

Q1 PRACTICAL DERMOSCOPY

[Translated article] Dermoscopy of Squamous Cell Carcinoma: From Actinic Keratosis to Invasive Forms

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KEYWORDS

Dermoscopy;
Dermatoscopy;
Actinic keratosis;
Bowen's disease;
Squamous cell carcinoma;
Keratoacanthoma

Abstract When performing the dermoscopy of squamous cell carcinoma and its precursors we differentiate among keratin-related, vascular, and pigment-related criteria. Non-pigmented actinic keratoses are characterized by the "strawberry pattern". Pigmented actinic keratosis shows a significant dermatoscopic overlap with lentigo maligna, but the presence of pigmented scales, erythema, and prominent follicles favors its diagnosis. Bowen's disease is characterized by clustered glomerular vessels, white-yellowish scales, and brown or grey dots arranged in lines in its pigmented variant. Finally, dermoscopy allows us to detect invasive squamous cell carcinoma in its early stages and differentiate it from its precursors. Furthermore, its presentation may vary depending on the degree of differentiation, with keratin-associated criteria predominating in well-differentiated tumors, while the atypical vascular pattern will predominate in poorly differentiated tumors.

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PALABRAS CLAVE

Dermatoscopia;
Queratosis actínica;
Enfermedad de Bowen;
Carcinoma epidermoide;
Queratoacantoma

Dermatoscopia del carcinoma epidermoide: de la queratosis actínica a las formas invasivas

Resumen En la evaluación dermatoscópica del carcinoma epidermoide y sus precursores diferenciaremos entre criterios relacionados con la queratina, criterios vasculares y criterios relacionados con el pigmento. Las queratosis actínicas no pigmentadas se caracterizan por el denominado "patrón en fresa". Las queratosis actínicas pigmentadas presentan un gran solapamiento con el léntigo maligno, pero la presencia de escamas pigmentadas, el eritema y los folículos prominentes favorecen su diagnóstico. La enfermedad de Bowen se caracteriza por la presencia de agregados de vasos glomerulares y escamas blanco-amarillentas, así como por

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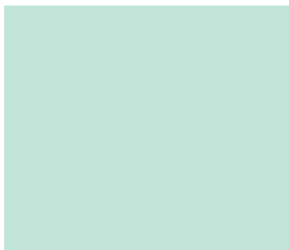
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puntos marrones o grises dispuestos en líneas en su variante pigmentada. Por último, la dermatoscopia puede permitirnos la detección del carcinoma epidermoide invasivo en sus fases incipientes y diferenciarlo de sus precursores. Además, este variará en su presentación en función del grado de diferenciación, predominando los criterios asociados a la queratina en tumores bien diferenciados, mientras que en tumores mal diferenciados predominará un patrón vascular atípico.

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39 Introduction

40 Squamous cell carcinoma (SCC) is the second most fre-
41 quent skin cancer, representing approximately 20% of all
42 non-melanoma skin cancers (NMSC). The incidence of SCC
43 is on the rise.^{1,2} The overall incidence rate in our region has
44 been estimated at 38.16/100,000 persons/year.³ Although
45 most SCCs evolve satisfactorily after surgical excision, there
46 is a subgroup of high-risk lesions with a high probability of
47 recurrence, metastasis, and disease-related death.^{1,3} Con-
48 sidering the progressive aging of the population and the
49 corresponding increase in the incidence of NMSC—especially
50 keratinocyte carcinomas—strategies aimed at the early
51 diagnosis of malignant lesions and their differentiation from
52 precursors are becoming increasingly important.

53 Dermoscopy is a non-invasive technique that is now
54 an essential part of clinical diagnosis in dermatology. In
55 experienced hands, it has been shown to improve diag-
56 nostic accuracy in both pigmented and non-pigmented skin
57 lesions. Specifically, dermoscopy increases sensitivity in the
58 diagnosis of SCC,⁴ with various patterns associated with
59 different types of lesions and stages of progression hav-
60 ing been described, which facilitates differentiating among
61 actinic keratosis (AK), Bowen's disease (BD), and invasive
62 SCC (iSCC)^{5,6} (Fig. 1). Similarly, pigmented AK (pAK) can
63 pose diagnostic challenges due to its clinical overlap with
64 lentigo maligna (LM), a scenario in which dermoscopy has
65 also demonstrated its validity.⁵ This review aims to syn-
66 thesize the existing literature with a practical approach,
67 addressing the spectrum of keratinocytic neoplasms and
68 their precursors from a dermoscopic perspective while uni-
69 fying the considerable terminological heterogeneity existing
70 in Spanish⁷ (Table 1).

71 This review will exclude SCC of the nail apparatus, which,
72 due to its peculiarities and specific dermoscopic criteria,
73 should be addressed in a separate work.

74 Dermoscopy of actinic keratosis

75 Non-pigmented actinic keratosis

76 Key aspects

77 Non-pigmented AK is frequently characterized by the
78 so-called "strawberry pattern," featuring a reddish pseudo-
79 reticulum interrupted by prominent follicles. These can
80 appear as "rosettes" under polarized light dermoscopy
81 (Fig. 2).

82 In an initial trial, Zalaudek et al. studied a total of 41
83 non-pigmented facial AKs and identified a total of 4 funda-
84 mental dermoscopic structures: a reddish pseudo-reticulum
85 (95%), superficial scales (85%), fine linear or wavy perifol-
86 lular vessels (81%), and prominent follicles (66%), and/or
87 surrounded by a white halo ("target follicles") (100%). The
88 combination of these structures forms the metaphorically
89 termed "strawberry pattern".^{8,9} This pattern was later
90 observed by the same group in 67% of lesions in their series,
91 and is significantly associated with the diagnosis of AK vs
92 BD/SCC/keratoacanthoma (KA) ($p < 0.001$).¹⁰

93 Cuellar et al. described structures called "rosettes"
94 (4 bright white dots resembling a "four-leaf clover") in
95 AK, only visible with polarized light¹¹ (Fig. 2). However,
96 these structures have subsequently been described in a wide
97 range of neoplasms and even in non-lesional photo-damaged
98 skin, which is why they are not considered specific.¹²
99 Lozano-Masdemont et al. later proposed that the "rosette
100 pattern," present in 35.8% of lesions in their series charac-
101 terized by this structure as the predominant feature could,
102 indeed, be specific.¹³

103 From a practical standpoint and according to Olsen's clin-
104 ical classification, we will be observing these structures with
105 relative frequency, which will eventually allow us to classify
106 AKs into 5 clinical/dermoscopic stages⁵ (Fig. 2):

107 *Grade 1 AK:* palpable, scarcely visible lesions characte-
108 rized by a reddish pseudo-reticulum and discrete scaling
109 under dermoscopy.

110 *Grade 2 AK:* visible, easily palpable, moderately kera-
111 totic lesions characterized by a reddish background with
112 prominent follicles or "rosettes" under dermoscopy. This
113 stage corresponds to the previously described "strawberry
114 pattern".⁵

115 *Grade 3 AK:* thick lesions with marked hyperkeratosis and
116 well-demarcated borders, predominantly showing compact
117 keratin masses as white/yellowish structureless areas under
118 dermoscopy.^{5,14}

119 Regarding locations, we should mention that non-
120 pigmented extrafacial AKs may present certain dermoscopic
121 differences. In their study, Reinehr et al. saw that whitish
122 scales (97.3%) and erythema (57.4%) were the most common
123 structures. The anatomical differences described in extrafac-
124 ial skin (primarily the lower density of adnexal structures)
125 imply that follicle-associated structures, and the reddish
126 pseudo-reticulum are less common findings.¹⁵

127 The validity of dermoscopy for diagnosing AKs was con-
128 firmed by Huerta-Brogueiras et al. in a prospective trial
129 of 178 clinically suggestive AK lesions, with sensitivity

