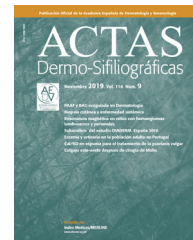




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REVIEW

Relation Between Atypical Fibroxanthoma and Pleomorphic Dermal Sarcoma: Histopathologic Features and Review of the Literature[☆]



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KEYWORDS

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Review

Abstract The relation between atypical fibroxanthoma and pleomorphic dermal sarcoma has led to confusion and debate in the literature. Both tumors present on sun-exposed skin, typically on the head and neck, in patients of advanced age. Both are comprised of a variable mix of histiocytoid, spindle, epithelioid, and/or giant multinucleated cells with pleomorphic nuclei. No immunohistochemical diagnostic techniques have emerged to distinguish these tumors. Diagnosis is by exclusion. Histologically, atypical fibroxanthoma is seen as a well-circumscribed dermal nodule but there will be no evidence of extensive subcutaneous invasion, tumor necrosis, or lymphovascular or perineural invasion. Therefore, if any of the aforementioned features is present, the diagnosis would be pleomorphic dermal sarcoma. This narrative review of the literature aims to identify the distinguishing and overlapping histopathologic features of these 2 tumors as they have been described in case series.

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PALABRAS CLAVE

Fibroxantoma atípico;
Sarcoma pleomórfico dérmico;
Relación histopatológica;
Revisión

Relación entre fibroxantoma atípico y sarcoma pleomórfico dérmico: histopatología de ambos y revisión de la literatura

Resumen La relación entre el fibroxantoma atípico (FXA) y el sarcoma pleomórfico dérmico (SPD) ha sido confusa y objeto de debate a lo largo de los años en la literatura científica. Son tumores que se presentan en pacientes de edad avanzada en piel fotoexpuesta, típicamente cabeza y el cuello. Están formados por una mezcla variable de células histiocitoides, fusiformes,

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epitelioides y gigantes multinucleadas con núcleos pleomórficos. No existen técnicas inmunohistoquímicas diagnósticas de estas entidades y su diagnóstico debe ser de exclusión. El FXA es una neoplasia dérmica, bien delimitada, con ausencia de infiltración difusa de tejido subcutáneo, necrosis tumoral o invasión linfovascular o perineural. Estando alguna de las características anteriores presente, debe hacerse el diagnóstico de SPD. En esta revisión narrativa de la literatura intentaremos determinar cuáles son las características histopatológicas precisas de ambas entidades según las series publicadas en la literatura y aquellos aspectos que las diferencian o relacionan.

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Introduction

The relation between atypical fibroxanthoma (AFX) and pleomorphic dermal sarcoma (PDS) has led to confusion and debate in the literature over the years.¹ Both tumors present in patients of advanced age, predominantly in men, and mainly on sun-exposed skin and on the head and neck, with lesions that are often ulcerated (Fig. 1).² Both are comprised of a variable admixture of histiocytoid, spindle-shaped, epithelioid, and multinucleated giant cells with pleomorphic nuclei (Fig. 2) often with abundant aberrant mitosis. There are no immunohistochemical diagnostic techniques for these entities and diagnosis should be by ruling out other possibilities.¹

These are uncommon tumors and their incidence is unknown, although the incidence of AFX in one Spanish health area has recently been estimated in the literature as 0.59 cases/100 000 inhabitants,³ which contrasts with the incidence of other tumors in Spain, such as basal cell carcinoma (113.05/100 000 inhabitants), squamous cell carcinoma (38.16/100 000), and melanoma (8.76/100 000 inhabitants).⁴ There are no figures available for the incidence of PDS.



Figure 1 Exophytic ulcerated tumor on the right temple, consistent with atypical fibroxanthoma after histopathologic study.

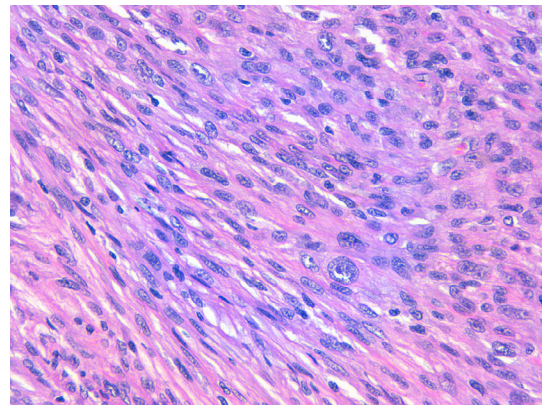


Figure 2 Detail of an admixture of epithelioid and spindle-shaped cells in an atypical fibroxanthoma (H&E, 40x).

Their etiology, in most cases, is related to chronic exposure to sunlight. The implication of ultraviolet light in their etiopathogenesis is supported by the demonstration of *p53* mutations.⁵

The current histopathologic definition of AFX includes the following characteristics¹:

- Well-circumscribed dermal neoplasm, comprised of an admixture of histiocytoid, spindle-shaped, epithelioid, and multinucleated giant cells with pleomorphic nuclei
- Exophytic, nodular, or polypoid growth (Fig. 3a)
- Absence of diffuse infiltrate of subcutaneous cellular tissue, tumor necrosis, or lymphovascular or perineural invasion
- Diagnosis by exclusion based on analysis of the entire resection piece, and after ruling out the main differential diagnoses—squamous-cell, sarcomatous, or spindle-cell carcinoma, spindle-cell melanoma, poorly differentiated leiomyosarcoma, and angiosarcoma—with an appropriate immunohistochemical panel
- Possible presence of an epidermal collarette
- The deep margin is usually predominantly expansive

