015 SH/EAHP workshop report – Part 4. Am J Clin Pathol. 2017;147:188–203, http://dx.doi.org/10.1093/ajcp/aqw213.
- 65. Miyake T, Iwatsuki K, Hirai Y, Yamamoto T, Hamada T, Fujii K, et al. The aim of the measurement of Epstein Barr virus DNA in hydroa vacciniforme and hypersensitivity to mosquito bites. J Med Virol. 2020;92:3689–96, http://dx.doi.org/ 10.1002/jmv.25811.

- 66. Bazin E. Léçons theoriques et cliniques sur les affectations géneriques de la peau. Delabrage. 1862.
- 67. Garcia-Garcia M, Morales Moya AL, Val IS, Prieto-Torres L. Hydroa vacciniforme lymphoproliferative disorder in a young Spanish woman: an infrequent case with fatal outcome. Am J Dermatopathol. 2024;46:54–9, http://dx.doi.org/10.1097/ dad.00000000002577.
- Xie Y, Wang T, Wang L. Hydroa vacciniforme-like lymphoproliferative disorder: a study of clinicopathology and whole-exome sequencing in Chinese patients. J Dermatol Sci. 2020;99:128–34, http://dx.doi.org/10.1016/j.jdermsci. 2020.06.013.
- 69. Zhang C, Chang L, Yang X, Khan R, Liu D. Severe atypical hydroa vacciniforme-like lymphoproliferative disorder in a patient with hyper IgE syndromes due to DOCK8 gene mutation. Indian J Dermatol Venereol Leprol. 2023;89:874–7, http://dx.doi.org/10.25259/ijdvl_409_2022.
- Cohen JI, Iwatsuki K, Ko YH, Kimura H, Manoli I, Ohshima K, et al. Epstein Barr virus NK and T cell lymphoproliferative disease: report of a 2018 international meeting. Leuk Lymphoma. 2020;61:808–19, http://dx.doi.org/10.1080/10428194. 2019.1699080.
- Magaña M, Sangüeza P, Gil-Beristain J, Sánchez-Sosa S, Salgado A, Ramón G, et al. Angiocentric cutaneous T-cell lymphoma of childhood (hydroa-like lymphoma): a distinctive type of cutaneous T-cell lymphoma. J Am Acad Dermatol. 1998;38:574–9, http://dx.doi.org/10.1016/s0190-9622(98)70120-3.
- Sangueza M, Plaza JA. Hydroa vacciniforme-like cutaneous T-cell lymphoma: clinicopathologic and immunohistochemical study of 12 cases. J Am Acad Dermatol. 2013;69:112–9, http://dx.doi.org/10.1016/j.jaad.2013.01.037.
- Rodríguez-Pinilla SM, Barrionuevo C, Garcia J, Martínez MT, Pajares R, Montes-Moreno S, et al. EBV-associated cutaneous NK/T-cell lymphoma: review of a series of 14 cases from Peru in children and young adults. Am J Surg Pathol. 2010;34:1773–82, http://dx.doi.org/10.1097/PAS.0b013e3181fbb4fd.
- Magaña M, Massone C, Magaña P, Cerroni L. Clinicopathologic features of hydroa vacciniforme-like lymphoma: a series of 9 patients. Am J Dermatopathol. 2016;38:20–5, http://dx.doi.org/10.1097/dad.00000000000385.
- 75. Plaza JA, Sangueza M. Hydroa vacciniforme-like lymphoma with primarily periorbital swelling: 7 cases of an atypical clinical manifestation of this rare cutaneous T-cell lymphoma. Am J Dermatopathol. 2015;37:20–5, http://dx.doi.org/10.1097/ dad.00000000000158.
- 76. Xu W, Tan J, Cai C, Lei L, Cao X, Zhou H, et al. Hydroa vacciniforme-like lymphoproliferative disorder: a retrospective study on clinicopathological characteristics of 32 cases. Pediatr Dermatol. 2022;39:372–5, http://dx.doi.org/ 10.1111/pde.14938.
- 77. Garzón E, Dávila-Rodríguez JJ. Hydroa vacciniforme-like lymphoproliferative disorder in Ecuadorian children: a case series. Indian J Dermatol Venereol Leprol. 2023;89:403–7, http://dx.doi.org/10.25259/ijdvl_847_19.
- Ren F, Zhu J, Perry DM, Pruitt L, Elston DM. Hydroa vacciniforme-like lymphoproliferative disorder: a retrospective cohort study of seven pediatric cases. Int J Dermatol. 2020;59:e290-2, http://dx.doi.org/10.1111/ijd.14931.
- 79. Feng X, Li F, Zhang Y, Wang L. Hydroa vacciniformelike lymphoproliferative disorder with eye involvement. Pediatr Dermatol. 2021;38:1387–8, http://dx.doi.org/ 10.1111/pde.14808.
- Ordoñez-Parra J, Mejía Cortes M, Tamayo-Buendía MM, Infante Gómez AM. Hydroa vacciniforme-like lymphoproliferative disorder (HV-LPD) is an Epstein Barr virus (EBV) associated disease. An Bras Dermatol. 2021;96:388–90, http://dx.doi.org/10.1016/j.abd.2020.06.023.