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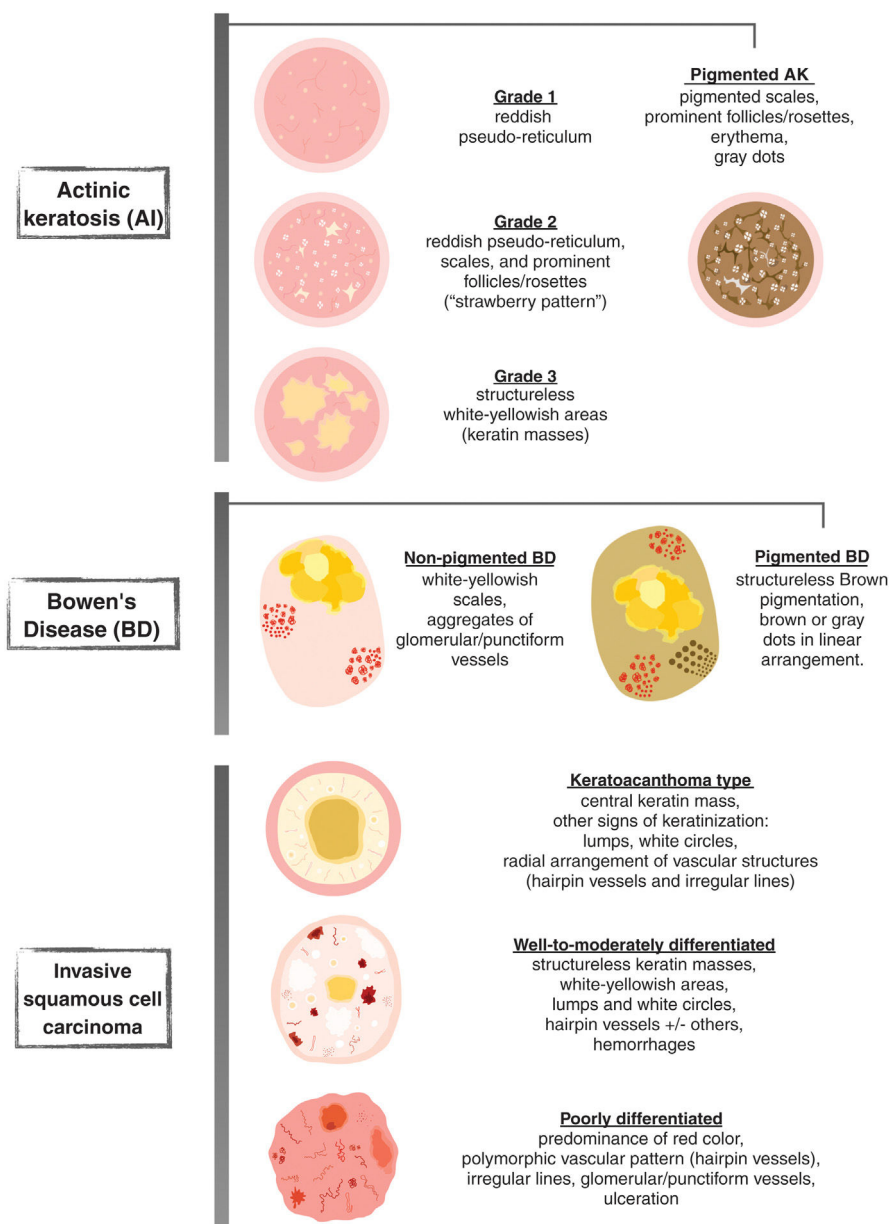


Figure 1 Schematic representation of dermoscopy of actinic keratoses, Bowen's disease, and keratoacanthoma/invasive squamous cell carcinoma.

130 and specificity rates of 98.7% and 95%, respectively, and
 131 a concordance of $\kappa=0.917$ between this technique and
 132 histopathology.¹⁶ Furthermore, dermoscopy also seems to
 133 be useful for post-therapeutic follow-up with cryotherapy,
 134 topical therapies, or photodynamic therapy (PDT).^{17,18}

135 **Pigmented actinic keratosis**

136 **Key aspects**

137 pAK and LM present significant dermoscopic overlapping,
 138 primarily based on pigment-related criteria such as gray
 139 dots, rhomboidal structures, or asymmetric perifollicular
 140 pigmentation.

141 Additional findings, such as prominent folli-
 142 cles/"rosettes," the presence of scales, erythema, or

143 an inner gray halo at follicular level favor the diagnosis of
 144 pAK (Fig. 3).

145 Diagnosing pAK can be challenging due to its clinical and
 146 dermoscopic similarity with LM¹⁹⁻²². In this regard, Akay
 147 et al. studied a total of 99 pigmented facial lesions (67
 148 of them pAKs) and observed that the latter could exhibit
 149 dermoscopic criteria, such as gray dots (70%), an annu-
 150 lar/granular pattern (39%), rhomboidal structures (36%), or
 151 asymmetric perifollicular pigmentation (25%).²³ The study
 152 by Moscarella et al. with 17 facial and extrafacial pAKs
 153 revealed that the most common structures were gray
 154 dots (76.5%), structureless brown areas (58.8%), pigmented
 155 pseudo-reticulum (35.3%), and the presence of white circles
 156 in 11.7% of the cases. Same as LM, these authors observed an
 157 annular/granular pattern and asymmetric perifollicular pig-
 158 mentation in 23.5% and 11.7% of cases, respectively²⁰, while

Table 1 Glossary of the main dermoscopic terms described in the literature in the spectrum of actinic keratosis, Bowen's disease, and invasive squamous cell carcinoma, along with their definition, schematic representation, and histopathological correlation.

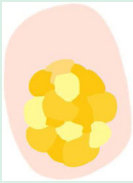
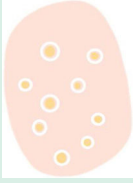
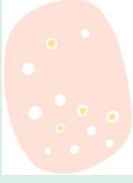
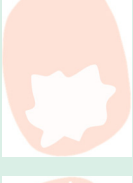
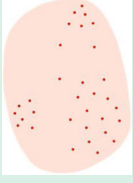
Criterion	Definition	Histopathological correlation	Schematic representation
<i>Keratin-related criteria</i>			
Superficial white-yellow or brown scales	Structureless white-yellow or brown opaque areas	Areas of hyperkeratosis and parakeratosis	
White clods ("keratin pearls")	Rounded white-yellowish structures surrounded by a whitish halo	Keratin pearls or horn swirls	
White circles	Concentric white structures surrounding a follicular orifice, which may have a central yellow globular area	Acanthosis and hypergranulosis of the infundibular epidermis with a central keratin plug	
Structureless white areas	Homogeneous areas that may cover a large part of the tumor, possibly associated with other white structures (circles, clods)	Hyperkeratosis and parakeratosis over a dysplastic epidermis or keratin in neoplastic cell aggregates	
"Rosettes"	4 bright white dots arranged like a "4-leaf clover"	Optical effect of cross-polarization, resulting from alternating hyperkeratosis and parakeratosis in follicular orifices and/or concentric fibrosis	
<i>Vascular criteria</i>			
Reddish pseudoreticulum	Structureless erythematous areas and wavy vessels surrounding follicular orifices	Localized increase in vascularization along with variable follicular hyperkeratosis and keratinocyte atypia	See Fig. 1
"Strawberry pattern"	Reddish pseudo-reticulum interrupted by prominent follicular orifices		See Fig. 1
Dotted vessels	Small caliber red dots resembling pinheads, densely packed together	Tips of short capillary loops in the papillary dermis	

Table 1 (Continued)


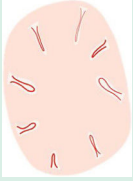
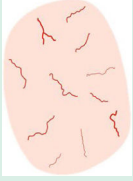
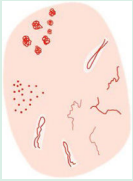
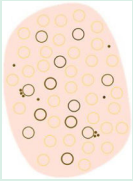
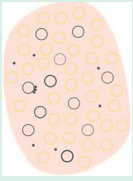