PDS has histopathologic features similar to AFX, but with diffuse invasion of subcutaneous cellular tissue or deep structures, tumor necrosis, or lymphovascular or perineural invasion⁶ (Fig. 3b). When these diagnostic criteria are strictly applied, the clinical behavior of AFX is considered

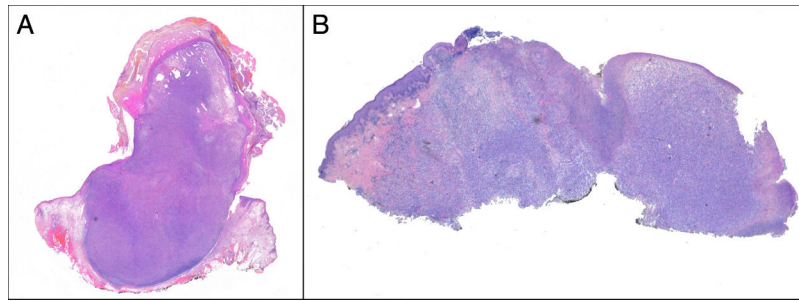


Figure 3 A, Atypical fibroxanthoma with polypoid morphology. Well-circumscribed dermal tumor with no invasion of subcutaneous cellular tissue (H&E). B, Pleomorphic dermal sarcoma under a magnifying glass. Extensive tumor tissue with involvement of the deep margin (H&E).

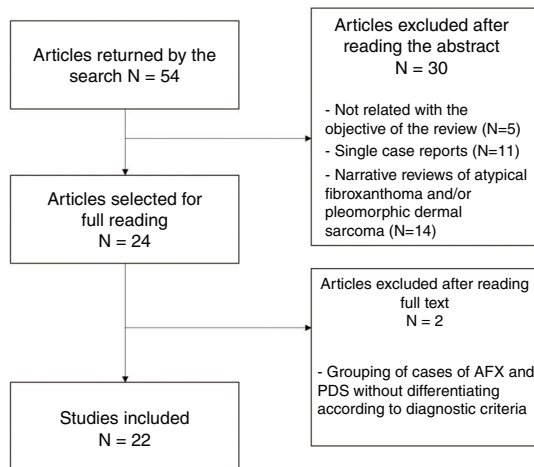


Figure 4 Flow diagram of the review strategy.

benign, with infrequent local recurrence. Those AFX lesions reported with metastases in the literature are probably other tumors that have been incorrectly diagnosed. The terms undifferentiated pleomorphic sarcoma of the skin and superficial and deep malignant fibrous histiocytoma are poorly documented and not currently used to refer to these tumors.²

Although AFX and PDS are tumors that have been studied in detail, there is still some debate in the scientific literature as to how these entities are related and there are some outstanding questions, such as whether AFX can transform into PDS over time, with the 2 belonging to the same entity and with a prognosis depending on the depth of the neoplasm.¹ In this literature review, we will attempt to determine the exact histopathologic features of the 2 entities, according to series published in the literature and those aspects that link them together.

Material and Methods

For this review, a literature search was performed in the PubMed (Medline), Cochrane Library, Embase, SciELO, and Trip databases with the following terms: “atypical fibroxanthoma” AND “pleomorphic dermal sarcoma,” on September 4, 2020. The initial search retrieved 126 results (53 after eliminating duplicates). The summaries and

abstracts were read and screened according to the following criteria:

- Criteria for inclusion in the review:
 - Case series published in peer-reviewed journals in the last 10 years, which included AFX and/or PDS, with both tumors described according to currently used terms. Additionally, series that deal with the relationship between these 2 tumors, according to a given feature.
- Exclusion criteria:
 - Not related with the objective of the review (5 publications)
 - Isolated case reports (11 publications)
 - Narrative reviews of AFX and/or PDS (14 publications)

In total, 24 studies were selected for the full text to be read; of these, 22 were finally included in the present review (Fig. 4). Two studies were excluded because of the grouping of AFX and PDS cases, without applying diagnostic criteria for differentiation between them.

Of the series included, the following data were collected: patients and tumor, site, diagnostic criteria used, pattern of invasion, size, mitotic index, ulceration, necrosis, perineural invasion, lymphovascular invasion, involvement of the deep margin, variants, immunohistochemistry (negative markers), prognosis, and relationship between tumors.

Results

The results of the series included for AFX,^{3,7-10} PDS,^{2,6} and mixed entities¹¹⁻²⁵ are summarized in Table 1.

The series are in general small, with few patients, rarely exceeding 30 tumors for each type. The site was mainly the head and neck, with all tumors reported at this site in many of the series.

Differences Between AFX and PDS

AFX and PDS were differentiated in most of the series with histopathologic criteria. In the series reported by Griewank et al.²⁴ in 2014, differentiation between AFX and PDS was performed by 2 dermatopathologists and in a subsequent analysis it was found that necrosis, invasion of subcutaneous tissue, invasion of fascia/muscle, involved margin,

Table 1 Case Series Included in the Review.