- 81. López de Cáceres CVB, Rodrigues-Fernandes CI, Rendón Henao J, Morais TML, Soares CD, de Almeida OP, et al. Oral manifestations of Hydroa vacciniforme-like lymphoproliferative disorder: a clinicopathological study of a Peruvian population. J Oral Pathol Med. 2021;50:530–9, http://dx.doi.org/10.1111/jop.13203.
- 82. Kim WY, Nam SJ, Kim S, Kim TM, Heo DS, Kim CW, et al. Prognostic implications of CD30 expression in extranodal natural killer/T-cell lymphoma according to treatment modalities. Leuk Lymphoma. 2015;56:1778–86, http://dx.doi.org/10.3109/10428194.2014.974048.
- İmren IG, Demirkan N, Çomut E, Duygulu Ş. Epstein Barr virus-associated hydroa vacciniforme-like lymphoproliferative syndrome: excellent response to antiviral therapy. Int J Dermatol. 2020;59:e452-4, http://dx.doi.org/10.1111/ijd.15141.
- 84. Lysell J, Wiegleb Edström D, Linde A, Carlsson G, Malmros-Svennilson J, Westermark A, et al. Antiviral therapy in children with hydroa vacciniforme. Acta Derm Venereol. 2009;89:393–7, http://dx.doi.org/10.2340/00015555-0670.
- **85.** Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, et al. WHO Classification of tumours of Haematopoietic and lymphoid tissues. Revised 4th ed. IARC; 2017.
- 86. Meucci E, Radice A, Fassio F, Iorno MLC, Macchia D. Omalizumab for prevention of anaphylactic episodes in a patient with severe mosquito allergy. Clin Case Rep. 2021;9: e04935.
- 87. Syrykh C, Péricart S, Lamaison C, Escudié F, Brousset P, Laurent C. Epstein Barr virus-associated T- and NK-cell lymphoproliferative diseases: a review of clinical and pathological features. Cancers (Basel). 2021;13:3315, http://dx.doi.org/10.3390/cancers13133315.
- Luo H, Yuan Z, Qin B. Case report: chronic active Epstein Barr virus infection with subcutaneous nodules and systemic damage. Front Med (Lausanne). 2022;9:759834, http://dx.doi.org/10.3389/fmed.2022.759834.
- 89. Shibata A, Ishiguro Y, Makita S, Yamaga Y, Kimura H, Akiyama M. A systemic form chronic active Epstein Barr virus infection diagnosed from erythema nodosum-like skin lesions. Eur J Dermatol. 2020;30:314–6.
- Uemura Y, Yamamoto M, Ishimura M, Kanegane H, Sawada A, Hirakawa A, et al. Phase II study of a JAK1/2 inhibitor ruxolitinib for systemic chronic active Epstein Barr virus disease: an investigator-initiated trial. Blood. 2022;140 Suppl. 1:6571–2, http://dx.doi.org/10.1182/blood-2022-168565.
- 91. Zanelli M, Parente P, Sanguedolce F, Zizzo M, Palicelli A, Bisagni A, et al. Intravascular NK/T-cell lymphoma: what we know about this diagnostically challenging, aggressive disease. Cancers (Basel). 2022;14:5458, http://dx.doi.org/10.3390/cancers14215458.
- 92. Wang L, Rocas D, Dalle S, Sako N, Pelletier L, Martin N, et al. Primary cutaneous peripheral T-cell lymphomas with a T-follicular helper phenotype: an integrative clinical, pathological and molecular case series study. Br J Dermatol. 2022;187:970–80, http://dx.doi.org/10.1111/bjd.21791.
- 93. Chiang CT, Chuang SS, Lin HF, Li WH, Chiang YY, Chen BJ. Primary cutaneous peripheral T-cell lymphoma with follicular helper T-cell phenotype: report of 2 Epstein Barr virus-positive cases. Am J Dermatopathol. 2023;45:73–80, http://dx.doi.org/10.1097/dad.0000000002254.
- **94.** Pfleger L, Tappeiner J. On the recognition of systematized endotheliomatosis of the cutaneous blood vessels (reticuloen-dotheliosis? Hautarzt. 1959;10:359–63.
- 95. Santucci M, Pimpinelli N, Massi D, Kadin ME, Meijer CJ, Müller-Hermelink HK, et al. Cytotoxic/natural killer cell cutaneous lymphomas. Report of EORTC Cutaneous Lymphoma Task Force Workshop. Cancer. 2003;97:610–27, http://dx.doi.org/10.1002/cncr.11107.

- Alegría-Landa V, Manzarbeitia F, Salvatierra Calderón MG, Requena L, Rodríguez-Pinilla SM. Cutaneous intravascular natural killer/T cell lymphoma with peculiar immunophenotype. Histopathology. 2017;71:994–1002, http://dx.doi.org/ 10.1111/his.13332.
- Okonkwo L, Jaffe ES. Intravascular large cell lymphoma of NK/T-cell type, EBV positive. Blood. 2017;130:837, http://dx.doi.org/10.1182/blood-2017-05-785857.
- Wang J, Yang X, Song Z, You Y. Cutaneous intravascular NK/T-cell lymphoma. Aust J Dermatol. 2020;61:61–3, http://dx.doi.org/10.1111/ajd.13123.
- 99. Yan J, Zhang F, Luo D, Yao S, Chen Y, Xu F, et al. Intravascular NK/T-cell lymphoma: a series of four cases. Int J Clin Exp Pathol. 2017;10:9541–50.
- 100. Fujikura K, Yamashita D, Sakamoto R, Ishikawa T, Chuang SS, Itoh T, et al. Intravascular NK/T-cell lymphoma: clinicopathological and integrated molecular analysis of two cases provides a clue to disease pathogenesis. J Clin Pathol. 2019;72:642–6, http://dx.doi.org/10.1136/jclinpath-2019-205727.
- 101. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. BMJ. 2021;372:n71, http://dx.doi.org/10.1136/bmj.n71.