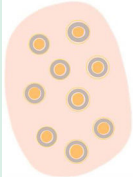
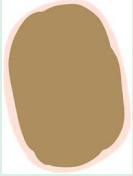
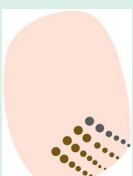
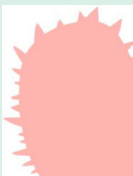

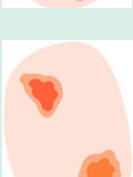
Criterion	Definition	Histopathological correlation	Schematic representation
Glomerular vessels	Larger caliber vessels than the dotted ones, convoluted or "coiled" resembling the renal glomerulus, often distributed in aggregates	Clustered and dilated capillaries in papillary dermis	
"Hairpin" vessels	Loop-shaped vessels in an oblique arrangement to skin surface, usually surrounded by a whitish halo in keratinizing tumors	Capillary loops in the papillary dermis in thick tumors	
Irregular linear vessels	Linearly or slightly curved vessels of irregular shape, size, and/or distribution	Tumoral neoangiogenesis	
Polymorphous vascular pattern	Vessels of various morphologies, often including "hairpin," irregular linear, and dotted/glomerular vessels in invasive squamous cell carcinoma	Tumoral neoangiogenesis	
Pigment-associated criteria			
Brown follicular dots and circles	Small brown rings inside follicular openings	Presence of melanin in basal follicular cells at infundibular level	
Gray follicular dots and circles	Small gray rings inside follicular openings	Presence of melanin at isthmic level along with melanophages in the adjacent dermis	
Rhomboidal structures	Confluent gray dots in linear arrangement of gray-to-brown color between follicular openings	Pigmented keratinocytes in the Malpighian layer and/or melanophages in the superficial dermis	

Table 1 (Continued)

Criterion	Definition	Histopathological correlation	Schematic representation
Inner gray halo	Subtle homogeneous gray or beige halo around follicular openings, forming an inner ring in relation to the pigmented pseudo-reticulum network	Area of preserved epidermis around follicular openings, with gray color as a result of the "Tyndall effect" of pigmented keratinocytes beneath this normal epidermis	
Structureless brown pigmentation	Homogeneous brown areas without other dermoscopic structures	Diffusely distributed melanin in basal keratinocytes	
Patchy or linearly arranged brown/gray dots/globules	Brown or gray dots arranged in a patchy or linear way, often found at the lesion periphery with radial orientation	Melanophages in dermal papillae near papillary vessels along with thin suprapapillary epidermis and/or increased number of pigmented keratinocytes	
Other criteria			
"Red starburst"	Presence of radial red lines or "hairpin" vessels around the central structureless white-yellow area of the lesion	May represent a sign of "horizontal" lesion growth	
Erosions	Small irregularly arranged structureless orange-to-red or brown areas	Loss of epidermis	
Ulceration	Large irregular or rounded structureless red or reddish-brown areas	Loss of epidermis and superficial dermis	

Source: Kittler et al.⁹, Zalaudek et al.¹⁰, Akay et al.²³, Lallas et al.²⁴, Ertop Doğan et al.²⁵, Nascimento et al.²⁶, Martín et al.²⁷, Zalaudek et al.²⁸, Zalaudek et al.³⁰, Cameron et al.³⁷, Rosendahl et al.⁴¹, Yélamos et al.⁵², and Kreusch⁵³.

159 Kelati et al. found an annular/granular pattern and rhom-
160 boidal structures in 19.4% and 82.8% of 232 pAKs.²² However,
161 certain dermoscopic criteria can aid in this difficult differ-
162 ential diagnosis. Lallas et al. studied a total of 70 LMs and 56
163 pAKs to find that the presence of white circles/prominent
164 follicles (OR, 13.5; p=0.006), scales (OR, 7.7; p=0.001),
165 and erythema (OR, 3.6; p=0.009) correlated with the diag-
166 nosis of pAK. Conversely, rhomboidal structures, intense
167 pigmentation, and non-prominent follicles were predictors
168 of LM.²⁴ A recent study including 53 pAKs confirmed that
169 erythema (35.8%), scales (77.4%), and prominent follicles
170 (52.8%) could be key in identifying these lesions while also

171 seeing structures shared with LM such as brown dots (22.6%)
172 and circles (43.4%), gray dots (45.4%) and circles (26.4%),
173 and structureless pigmented areas (30.2%).²⁵ Finally, the
174 presence of an "inner gray halo" at follicular level was
175 reported by Nascimento et al. as a predictor of pAK vs LM,
176 reporting this dermoscopic sign in 91.4% of pAKs vs 23.8%
177 of LMs (p<0.01), with excellent inter-observer agreement
178 ($\kappa = 0.846$).²⁶

179 **Other variants**

180 Bowenoid AKs exhibit glomerular vessels of regular
181 distribution^{27,28} (Fig. 2), while lichenoid AKs can exhibit

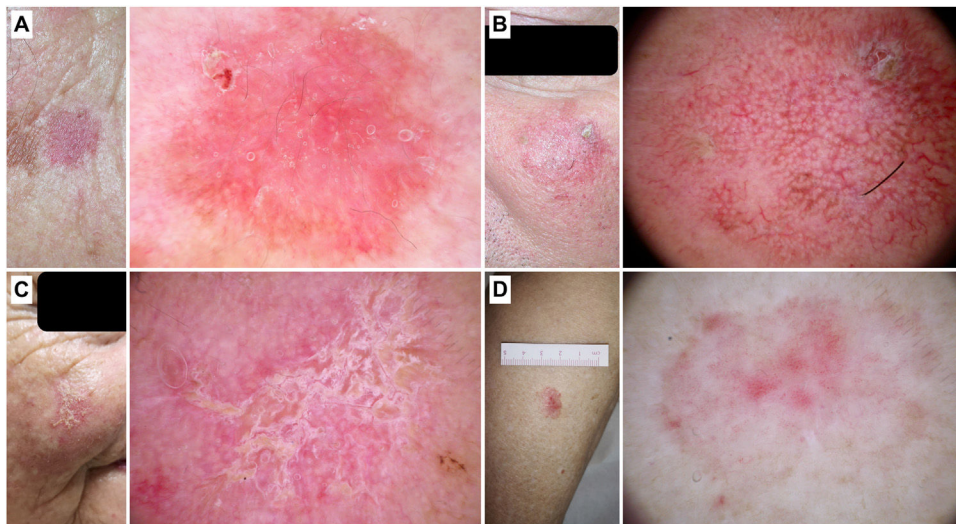


Figure 2 Non-pigmented actinic keratoses. A) Grade 1 actinic keratosis on the face of an 81-year-old woman. Dermoscopy shows a reddish pseudo-reticulum and discrete scaling. B) Grade 2 facial actinic keratosis in a 72-year-old man. Dermoscopic “strawberry pattern,” with prominent follicles/“rosettes,” perifollicular wavy vessels, and keratin. C) Grade 3 actinic keratosis on the face of an 80-year-old woman. Dermoscopy shows keratin masses as structureless white-yellowish areas. D) Bowenoid actinic keratosis on the leg of an 80-year-old woman. Dermoscopy shows multiple dotted/glomerular vessels.

gray annular/granular patterns due to melanophagia.⁵ Recently, the “iceberg sign” was described in AKs with a bluish surface coloration. The presence of this sign has been associated with the use of certain violet shampoos and the following deposition of amorphous basophilic material in the stratum corneum.²⁹

Dermoscopy of in situ squamous cell carcinoma (Bowen’s disease)

Non-pigmented Bowen’s disease

Key aspects

Non-pigmented BD is characterized by the presence of aggregates of glomerular or dotted vessels along with white/yellowish scales (Fig. 4).