Study	Patients and Tumors	Site	Diagnostic Criteria Used	Invasion Pattern	Tumor Size	Mitotic Index	Presence of Ulceration	TN	Involvement of Deep Margin	Variants Cytology	Immunohistochemistry (Negative Markers)	Prognosis	Relationship Between Tumors
Miller et al. (2012) ²	32 PDS	Head (96.8%)	Invasion of deep SCT (at least deep areas of subcutaneous tissue, muscle, fascia or galea), and/or TN, LVI, PNI	Infiltrative (59.3%)	Median 25 mm (range 7 to 60 mm)	19.5 mitosis/10 HGF (range 6 to 51)	25 (78%)	TN (53%)	34.3%	Classic 40.6%	AE1/3	Median follow-up 24 months (available for 29 patients)	NA
		Forearm (3.2%)	S100, CD34 and desmin negative	Expansive (37.5%) Not determined (3,1%)					PNI (28.1%) LVI (25%)	Myxoid changes 18.7% Pseudoangiomatic hemorrhagic 9.3%	Spindle-cell predominance 59.3% cam5.2 34bE12 Storiform patterns 6.2% Keloid changes 6.2% Telangiectatic spaces 3.1% Osteoclast-like 3.1%	MNF116 Met 10%	LR 28%
Thum et al. (2013) ¹¹	11 AFX	HaN (100%)	Centered on the dermis No evidence of epidermal precursor	Expansive (number of cases NS)	Median 12 mm (range 7 to 23)	High (without specifying tumor type)	10 cases (without specifying tumor type)	Absent	NS (at least 1)	Pseudoangiomatic hemorrhagic (100%)	AE1/AE3 MNF116 CK14 CK5/6 S100 melan A HMB-45 Desmin p63	Median follow-up 43.1 months (10 patients) LR 0%	Both tumors negative for ERG and CD34
	3 PDS*		Invasion of underlying fascia	NS	Median 35 mm (range 20 to 40)				NS (at least 1)				
Hollmig et al. (2013) ²⁶	8 PDS	HaN (100%)	Tumor invading SCT	NS	NS	NS	NS	NS	NS (4 cases with biopsy insufficient for assessment)	NS	NS	Median follow-up 19.5 months	Similar immunohistochemical features
	14 AFX		Tumor limited to the dermis					PNI LVI				LR 62.5%	LN-2 is not a good diagnostic or prognostic marker in these tumors
Zschoche et al. (2014) ¹⁸	25 AFX	HaN (100%)	Pattern of exophytic growth	NS	NS	NS	NS	NS	NS	NS	NS	Met 25% Mean follow-up 43 months LR 14.2% Met 14.2% NS	Both tumors have a similar density of intratumoral lymphatic vessels

Table 1 (Continued)

Study	Patients and Tumors	Site	Diagnostic Criteria Used	Invasion Pattern	Tumor Size	Mitotic Index	Presence of Ulceration	TN	Involvement of Deep Margin	Variants/Cytology	Immunohistochemistry (Negative Markers)	Prognosis	Relationship Between Tumors
			No invasion of SCT					PNI LVI					There are no significant differences between AFX and PDS when the 3 lymphatic subgroups are compared
Nonaka, Bishop (2014) ¹⁹	22 PDS	HaN (95.5%) Arm (4.5%)	TSC invasion Tumors in the deep dermis without exophytic growth pattern, still without SCT involvement										
	19 AFX	HaN (100%)	Tumor limited to the dermis	NS (data grouped by AFX and PDS)	NS (data grouped by AFX and PDS)	NS (data grouped by AFX and PDS)	NS (data grouped by AFX and PDS)	NS (data grouped by AFX and PDS)	NS	NS	AE1/AE3	NS (data grouped by AFX and PDS)	Not evaluated because pooled data are presented
	34 PDS		Tumor extends to the SCT								MNF116 34bE12 CK5/6 CK14 p63		
Griewank et al. (2014) ²⁰	27 AFX	HaN (96.2%)	Absence of invasion of SCT, TN, PNI, and LVI	Expansive (100%)	Median 9 mm (range 4-30)	Median 20 mitosis/mm ² (range 7-53)	44.4%	PNI LVI Absent	NS	NS	MNF116	NS	Mutations in the TERT promotor are present in 93% of AFX and 76% of PDS. The number of CC > T mutations suggests a pathogenic role for UV light in both tumors
		Unknown (3.8%)									AE1/ AE3 CD31 (focal in 1 case) CD34 S100 Desmin		
	34 PDS	HaN (94.2%)	Presence of invasion of SCT, TN, PNI, or LVI	Invasive (50%)	Median 22 mm (range 6-60)	Median 19 mitosis/mm ² (range 5-51)	76.4%	TN (55.8%)					
		Unknown (5.8%)		Expansive (47%) Not evaluable (2.9%)				PNI (26.4%) LVI (26.4%)					
Harding-Jackson et al. (2015) ⁷	15 AFX	HaN (86.6%)	Circumscribed with expansive border	Expansive (100%)	NS	Range 5-38 per 10 HGF	60%	Absent	NS	NS	Desmin Calponin h-caldesmon	At least 2 years of follow-up	NA
		Unknown (13.3%)	Absence of TN, PNI, LVI or deep extension to SCT								S100	LR 6.6% (1 case)	
											P63 Broad spectrum CK CD31 CD34	Met 0%	

Table 1 (Continued)