In the initial study by Zalaudek et al. that analyzed 21 cases of BD found that the most common dermoscopic structures were glomerular vessels (90%) and white/yellowish scales (90%).³⁰ These data were corroborated by the same group, observing these criteria in 43.3% and 83.1% of all 71 cases of BD, with significant differences vs AK. The authors also described the presence of a “red starburst” pattern in 28.2% of BD cases. Based on the AK-BD-SCC progression model, they hypothesized that this criterion could represent an initial step in such progression.¹⁰ In this regard, Pan et al. conducted a retrospective observational study with 50 cases of BD, 150 basal cell carcinomas (BCC), and 100 cases of psoriasis, concluding that the combination of aggregated glomerular vessels and hyperkeratosis achieved a diagnostic likelihood of 98% for BD.³¹ Of note that vessel morphology may be influenced by the magnification used. Thus, with standard handheld dermatoscopes, we can see aggregates of dotted vessels.³² In a recent study, Papageorgiou et al. observed that dotted and glomerular vessels were the main

predictors of BD vs BCC. However, they also noted that dotted or glomerular vessels can be detected in BCCs located on the lower limbs, likely due to venous stasis (25% and 19.3% in the study, respectively).³³ Aside from these fundamental findings, other described dermoscopic criteria include hemorrhages, focal hypopigmentation, irregular linear vessels, or “hairpin” vessels, among others.³⁴⁻³⁶ Additionally, dermoscopy seems useful at post-therapeutic follow-up with imiquimod. In a small patient series, Mun et al. saw reported that persistent glomerular vessels after treatment would be suggestive of the presence of residual tumor.³⁴

Pigmented Bowen’s disease

Key aspects

Pigmented BD is characterized by the criteria present in the non-pigmented form and the presence of structureless brown pigmentation areas and brown or gray dots arranged in lines (Fig. 4).

In an initial study by Zalaudek’s group that analyzed 10 pigmented BD cases and reported brown globules with a patchy distribution (90%), as well as structureless gray or brown areas (80%).³⁰ Afterwards, Cameron et al. published a retrospective study with 52 pigmented BD cases reporting the presence of brown or gray dots with a linear distribution pattern in 21.2% of cases. In 48.1% of cases, however, a structureless pigmentation pattern predominated, while 34.6% exhibited a combination of structureless pigmentation and dots. Most cases exhibited a monomorphic vascular pattern (82.9%), with predominance of glomerular vessels (44.2%), with a linear vessel distribution in 11.5%.³⁷ Other studies have corroborated these findings in varying percentages, mainly the presence of glomerular vessels (50% up to 100%), pigmented dots/globules (30% up to 80%), or structureless pigmentation areas (70% up to 78%).^{34,36,38}

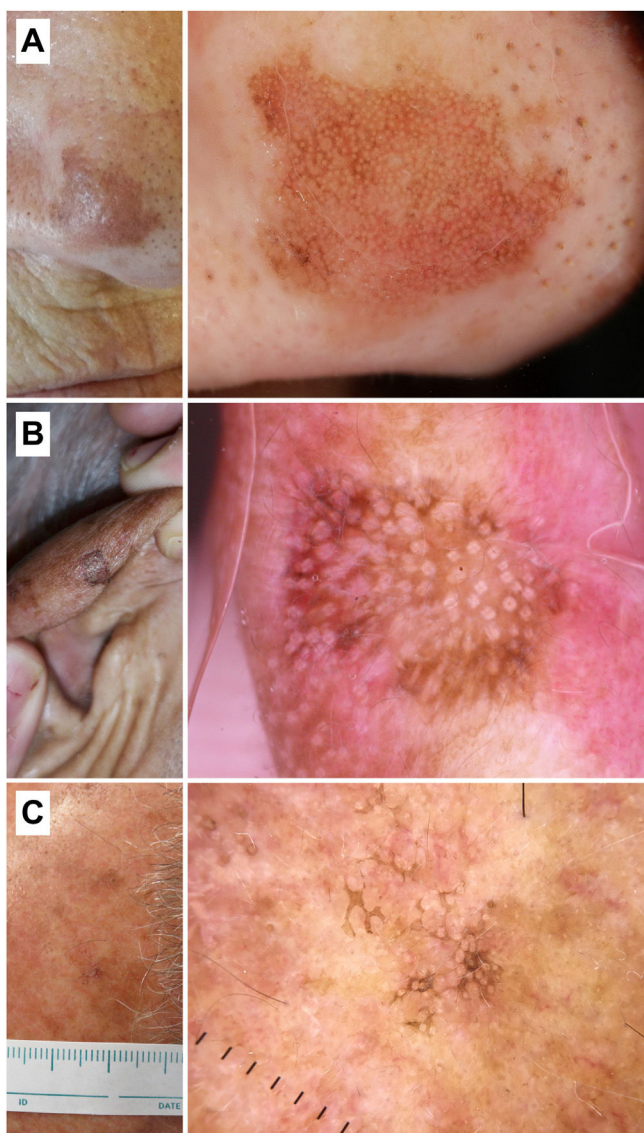


Figure 3 Pigmented actinic keratoses. A) Pigmented actinic keratosis on the nasal pyramid of a 70-year-old woman. Dermoscopy shows an annular/granular pattern, erythema, and prominent follicles. B) Pigmented actinic keratosis on the helix of an 85-year-old man. Dermoscopy shows a structureless brown area, erythema, and multiple "rosettes." C) Pigmented actinic keratosis on the temple of a 56-year-old man. Dermoscopy shows pigmented scales and prominent follicles/"rosettes."

While these findings have proven reproducible in the genital area³⁸ (Fig. 4), a study that analyzed a total of 79 head and neck lesions concluded that dermoscopic patterns in this location differ from previously published data and are similar to pAKs based on a lower presence of glomerular vessels (7.6%) and dots with linear arrangement (13.9%), the observation of structures, such as pigmented circles (48.1%) and white circles (17.7%), rhomboidal structures (41.8%), and structureless pigmentation areas (86.1%), as well as the predominance of irregular linear vessels (29.2%)³⁹ (Fig. 5). Other less consistently described dermoscopic criteria include projections, pigmented reticulum, hypopigmented areas, or ulceration^{30,34,36,39}.

Dermoscopy of invasive squamous cell carcinoma

Key aspects

In iSCC, we can identify dermoscopic criteria associated with keratinization (white clods and circles, structureless white areas), vascular criteria (irregular linear vessels, dotted/glomerular vessels, and "hairpin" vessels), and other criteria (ulceration, hemorrhages).

In well-to-moderately differentiated SCC, keratinization-associated criteria are predominant, while in poorly differentiated SCC, a polymorphous vascular pattern predominates (Fig. 6).

In the differential diagnosis with AK, the presence of dotted/glomerular vessels, "red starburst," "hairpin" vessels, structureless white areas, and perifollicular white circles should prompt a skin biopsy to rule out iSCC.

The study by Zalaudek et al. of 78 iSCCs and 24 KAs reported that "hairpin" vessels (38.5%), irregular linear vessels (17.9%), target follicles (41%), structureless white areas (42.3%), central keratin (39.4%), and ulceration (17.9%) were significantly associated with the diagnosis of iSCC ($p < 0.001$), with similar frequencies in the KA group except for irregular linear vessels, which were more common in the latter.¹⁰ On the other hand, Jaimes et al. coined the term "keratin pearls" after studying a total of 15 well-differentiated KAs/SCCs and observed that all lesions exhibited rounded white/yellowish structures surrounded by a whitish halo⁴⁰ (Fig. 7). These structures, also known as white clods, were present in 25.6% and 16.7% of KA and SCC cases in the study by Rosendahl et al., respectively. This study, designed as a retrospective and prospective study with 43 KAs/60 iSCCs and 29 KAs/32 iSCCs/145 other lesions, respectively, concluded: 1) central keratin was more common in KA (51.2%) vs iSCC (30.0%) ($p = 0.03$); 2) the presence of keratin was more common in the KA/iSCC group vs other lesions (78.7% vs 30.3%; $p < 0.001$), with a sensitivity rate and positive predictive value (PPV) of 79% and 92% vs the BCC group, respectively; 3) structureless white areas (39.3% vs 18.6%; $p = 0.02$) and white circles (44.3% vs 13.1%; $p < 0.001$) were more common in the KA/iSCC group, with an 87% specificity rate for the latter vs other lesions; and 4) in the multivariate model, keratin, hemorrhages, and white circles were the only independent predictors of KA/iSCC diagnosis, with the latter reaching the highest ORs at 6.1 (95%CI, 2.4-13.3; $p < 0.001$).⁴¹ In this context, of note that iSCC and KA can exhibit overlapping dermoscopic patterns, meaning that histopathological examinations will be necessary in most cases.⁴²

Based on the degree of tumor differentiation, several dermoscopic patterns can be observed.^{43,44} In conclusion, well-to-moderately differentiated iSCCs will more frequently exhibit "hairpin" vessels, structureless white or yellowish areas, white clods, and circles. Specifically, KA has been associated with the presence of a central keratin mass surrounded by "hairpin" or irregular linear vessels of a radial distribution.^{10,28} Conversely, poorly differentiated tumors will predominantly exhibit a polymorphous/atypical vascular pattern and ulceration.⁵ Lallas et al. found that the predominance of red color was associated with a 13-fold greater likelihood of being a poorly differentiated iSCC.⁴⁴ A similar pattern with predominant atypical vascularization

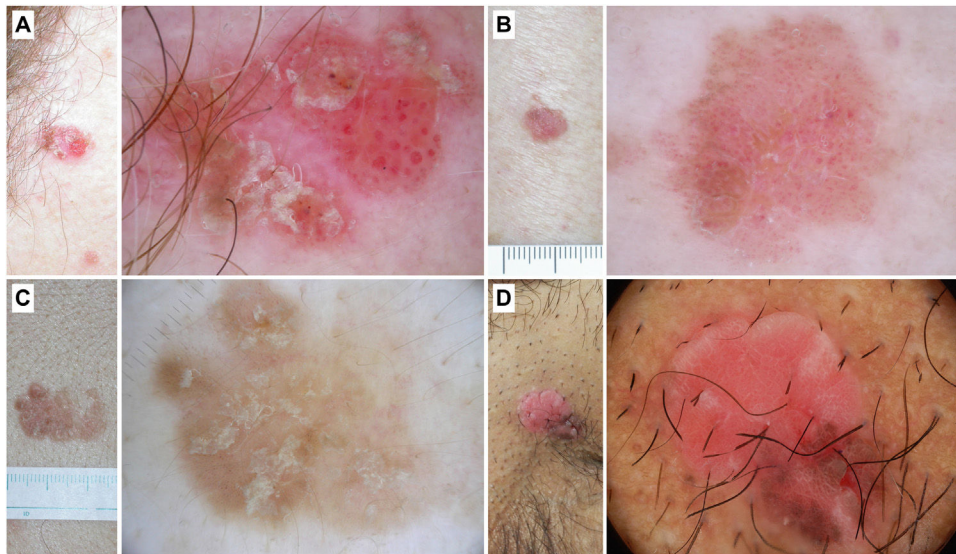


Figure 4 Bowen's disease. A) Non-pigmented Bowen's disease on the temple of a 79-year-old woman. Dermoscopic pattern consists of white-yellowish scales and aggregates of glomerular vessels. B) Non-pigmented Bowen's disease on the back of a 55-year-old man. Dermoscopy shows discrete scaling and glomerular and dotted vessels. C) Pigmented Bowen's disease on the back of a 31-year-old man. Dermoscopy shows structureless brown pigmentation, aggregates of glomerular vessels, and a linear arrangement of brown and gray dots. D) Genital Bowen's disease on the pubis of a 59-year-old man. Dermoscopy shows a structureless brown pigmentation area, brown dots in linear arrangement, dotted vessels, and bright white lines.

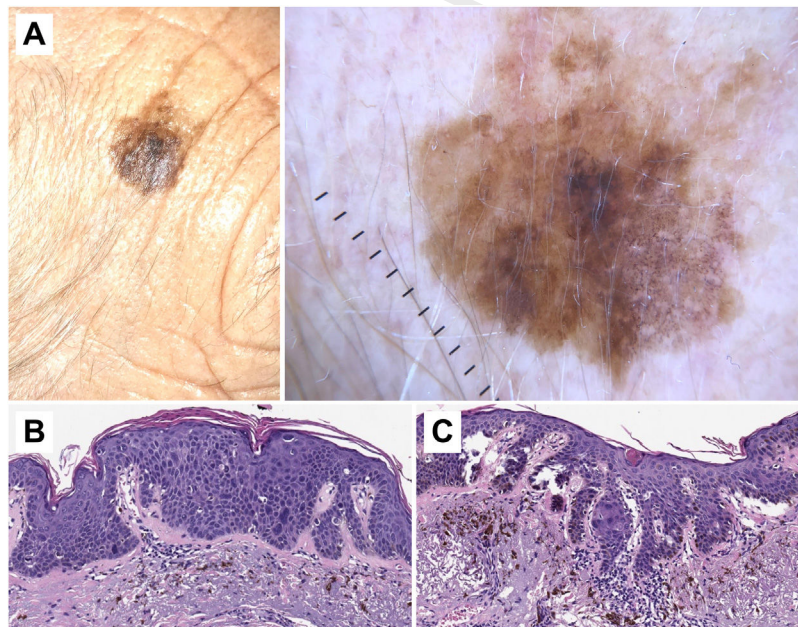


Figure 5 Histopathological correlation in pigmented actinic keratosis with areas of in situ squamous cell carcinoma. A) Pigmented atypical lesion on the temple of a 75-year-old woman. Dermoscopic image shows homogeneous brown pigmentation regions, brown circles, blue-gray granules, and gray dots in linear arrangement in the lower pole. In this case, dermoscopic findings are not suitable to establish a reliable diagnosis, making histopathological examination of paramount importance. B) Histopathological image of an area of in situ squamous cell carcinoma (Bowen's disease) with melanophages on the superficial dermis (hematoxylin and eosin, $\times 10$). C) Area of proliferative actinic keratosis with pigmented keratinocytes and melanophages on the superficial dermis (hematoxylin and eosin, $\times 10$).

Q2

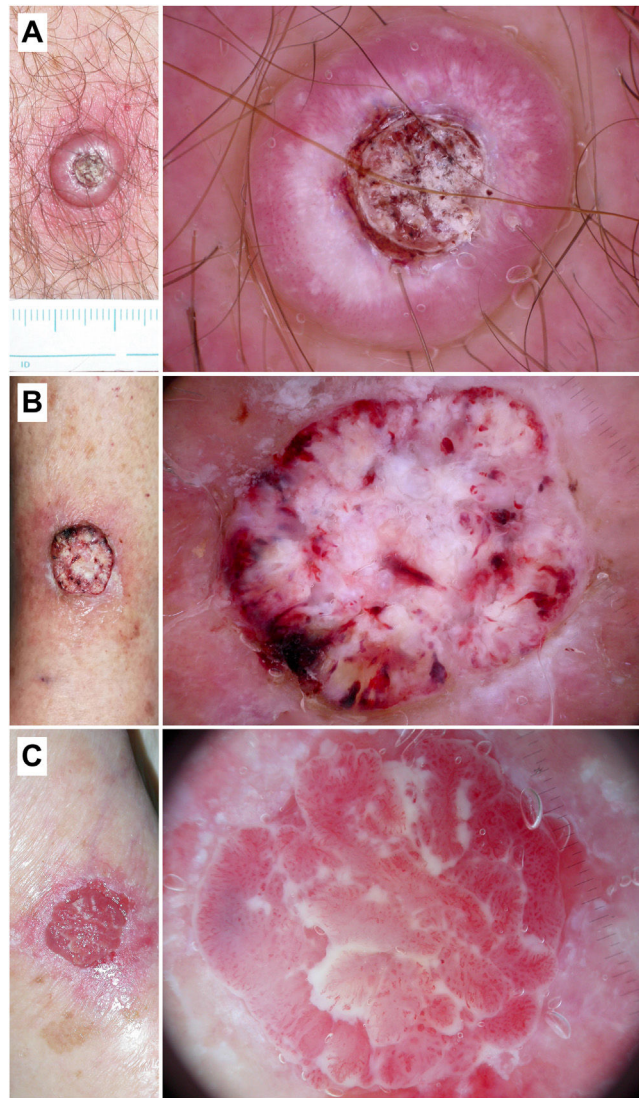


Figure 6 Keratoacanthoma and invasive squamous cell carcinoma. A) Keratoacanthoma on the chest of a 54-year-old man. Dermoscopy shows a central keratin mass, white clods and circles, structureless white areas, and "hairpin" vessels with a radial distribution. B) Well-differentiated squamous cell carcinoma on the leg of an 88-year-old woman. Dermoscopy shows structureless white-yellowish areas, white clods and circles, a polymorphous vascular pattern, and hemorrhages. C) Poorly differentiated squamous cell carcinoma on the arm of an 89-year-old woman. Dermoscopy shows a predominance of red color and an overtly atypical vascular pattern, with few keratinization-associated criteria.