Study	Patients and Tumors	Site	Diagnostic Criteria Used	Invasion Pattern	Tumor Size	Mitotic Index	Presence of Ulceration	TN	Involvement of Deep Margin	Variants/ Cytology	Immunohistochemistry (Negative Markers)	Prognosis	Relationship Between Tumors
Wang et al. (2015) ⁸	11 AFX	HaN (100%)	Include tumors with SCT, TN, and PNI invasion	Invasive (90.9%)	Median 8 mm (range 3-18)	Median 20 mitosis per 10 HGF (range 10-55)	27.27%	PNI LVI	NS	Spindle-cell (6/11)	NS	Selection of 9 cases with metastasis in 152 (5.9%) + 2 selected from another center 2 cases without SCT involvement metastasized	NA
				TN 18.18%									
				PNI 45.45%									
Tardío et al. (2016) ²¹	18 PDS	HaN (100%)	Invasion of deep subcutaneous tissue and/or TN and/or PNI and/or LVI	Expansive (17%)	Median 15 mm (range 7-70)	Median of 22 mitosis per 5 mm ² (range 2-126)	55.5%	TN (17%)	22.2%	Spindle-cell and epithelioid (1/11) Spindle-cell and pleomorphic (1/11) Pleomorphic (1/11) Spindle-cell fascicular	Cytokeratins	Median follow-up of 33 months (available in 15/18)	Same histopathologic and immunohistochemical features, except for lack of invasion of deep SCT, TN, or LVI in AFX Only differences in prognosis
				Invasive (83%)									
				LVI (17%)									
Helbig et al. (2016) ²²	45 AFX with follow-up of more than 12 months (for comparison)	HaN (100%)	Absence of deep subcutaneous tissue invasion, TN, PNI, or LVI	NS	Median 11 mm (range 4-30)	NS	NS	PNI LVI Absent	NS	NS	NS (the same)	Follow-up of more than 12 months (median 48 months) LR 2.2% (1 case) No Met NS	All AFX and PDS presented similar oncogene expression (overexpression of TP53, CCND1, CDK4) One case with AFX and PDS showed mutational profiles of TP53 and PIK3CA, identical in both tumors
				NS									
				NS									
Helbig et al. (2016) ²²	5 AFX	Thigh (20%)	Limited to the dermis	NS	NS	NS	NS	NS	NS	NS	At least 1 cytokeratin (CK 5/6 or pancytokeratin)	NS	
		HaN (80%)											
											Two melanocytic markers (S100, melan A, or HMB-45)		

Table 1 (Continued)

Study	Patients and Tumors	Site	Diagnostic Criteria Used	Invasion Pattern	Tumor Size	Mitotic Index	Presence of Ulceration	TN	Involvement of Deep Margin	Variants/ Cytology	Immunohistochemistry (Negative Markers)	Prognosis	Relationship Between Tumors
								PNI LVI					
Helbig et al. (2017) ²³	6 PDS (1 patient with AFX and PDS separated by 3 years) 5 AFX	HaN (100%) HaN (80%)	TSC invasion NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	Amplifications and deletions detected in 6 of 27 PDS; these were not detected in AFX
Griewank et al. (2018) ²⁴	26 PDS 41 AFX	Unknown (20%) HaN (96.1%) Shoulder (3.9%) HaN (90.2%)	Established after analysis	Invasive (2.4%)	Median 8 mm (range 4-30)	Median of 17 mitosis/mm ² (3-52)	34.1%	TN 2.4% (1 case included)	NS	NS	Pancytokeratin (MNF116, AE1/AE3) CD31, or CD34	NS	AFX and PDS present recurrent mutations in FAT1, NOTCH1/2, CDKN2A, TP53, and the TERT promoter
		Arm (4.8%)	SS association of PDS with TN, SCT invasion, invasion of fascia/muscle, involved border, and LVI	Expansive (73.1%)			Not determined 17.1%	No PNI			S100		
		Unknown (4.8%)		Not evaluable (24.3%)				No LVI			Desmin Smooth muscle actin melan A (1 case of aberrant expression)		
	40 SPD**	HaN (92.5%)		Invasive (40%)	Median 20 (range 4-60)	Median of 21 mitosis/mm ² (range 5-44)	55%	TN 40%					
		Arm (2.5%)		Expansive (25%)			Not determined 25%	PNI 22.5%					
		Unknown (5%)		Not evaluable (35%)				LVI 20%					
Helbig et al. (2018) ²⁵	25 AFX	HaN (96%)	No invasion of SCT	NS	NS	NS	NS	NS	NS	NS	AE1/AE3	NS (only available for the 2 patients with <i>MitF</i> expression: free of tumor progression)	Expression of <i>MitF</i> in 1/25 AFX and 1/25 PDS; expression of α -SMA in 40% of AFX and 36% of PDS
		Leg (4%)									CK5/6 p40 SOX10 ERG CD10+ necessary		
	25 PDS***	HaN (92%) Shoulder (8%)	Invasion of SCT, fascia, or muscle										

Table 1 (Continued)