321 has been reported in combined SCC/Merkel cell carcinoma
322 tumors⁴⁵.

323 Regarding locations, we should mention that lip SCC
324 shares most dermoscopic characteristics with cutaneous
325 SCC.⁴⁶ Benati et al. published a series of cases of 22 lip SCC,
326 in which the most relevant structures were scales (100%),
327 perivascular white halos (86%), structureless white areas
328 (91%), white circles (59%), and a polymorphous vascular pat-
329 tern (68%).⁴⁷ In a recent multicentric retrospective study of
330 177 lip lesions (107 of them SCC), Lallas et al. saw that the
331 presence of white clods and ulceration were predictors of
332 SCC diagnosis vs controls (OR, 6.38 and 4.11, respectively).⁴⁸

333 On the differential diagnosis with other lesions, early
334 detection of iSCC and its differentiation from AK is essen-

335 tial, a common scenario in the follow-up of patients with
336 actinic damage. The "red starburst" outburst described by
337 the study of Zalaudek et al. was observed in 29.5% of iSCC
338 cases, with no significant differences being reported in fre-
339 quency vs BD/in situ SCC. In any case, this dermoscopic
340 pattern should be considered when planning to perform a
341 skin biopsy in the context of a patient with actinic damage.¹⁰
342 Papageorgiou et al. collected 50 incipient cases of iSCC
343 and 45 AK with histopathological confirmation and found
344 that the presence of dotted/glomerular vessels (OR, 3.83),
345 "hairpin" vessels (OR, 12.12), and structureless white areas
346 (OR, 3.58) were the main predictors for SCC diagnosis on the
347 multivariate analysis. The univariate model also suggested
348 that since ulceration, perivascular white halos, and white

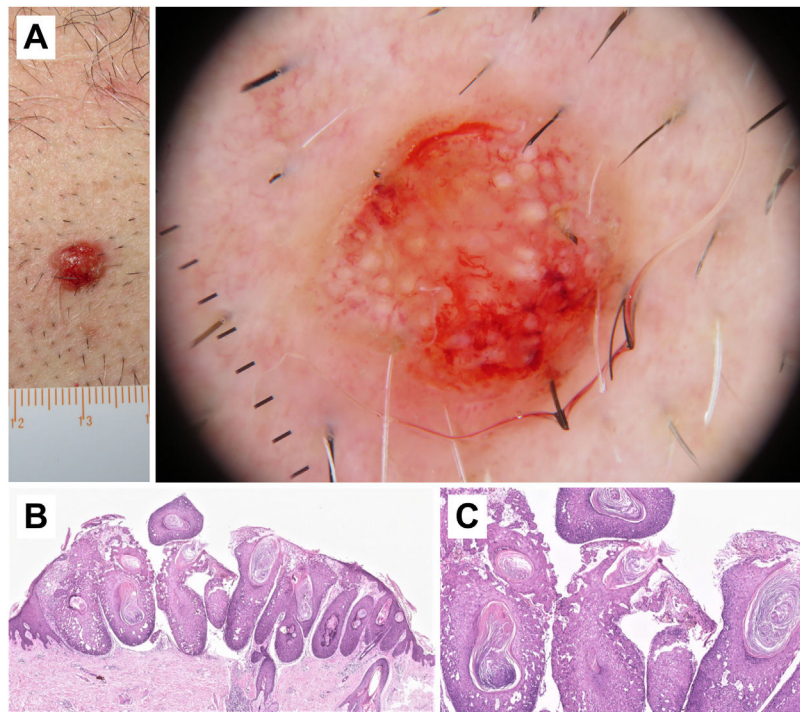


Figure 7 Histopathological correlation in microinvasive squamous cell carcinoma. A) Erythematous papule on the chest of a 46-year-old man. B) Dermoscopic image shows white clods and a polymorphous vascular pattern with irregular linear, “hairpin,” and glomerular vessels. C) Histopathological image shows epithelial proliferation with acanthosis and bulbous nests, revealing acantholysis and central keratinization phenomena (hematoxylin and eosin, $\times 2$). D) Higher magnification image showing tumor cells and keratinization areas (“horn pearls”), corresponding to the white clods seen in dermoscopy (hematoxylin and eosin, $\times 5$).

circles could be predictors of SCC, they should also be taken into consideration.⁴⁹ iSCC can also significantly overlap with common benign lesions, such as irritated seborrheic keratosis (ISK), particularly in well-differentiated SCC cases.⁵⁰ A study conducted by the same group analyzed 104 cases of SCC and 61 ISK and observed that the presence of dotted vessels (OR, 10.4), branched linear vessels (OR, 5.3), structureless white areas (OR, 6.78), white circles (OR, 23.45), or the irregular (OR, 2.55) or peripheral (OR, 2.8) distribution of vascular structures were predictors of SCC diagnosis vs ISK.⁵¹

Conclusions

Patients with actinic damage/cancerization field generally present with dozens of skin lesions of varying biological behavior. In this context, an accurate differential diagnosis that allows us to select malignant lesions amenable to surgery and reliably identify “pre-malignant” lesions amenable to other treatments is desirable. Based on the available evidence, dermoscopy can be key in this endeavor, and therefore, appropriate training beyond melanocytic neoplasms, integrating the spectrum of keratinocytic carcinomas, is essential.

Conflicts of interest

None declared.

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References

1. Que SKT, Zwald FO, Schmults CD. Cutaneous squamous cell carcinoma: Incidence, risk factors, diagnosis, and staging. *J Am Acad Dermatol.* 2018;78:237–47, <http://dx.doi.org/10.1016/J.JAAD.2017.08.059>.
2. Stratigos AJ, Garbe C, Dessinioti C, Lebbe C, Bataille V, Bastholt L, et al. European interdisciplinary guideline on invasive squamous cell carcinoma of the skin: Part 1. Epidemiology, diagnostics and prevention. *Eur J Cancer.* 2020;128:60–82, <http://dx.doi.org/10.1016/J.EJCA.2020.01.007>.
3. Tejera-Vaquero A, Descalzo-Gallego MA, Otero-Rivas MM, Posada-García C, Rodríguez-Pazos L, Pastushenko I, et al. Skin Cancer Incidence and Mortality in Spain: A Systematic Review and Meta-Analysis. *Actas Dermosifiliogr.* 2016;107:318–28, <http://dx.doi.org/10.1016/J.AD.2015.12.008>.
4. Lin MJ, Xie C, Pan Y, Jalilian C, Kelly JW. Dermoscopy improves diagnostic accuracy for clinically amelanotic nodules. *Australas J Dermatol.* 2019;60:45–9, <http://dx.doi.org/10.1111/AJD.12902>.
5. Zalaudek I, Argenziano G. Dermoscopy of actinic keratosis, intraepidermal carcinoma and squamous cell carcinoma. *Curr Probl Dermatol.* 2015;46:70–6, <http://dx.doi.org/10.1159/000366539>.

6. Zaballos Diego P. Dermoscopy Can Indicate the Grade of Actinic Keratoses. *Actas Dermosifiliogr.* 2017;108:798, <http://dx.doi.org/10.1016/J.AD.2017.07.005>.
7. Valdés-Morales KL, Peralta-Pedrero ML, Jurado-Santa Cruz F, Morales-Sánchez MA. Diagnostic Accuracy of Dermoscopy of Actinic Keratosis: A Systematic Review. *Dermatol Pract Concept.* 2020;10, <http://dx.doi.org/10.5826/DPC.1004A121>.
8. Zalaudek I, Giacomel J, Argenziano G, Hofmann-Wellenhof R, Micantonio T, et al. Dermoscopy of facial nonpigmented actinic keratosis. *Br J Dermatol.* 2006;155:951-6, <http://dx.doi.org/10.1111/J.1365-2133.2006.07426.X>.
9. Kittler H, Marghoob AA, Argenziano G, Carrera C, Curiel-Lewandrowski C, Hofmann-Wellenhof R, et al. Standardization of terminology in dermoscopy/dermatoscopy: Results of the third consensus conference of the International Society of Dermoscopy. *J Am Acad Dermatol.* 2016;74:1093, <http://dx.doi.org/10.1016/J.JAAD.2015.12.038>.
10. Zalaudek I, Giacomel J, Schmid K, Bondino S, Rosendahl C, Cavicchini S, et al. Dermatoscopy of facial actinic keratosis, intraepidermal carcinoma, and invasive squamous cell carcinoma: A progression model. *J Am Acad Dermatol.* 2012;66:589-97, <http://dx.doi.org/10.1016/J.JAAD.2011.02.011>.
11. Cuellar F, Vilalta A, Puig S, Palou J, Salerni G, Malveyh J. New dermoscopic pattern in actinic keratosis and related conditions. *Arch Dermatol.* 2009;145:732, <http://dx.doi.org/10.1001/ARCHDERMATOL.2009.86>.
12. Liebman TN, Scope A, Rabinovitz H, Braun RP, Marghoob AA. Rosettes may be observed in a range of conditions. *Arch Dermatol.* 2011;147:1468, <http://dx.doi.org/10.1001/ARCHDERMATOL.2011.312>.
13. Lozano-Masdemont B, Polimón-Olabarrieta I, Marinero-Escobedo S, Gutiérrez-Pecharrmán A, Rodríguez-Lomba E. Rosettes in actinic keratosis and squamous cell carcinoma: Distribution, association to other dermoscopic signs and description of the rosette pattern. *J Eur Acad Dermatol Venereol.* 2018;32:48-52, <http://dx.doi.org/10.1111/JDV.14474>.
14. Çayirli M, Köse O, Demiriz M. Dermatoscopy of facial nonpigmented actinic keratosis regarding to grades of the lesions. *J Eur Acad Dermatol Venereol.* 2013;27:1185-7, <http://dx.doi.org/10.1111/JDV.12020>.
15. Reinehr CPH, Garbin GC, Bakos RM. Dermatoscopic Patterns of Nonfacial Actinic Keratosis: Characterization of Pigmented and Nonpigmented Lesions. *Dermatol Surg.* 2017;43:1385-91, <http://dx.doi.org/10.1097/DSS.0000000000001210>.
16. Huerta-Brogeras M, Olmos O, Borbujo J, Hernández-Núñez A, Castaño E, Romero-Maté A, et al. Validation of dermoscopy as a real-time noninvasive diagnostic imaging technique for actinic keratosis. *Arch Dermatol.* 2012;148:1159-64, <http://dx.doi.org/10.1001/ARCHDERMATOL.2012.1060>.
17. Pampena R, Condorelli A, Cornacchia L, Guiducci L, Raucci M, Benati E, et al. Treatment monitoring of 5-fluorouracil 0.5%/salicylic acid 10% lesion-directed therapy for actinic keratosis using dermoscopy and in-vivo reflectance confocal microscopy. *Dermatol Ther.* 2020;33, <http://dx.doi.org/10.1111/DTH.13744>.
18. Lee JH, Won CY, Kim GM, Kim SY. Dermoscopic features of actinic keratosis and follow up with dermoscopy: A pilot study. *J Dermatol.* 2014;41:487-93, <http://dx.doi.org/10.1111/1346-8138.12282>.
19. Zalaudek I, Ferrara G, Leinweber B, Mercogliano A, D'Ambrosio A, Argenziano G. Pitfalls in the clinical and dermoscopic diagnosis of pigmented actinic keratosis. *J Am Acad Dermatol.* 2005;53:1071-4, <http://dx.doi.org/10.1016/J.JAAD.2005.08.052>.
20. Moscarella E, Rabinovitz H, Zalaudek I, Piana S, Stanganelli I, Oliviero MC, et al. Dermoscopy and reflectance confocal microscopy of pigmented actinic keratoses: A morphological study. *J Eur Acad Dermatol Venereol.* 2015;29:307-14, <http://dx.doi.org/10.1111/JDV.12532>.
21. Stolz W, Schiffner R, Burgdorf WHC. Dermatoscopy for facial pigmented skin lesions. *Clin Dermatol.* 2002;20:276-8, [http://dx.doi.org/10.1016/S0738-081X\(02\)00221-3](http://dx.doi.org/10.1016/S0738-081X(02)00221-3).
22. Kelati A, Baybay H, Moscarella E, Argenziano G, Galouj S, Mernissi FZ. Dermoscopy of Pigmented Actinic Keratosis of the Face: A Study of 232 Cases [Article in English, Spanish]. *Actas Dermosifiliogr.* 2017;108:844-51, <http://dx.doi.org/10.1016/J.AD.2017.05.002>.
23. Akay BN, Kocyigit P, Heper AO, Erdem C. Dermatoscopy of flat pigmented facial lesions: Diagnostic challenge between pigmented actinic keratosis and lentigo maligna. *Br J Dermatol.* 2010;163:1212-7, <http://dx.doi.org/10.1111/J.1365-2133.2010.10025.X>.
24. Lallas A, Tschandl P, Kyrgidis A, Stolz W, Rabinovitz H, Cameron A, et al. Dermoscopic clues to differentiate facial lentigo maligna from pigmented actinic keratosis. *Br J Dermatol.* 2016;174:1079-85, <http://dx.doi.org/10.1111/BJD.14355>.
25. Ertop Doğan P, Akay BN, Okçu Heper A, Rosendahl C, Erdem C. Dermatoscopic findings and dermatopathological correlates in clinical variants of actinic keratosis, Bowen's disease, keratoacanthoma, and squamous cell carcinoma. *Dermatol Ther.* 2021;34, <http://dx.doi.org/10.1111/DTH.14877>.
26. Nascimento MM, Shitara D, Enokihara MMSS, Yamada S, Pellacani G, Rezza GG. Inner gray halo, a novel dermoscopic feature for the diagnosis of pigmented actinic keratosis: Clues for the differential diagnosis with lentigo maligna. *J Am Acad Dermatol.* 2014;71:708-15, <http://dx.doi.org/10.1016/J.JAAD.2014.05.025>.
27. Martín JM, Bella-Navarro R, Jordá E. Vascular patterns in dermoscopy [Article in Spanish]. *Actas Dermosifiliogr.* 2012;103:357-75, <http://dx.doi.org/10.1016/J.AD.2011.11.005>.
28. Zalaudek I, Kreusch J, Giacomel J, Ferrara G, Catrical C, Argenziano G. How to diagnose nonpigmented skin tumors: A review of vascular structures seen with dermoscopy: Part II. Non-melanocytic skin tumors. *J Am Acad Dermatol.* 2010;63:377-86, <http://dx.doi.org/10.1016/J.JAAD.2009.11.697>.
29. Mir-Bonafé JF, Mir-Bonafé M, Mozos A, López-Sánchez C, Piquero-Casals J, de Dios-Velázquez Á, et al. Iceberg sign in actinic keratosis neglecta caused by toning shampoo for blonde and white hair. *Int J Dermatol.* 2020;59:e64-5, <http://dx.doi.org/10.1111/IJD.14717>.
30. Zalaudek I, Argenziano G, Leinweber B, Citarella L, Hofmann-Wellenhof R, Malveyh J, et al. Dermoscopy of Bowen's disease. *Br J Dermatol.* 2004;150:1112-6, <http://dx.doi.org/10.1111/J.1365-2133.2004.05924.X>.
31. Pan Y, Chamberlain AJ, Bailey M, Chong AH, Haskett M, Kelly JW. Dermatoscopy aids in the diagnosis of the solitary red scaly patch or plaque-features distinguishing superficial basal cell carcinoma, intraepidermal carcinoma, and psoriasis. *J Am Acad Dermatol.* 2008;59:268-74, <http://dx.doi.org/10.1016/J.JAAD.2008.05.013>.
32. Bugatti L, Filosa G, De Angelis R. The specific dermoscopic criteria of Bowen's disease. *J Eur Acad Dermatol Venereol.* 2007;21:700-1, <http://dx.doi.org/10.1111/J.1468-3083.2006.01995.X>.
33. Papageorgiou C, Apalla Z, Variaah G, Matiaki FC, Sotiriou E, Vakirlis E, et al. Accuracy of dermoscopic criteria for the differentiation between superficial basal cell carcinoma and Bowen's disease. *J Eur Acad Dermatol Venereol.* 2018;32:1914-9, <http://dx.doi.org/10.1111/JDV.14995>.
34. Mun JH, Kim SH, Jung DS, Ko HC, Kwon KS, Kim MB. Dermoscopic features of Bowen's disease in Asians. *J Eur Acad Dermatol Venereol.* 2010;24:805-10, <http://dx.doi.org/10.1111/J.1468-3083.2009.03529.X>.