Study	Patients and Tumors	Site	Diagnostic Criteria Used	Invasion Pattern	Tumor Size	Mitotic Index	Presence of Ulceration	TN PNI LVI	Involvement of Deep Margin	Variants/ Cytology	Immunohistochemistry (Negative Markers)	Prognosis	Relationship Between Tumors
Gaiser et al. (2018) ¹³	51 AFX	HaN (100%)	NS	NS	NS	NS	NS	NS	NS	NS	NS	LR 1.9% (1 case)	MYC amplification is not a determining genetic factor in the process of tumor genesis for AFX or PDS
Koelsche et al. (2019) ¹⁴	24 PDS	HaN (91.7%) Shoulder (8.3%)	NS	NS	NS	NS	NS	NS	NS	NS	NS	No Met NS	The DNA methylation profile does not distinguish between AFX and PDS Loss of 9p and 13q and gain of 8q with similar frequency Homozygous deletion of CDKN2A more frequent in PDS than AFX
	17 AFX	HaN (100%)										NS	
Müller et al. (2019) ⁹	15 PDS 41 AFX (40 patients)	HaN (93.3%) Unknown (6.6%) HaN (90.3%) Limbs (9.7%)	NS	NS	Mean of 5.7 cm ² (range 0.06 and 40)	Median 4.8 mitosis/10 HGF (range 1-50.4)	68.2%	TN 14.6% No PNI	NS	NS	NS	LR 7.3% No Met	NA High expression of SEC62 in tumors with necrosis, size > 5 cm ² , high Clark levels
Nassios et al. (2019) ¹⁵	4 AFX	NS	NS	NS	NS	NS	NS	No LVI NS	NS	NS	NS	NS	Similar expression of GPR4, TDAG8, OGR1, and G2A in AFX and SPD studied

Table 1 (Continued)

Study	Patients and Tumors	Site	Diagnostic Criteria Used	Invasion Pattern	Tumor Size	Mitotic Index	Presence of Ulceration	TN PNI LVI	Involvement of Deep Margin	Variants/ Cytology	Immunohistochemistry (Negative Markers)	Prognosis	Relationship Between Tumors
Miller et al. (2020) ¹⁶	5 PDS 21 AFX	HaN 90.5%	Atypical spindle-cell proliferation in sun-damaged skin with minimal or no SCT invasion, without TN, LVI, PNI, or epithelial/melanocytic/vascular/smooth muscle differentiation by morphology or immunostaining	NS	Median 0.9 mm (range 0.2-2.8)	NS	NS	Absent	NS	NS	Cytokeratins	Follow-up in 7 patients	Mutations detected in <i>PIK3CA</i> (and not in PDS), more frequent <i>CDKN2A</i> deletions in PDS compared with AFX, <i>CDKN2a</i> mutations in AFX were not observed in PDS but were observed in AFX, increased mutations in the TERT promotor in PDS compared with AFX.
		Knee 4.7% Chest 4.7%									S100 Others (see supplementary table in original article)		
	17 PDS	HaN 88.2%	Presence of deep subcutaneous invasion, necrosis, lymphovascular invasion, and/or perineural invasion		NS	NS	NS	NS	NS	NS		Follow-up in 2 patients (both LR)	
		Thigh 5.8% Supraclavicular 5.8%											
Lonie et al. (2020) ⁶	27 PDS	HaN (96.3%)	Cytokeratins, S100, and CD34 negative, without specifying other histopathologic features	NS	NS	Numerous 35%	NS	NS	NS	NS	Cytokeratins	Median follow-up of 46.4 months	NA

Table 1 (Continued)

Study	Patients and Tumors	Site	Diagnostic Criteria Used	Invasion Pattern	Tumor Size	Mitotic Index	Presence of Ulceration	TN	Involvement of Deep Margin	Variants/ Cytology	Immunohistochemistry (Negative Markers)	Prognosis	Relationship Between Tumors
		Trunk (3.7%)				Frequent 33.3%		PNI LVI			S100 CD34	LR 7.3% Met 3.7% (1 case) NS	Cathepsin-K was moderately positive and diffuse in all cases of AFX and PDS
Ricci et al. (2020) ¹⁷	5 AFX	HaN (100%)	NS	NS	NS	NS	NS	NS	NS	NS	Cytokeratins		
Bitel et al. (2020) ¹⁰	4 PDS 105 AFX	HaN (50%) Trunk (50%) HaN (97.9%)	NS	NS	15 mm +/- 3 mm	High 63%	47.6%	NS	NS	Spindle-cell 70%	NS	Mean follow-up of 30 months in 36 patients LR 22.9%	NA
	(85 patients)	Shoulder (2.1%)	Includes tumors invading muscle/fascia/cartilage			Intermediate 17.8%				Mixed 30%		No Met	Identification as a risk factor for recurrence of invasion of deep structures
Iglesias-Pena et al. (2020) ³	62 AFX	HaN (96.8%) Trunk (3.2%)	Invasion of deep structures Compatible immunohistochemical panel Absence of TN, PNI, and LVI	NS	Mean 12.3 mm (range 3-40)	Median of 7.09 mitosis/10 HGF (range 0-31)	50%	Absent	8.1%	Classic 88.7% Nonpleomorphic spindle-cell 4.8% Hemosiderotic 4.8% Keloid 4.8%	AE1-AE3 CD34 HMB45 Desmin EMA S100 (1 focal)	Median follow-up of 34 months LR 6.5% (4 cases) No Met	NA

Abbreviations: AFX, atypical fibroxanthoma; HaN, head and neck; HPF, high power field; LR, local recurrence; LVI, lymphovascular invasion; Met, metastasis; NA, not applicable; NS, not specified; PDS, pleomorphic dermal sarcoma; PNI, perineural invasion; SCT, subcutaneous cellular tissue; SS, statistically significant; TN, tumor necrosis.

*Included in the series of Miller et al. (2012).

**20 of these included in the series of Miller et al. (2012).

***Four of these included in the series of the same author Helbig et al. (2016).

and lymphovascular invasion are criteria significantly associated with PDS.

The diagnostic criteria used to differentiate between AFX and PDS are not homogeneous in the studies, and often they are not clearly specified. The tumors with tumor necrosis, lymphovascular invasion, perineural invasion, or invasion of the fascia/muscle were generally classified as PDS, although some series classified tumors with some of these features as AFX.^{8-10,24}

Invasion of subcutaneous cellular tissue deserves special mention for differentiating between AFX and PDS. In some series,^{12,18-20,22,25} simple invasion of subcutaneous cellular tissue is automatically considered a criterion for PDS whereas in others,^{2,7,11,21} this infiltration should be *deep*, in some cases without specifying how deep. One series went as far as to consider PDS those dermal tumors that did not invade subcutaneous tissue because they did not have an exophytic pattern of growth (Table 2).¹⁸

Many series did not specify the pattern of invasion of the tumors. When this was specified, most AFX had an expansive pattern, whereas the patterns reported for PDS had a mainly infiltrative pattern of growth.^{2,7,20,21,24} Of note is the percentage of AFX with invasive pattern in the series of Wang (90.9%), with 11 metastatic AFX lesions, which included tumors with extensive invasion of subcutaneous tissue, tumor necrosis, and perineural infiltration; these are criteria that should trigger reclassification as PDS.⁸

Tumor size^{11,20,24} and mitotic index,^{20,24} when reported, are clearly larger in PDS than AFX. Likewise, ulceration is reported more often in PDS in studies in which this feature is reported separately.^{20,24}

In the series of AFX by Müller et al.,⁹ a significantly larger increase in *SEC62* expression is reported in tumors with necrosis and a trend to greater expression in tumors with high Clark levels and tumor size greater than 5cm².

At the genetic level, of note is the series published in 2020 by Miller et al.,¹⁶ who reported mutations in *PIK3CA* (and not in PDS), more frequent *CDKN2A* deletions in PDS compared with AFX, *CDKN2a* mutations in AFX that were not observed in PDS, and increased mutations in the *TERT* promoter in PDS compared with AFX.

Finally, in terms of prognosis, differences between AFX and PDS are noteworthy. In series that included both types of tumor,^{21,26} local recurrence and metastases occur markedly more frequently in PDS. Of note is that the series that report metastatic AFX with specification of histopathologic features^{8,10} include tumors with invasion of deep structures, tumor necrosis, or perineural invasion (which, according to the current criteria, would be considered PDS).

Similarities between AFX and PDS

In this review, we can identify similarities between AFX and PDS in the different series that compare these tumors.

In the immunohistochemical studies, no study identified a marker that was expressed differently in the 2 types of tumor.²⁶ The immunohistochemical markers used vary from one study to another, but most include some cytokeratins, S100, and vascular markers (CD31, CD34), although this is not always clearly specified.^{8-10,12-15,18,23}

The investigations by Zschoche et al.¹⁸ did not show any differences in lymphatic architecture in more than 20 examples of each type of tumor.

In the series of Nonaka and Bishop,¹⁹ AFX and PDS are presented grouped as sarcoma-like tumors, with no immunohistochemical evidence of epithelial differentiation and no histologic signs of a squamous-cell carcinoma component. This meant that the characteristics of these tumors could not be assessed separately for the purposes of this review. Interestingly, these authors showed that the prognosis for AFX and PDS, when these tumors are grouped together, was similar to that of other sarcoma-like tumors with an epithelial component, suggesting that at least some cases of AFX and/or PDS could be related to squamous-cell carcinoma, with the former representing complete loss of epithelial phenotype.

At the genetic level, the series by Griewank et al.²⁰ published in 2018 shows that mutations in the *TERT* promoter are present in 93% of AFX and in 76% of PDS. The number of CC > TT mutations suggests a pathogenic role for UV light in both tumors. In the series reported by Helbig et al.,²² AFX and PDS were found to have a similar expression of oncogenes, with overexpression of *TP53*, *CCND1*, and *CDK4*, leading the authors to reaffirm their hypothesis that AFX is the noninvasive precursor of PDS. In another series reported by Helbig et al.²⁵ in 2018, with 25 AFX and 25 PDS lesions, the authors found *MiTF* expression in one AFX and one PDS, as well as expression of α -SMA in 40% of AFX and 36% of PDS. Gaiser et al.¹³ also ruled out amplification of the *MYC* gene as an important process in tumor genesis of AFX and PDS. Koelsche et al.¹⁴ demonstrated that the DNA methylation profile does not distinguish between AFX and PDS; in their study they observed loss of 9q and 13q and gain of 8q in a similar frequency in both tumors. Homozygous *CDKN2A* deletion was most frequent in PDS (6/15) compared with AFX (2/17), although the sample size was relatively small.

Nassios et al.¹⁵ studied expression of proton-sensitive protein G coupled receptors and found similar expression of *GPR4*, *TDQAG8*, *OGR1*, and *G2A* in the AFX and PDS studied.

Discussion

Except for the aforementioned histopathological criteria (invasion of deep structures, tumor necrosis, perineural invasion, and lymphovascular invasion), it is not clear where the differences between AFX and PDS actually lie. The scientific literature is very confusing due to the different terminology, different diagnostic criteria used by the authors, and insufficient immunohistochemical characterization of the tumors,²⁷ especially in older articles.²⁸ It is essential to perform an exhaustive assessment of resected pieces when such tumors are suspected, as the final diagnosis has implications in the subsequent management of the patients.²⁹

Variants of AFX have been described in the literature and knowledge of these is important to avoid diagnostic errors. These include nonpleomorphic spindle-cell AFX, clear-cell AFX, hemosiderotic AFX, myxoid AFX, AFX rich in osteoclast-like giant cells, keloid AFX, and granular cell AFX.¹ These changes have also been reported in some PDS, either involving the tumor areas or the entire lesion.²

Table 2 Differences in Considering AFX and PDS, According to Subcutaneous Tissue Invasion in Some of the Case Series Included.