- 537 35. Yang Y, Lin J, Fang S, Han S, Song Z. What's new in der-
538 moscopy of Bowen's disease: Two new dermoscopic signs and
539 its differential diagnosis. *Int J Dermatol.* 2017;56:1022–5,
540 <http://dx.doi.org/10.1111/IJD.13734>.
- 541 36. Payapvipapong K, Tanaka M. Dermoscopic classification
542 of Bowen's disease. *Australas J Dermatol.* 2015;56:32–5,
543 <http://dx.doi.org/10.1111/AJD.12200>.
- 544 37. Cameron A, Rosendahl C, Tschandl P, Riedl E, Kit-
545 tler H. Dermatoscopy of pigmented Bowen's disease.
546 *J Am Acad Dermatol.* 2010;62:597–604, <http://dx.doi.org/10.1016/J.JAAD.2009.06.008>.
- 547 38. Giuffrida R, Conforti C, Resende FSS, Hamilko de Barros M,
548 Uranitsch M, Favero F, et al. Clinical and dermoscopic fea-
549 tures of genital pigmented Bowen disease. *Clin Exp Dermatol.*
550 2018;43:813–6, <http://dx.doi.org/10.1111/ced.13633>.
- 551 39. Inskip M, Cameron A, Akay BN, Gorji M, Clark SP, Rosendahl
552 N, et al. Dermoscopic features of pigmented intraepi-
553 dermal carcinoma on the head and neck. *J Dtsch Der-
554 matol Ges.* 2020;18:969–76, <http://dx.doi.org/10.1111/DDG.14220>.
- 555 40. Jaimes N, Zalaudek I, Braun RP, Tan BH, Busam KJ, Marghoob
556 AA. Pearls of keratinizing tumors. *Arch Dermatol.* 2012;148:976,
557 <http://dx.doi.org/10.1001/ARCHDERMATOL.2011.3475>.
- 558 41. Rosendahl C, Cameron A, Argenziano G, Zalaudek I, Tschandl
559 P, Kittler H. Dermoscopy of squamous cell carcinoma
560 and keratoacanthoma. *Arch Dermatol.* 2012;148:1386–92,
561 <http://dx.doi.org/10.1001/ARCHDERMATOL.2012.2974>.
- 562 42. Sgouros D, Theofili M, Damaskou V, Theotokoglou S,
563 Theodoropoulos K, Stratigos A, et al. Dermoscopy as a
564 Tool in Differentiating Cutaneous Squamous Cell Carci-
565 noma From Its Variants. *Dermatol Pract Concept.* 2021;11,
566 <http://dx.doi.org/10.5826/DPC.1102A50>.
- 567 43. Manfredini M, Longo C, Ferrari B, Piana S, Benati E,
568 Casari A, et al. Dermoscopic and reflectance confocal
569 microscopy features of cutaneous squamous cell carci-
570 noma. *J Eur Acad Dermatol Venereol.* 2017;31:1828–33,
571 <http://dx.doi.org/10.1111/JDV.14463>.
- 572 44. Lallas A, Pyne J, Kyrgidis A, Andreani S, Argenziano G, Cavaller
573 A, et al. The clinical and dermoscopic features of invasive cuta-
574 neous squamous cell carcinoma depend on the histopathological
575 grade of differentiation. *Br J Dermatol.* 2015;172:1308–15,
<http://dx.doi.org/10.1111/BJD.13510>.
- 576 45. Suárez AL, Louis P, Kitts J, Busam K, Myskowski PL, Wong RJ,
577 et al. Clinical and dermoscopic features of combined cuta-
578 neous squamous cell carcinoma (SCC)/neuroendocrine [Merkel
579 cell] carcinoma (MCC). *J Am Acad Dermatol.* 2015;73:968–75,
580 <http://dx.doi.org/10.1016/J.JAAD.2015.08.041>.
- 581 46. Güleç AT. Diagnosing squamous cell carcinoma of the lip
582 using dermoscopy. *J Am Acad Dermatol.* 2017;76:S82–3,
583 <http://dx.doi.org/10.1016/J.JAAD.2016.10.026>.
- 584 47. Benati E, Persechino F, Piana S, Argenziano G, Lallas A,
585 Moscarella E, et al. Dermoscopic features of squamous cell
586 carcinoma on the lips. *Br J Dermatol.* 2017;177:e41–3,
587 <http://dx.doi.org/10.1111/BJD.15274>.
- 588 48. Lallas A, Martínez G, Arceu M, Kyrgidis A, Liopyris K, Bran-
589 caccio G, et al. Clinical and dermatoscopic predictors of
590 squamous cell carcinoma of the lips: A case-control, multi-
591 centric study. *J Eur Acad Dermatol Venereol.* 2022;36:222–7,
592 <http://dx.doi.org/10.1111/JDV.17790>.
- 593 49. Papageorgiou C, Lallas A, Manoli SM, Longo C, Lai M,
594 Liopyris K, et al. Evaluation of dermoscopic criteria for
595 early detection of squamous cell carcinoma arising on
596 an actinic keratosis. *J Am Acad Dermatol.* 2022;86:791–6,
597 <http://dx.doi.org/10.1016/J.JAAD.2021.03.111>.
- 598 50. Papageorgiou V, Apalla Z, Sotiriou E, Papageorgiou C, Lazaridou
599 E, Vakirlis S, et al. The limitations of dermoscopy: False-positive
600 and false-negative tumours. *J Eur Acad Dermatol Venereol.*
601 2018;32:879–88, <http://dx.doi.org/10.1111/JDV.14782>.
- 602 51. Papageorgiou C, Spyridis I, Manoli SM, Busila I, Nasturica IE, Lal-
603 las K, et al. Accuracy of dermoscopic criteria for the differential
604 diagnosis between irritated seborrheic keratosis and squa-
605 mous cell carcinoma. *J Am Acad Dermatol.* 2021;85:1143–50,
606 <http://dx.doi.org/10.1016/J.JAAD.2020.02.019>.
- 607 52. Yélamos O, Braun RP, Liopyris K, Wolner ZJ, Kerl K, Gerami
608 P, et al. Dermoscopy and dermatopathology correlates of
609 cutaneous neoplasms. *J Am Acad Dermatol.* 2019;80:341–63,
610 <http://dx.doi.org/10.1016/j.jaad.2018.07.073>.
- 611 53. Kreusch JF. Vascular patterns in skin tumors. *Clin Der-
612 matol.* 2002;20:248–54, [http://dx.doi.org/10.1016/s0738-081x\(02\)00227-4](http://dx.doi.org/10.1016/s0738-081x(02)00227-4).
- 613
614
615