Simple Invasion of SCT as Criterion for PDS	Deep Invasion of SCT Required to Consider PDS Diagnosis
Hollmig et al. (2013) ²⁶	Miller et al. (2012) ²
Zschoche et al. (2014) ¹⁸ *	Thum et al. (2013) ¹¹
Nonaka, Bishop (2014) ¹⁹	Harding-Jackson et al. (2015) ⁷
Griewank et al. (2014) ²⁰	Tardío et al. (2016) ²¹
Helbig et al. (2016) ²²	
Helbig et al. (2018) ²⁵	

*PDS was considered even without SCT invasion if tumor growth was not exophytic.

Abbreviations: PDS, pleomorphic dermal sarcoma; SCT, subcutaneous cellular tissue.

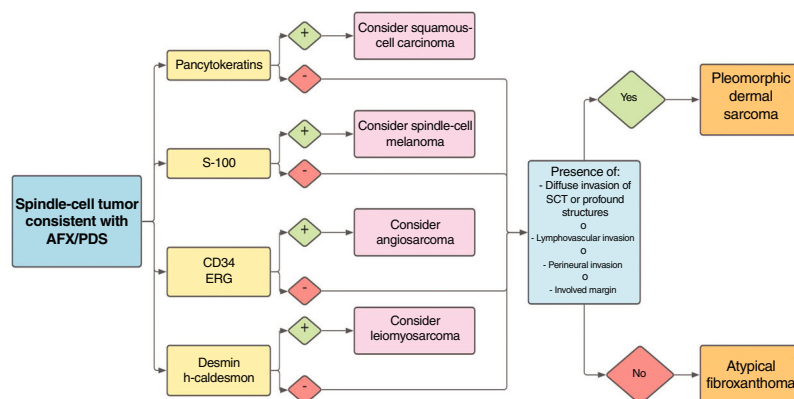


Figure 5 Diagnostic algorithm for atypical fibroxanthoma and pleomorphic dermal sarcoma. Abbreviations: AFX, atypical AFX; PDS, pleomorphic dermal sarcoma.

In AFX, a chronic inflammatory infiltrate can be observed, above all in the tumor periphery and the presence of severe solar elastosis is usually observed in the adjacent dermis.⁵

The immunohistochemical markers that can help differentiate between AFX and other tumors include CD99, S-100, CD34, cytokeratins, desmin, CD10, vimentin, HMB-45, CD68, and p63, among others.⁹ Differential diagnosis of these tumors with angiosarcoma is important and CD34 and ERG have been shown to be useful in this context, particularly in AFX-PDS with pseudoangiomatous/hemorrhagic pattern (Fig. 5).¹¹ To date, attempts to differentiate between AFX and PDS with immunohistochemical markers (CD99, LN-2) have not proved successful.^{12,30}

Strong positivity for CD10 in AFX and PDS can support diagnosis. However, it is important to note that any tumor with spindle-cell morphology can be positive for this marker, including sarcomas such as mixofibrosarcoma.⁵

It is also important to highlight that dendritic cells present in biopsies of AFX and PDS are positive for S100, but the tumor cells should be negative for this marker to make a diagnosis.⁵

Of note in this sense is the series of by Müller et al.,⁹ in which a significant increase in *SEC62* expression was reported in tumors with necrosis and a trend to greater expression in tumors with high Clark levels and tumor size greater than 5 cm². Although the histopathologic features for diagnosis of AFX are not specified in the study, the fact that large tumors with necrosis and subcutaneous invasion were included suggests that, according to current criteria,

some tumors would actually be PDS. This increased expression of *SEC62* in PDS compared with AFX could be the object of new lines of research in the future, with inclusion of both types of tumor with stricter diagnostic criteria.

Given that diagnosis of AFX and PDS is by exclusion, negative immunohistochemical markers suggestive of other entities are those that are used for ruling them out. In the literature, the number and type of markers that need to be negative for diagnosis have not been established. Thus, in many case series, immunohistochemical characterization of tumors is insufficient and can, in actual fact, correspond to other entities.³¹

It is worth asking whether, faced with tumors as closely related as AFX and PDS, it makes sense to continue using different terms. In situ melanoma does not have the same prognosis as ulcerated melanoma, with lymphovascular invasion and microsatellitosis, but we still refer to both lesions as melanoma. If to date, no significant differences have been found at the cellular, immunohistochemical, genetic, or molecular level, it makes sense to group these tumors in the same spectrum and use the same term to describe them. This is what Winchester et al.³² did in a series of 319 patients with tumors encompassed by the term *undifferentiated pleomorphic sarcoma*, which included cases of AFX and PDS grouped together, and for this reason, they were not included in this review. In that series, local recurrence was reported in 45 patients (14.1%) and distant metastasis in 33 patients (10.3%). After a complete analysis, the authors concluded that the aggressive behav-

ior in these tumors grouped together depends on invasion beyond subcutaneous fat, tumor size greater than 2 cm, immunosuppression, and presence of lymphovascular invasion. Another similar approach was taken by Cesinaro et al.²⁸ in a series of 71 tumors with features consistent with AFX or PDS grouped together and with a follow-up time of between 17 and 125 months. Like the previous study, this one was not included in our review. Only 4 local recurrences were reported and there were no cases of metastasis. The spindle-cell morphology was associated with subcutaneous invasion and recurrence. Some authors consider AFX and PDS to be on the same spectrum of tumors, although they continue to use the individual terms to differentiate between them.³³

With regard to subcutaneous invasion, we have shown in this review that the criteria for determining when it is present are by no means homogeneous. These variations of criteria when classifying the tumors have probably led to overdiagnosis of either AFX or PDS, depending on the series, with the subsequent impact on the rates of recurrence and/or metastasis. In the literature, AFX has been defined as a dermal neoplasm limited to the dermis or with *minimal* invasion of subcutaneous cellular tissue. The exact limit of the extent of invasion required to safely denote the tumor as AFX has never been uniformly established.^{34,35} According to the classification of the World Health Organization, initially it is specified that AFX is limited to the dermis, *without invasion of the subcutaneous cellular tissue* to then affirm in the differential diagnosis that lesions that resemble AFX but are larger or show *substantial invasion of subcutaneous cellular tissue* or beyond that, perineural invasion, lymphovascular invasion, or necrosis, should be classified as dermal pleomorphic sarcoma.³⁶ On the other hand, and although this is an infrequent situation, cases have been described of AFX without subcutaneous invasion that lead to recurrence with invasion of deeper structures³ or even metastasis⁸; so though uncommon, it is not impossible.

Finally, diagnosis of AFX is currently considered to be performed based on an entire resection piece and not with partial biopsies, given that it is very important to assess the pattern of invasion, invasion of structures, and the other criteria for PDS.¹ Although it was attempted to gather information systematically, the only study that specifies the number of cases diagnosed with involvement of the deep margin is that of Iglesias-Pena et al.,³ who also described 2 cases of spontaneous regression after an incisional biopsy. This is relevant because diagnosis of AFX and/or PDS, when the deep margin is positive, always generates a certain degree of uncertainty. It is necessary to establish some clear diagnostic criteria to classify tumors suggestive of AFX that regress after biopsy or incorrectly resected tumors, with involved margins that do not allow distinction between AFX and PDS, particularly when later margin widening gives a negative result.

Conclusions

The series of AFX and PDS published in recent years classify these tumors mainly according to histopathologic features of subcutaneous invasion, tumor necrosis, perineural

invasion, and lymphovascular invasion. The series of AFX that specify strict histopathologic criteria show that this is a benign neoplasm, but we lack larger series of patients with PDS to allow us to clearly establish what characteristics are associated with local recurrence or metastasis. Data published to date do not allow clear differentiation between these 2 tumors at other levels, and so some authors consider them the same entity that can follow a more aggressive course according to certain features of poor prognosis. Further studies are needed with large series of patients and careful description of histopathologic features to allow us to establish more rigorously factors of poor prognosis and thus help us choose the most appropriate treatment for our patients.

Conflicts of interest

The authors declare that they have no conflicts of interest.

References

- Mentzel T, Requena L, Brenn T. Atypical fibroxanthoma revisited. *Surg Pathol Clin.* 2017;10:319–35, <http://dx.doi.org/10.1016/j.path.2017.01.007>.
- Miller K, Goodlad JR, Brenn T. Pleomorphic dermal sarcoma: adverse histologic features predict aggressive behavior and allow distinction from atypical fibroxanthoma. *Am J Surg Pathol.* 2012;36:1317–26, <http://dx.doi.org/10.1097/PAS.0b013e31825359e1>.
- Iglesias-Pena N, López-Solache L, Martínez-Campayo N, Meilán-Sánchez I, Yebra-Pimentel MT, Balboa-Barreiro V, et al. Incidence rate and clinicopathological features of 62 atypical fibroxanthomas in a North-Western Spanish population. *Australas J Dermatol.* 2020;61:e22–7, <http://dx.doi.org/10.1111/ajd.13102>.
- Tejera-Vaquero A, Descalzo-Gallego MA, Otero-Rivas MM, Posada García C, Rodríguez-Pazos L, Pastushenko I, et al. Incidencia y mortalidad del cáncer cutáneo en España: revisión sistemática y metaanálisis. *Actas Dermosifiliogr.* 2016;107:318–28, <http://dx.doi.org/10.1016/j.ad.2015.12.008>.
- Cocharan AJ. Sentinel node biopsies. In: Calonje EJ, Brenn T, Lazar A, McKee P, editors. *McKee's Pathology of the Skin.* 4th ed Saunders; 2011.
- Lonie S, Yau B, Henderson M, Gyorki D, Angel C, Webb A. Management of pleomorphic dermal sarcoma. *ANZ J Surg.* 2020, <http://dx.doi.org/10.1111/ans.15909>.
- Harding-Jackson N, Sanguenza M, Mackinnon A, Suster S, Plaza JA. Spindle cell atypical fibroxanthoma: myofibroblastic differentiation represents a diagnostic pitfall in this variant of AFX. *Am J Dermatopathol.* 2015;37:509–16, <http://dx.doi.org/10.1097/DAD.0000000000000313>.
- Wang WL, Torres-Cabala C, Curry JL, Ivan D, McLemore M, Tetzlaff M, et al. Metastatic atypical fibroxanthoma: a series of 11 cases including with minimal and no subcutaneous involvement. *Am J Dermatopathol.* 2015;37(6):455–61, <http://dx.doi.org/10.1097/DAD.0000000000000237>.
- Müller CSL, Kreie L, Bochen F, Pfuhl T, Smola S, Gräber A, et al. Expression of 3q oncogene SEC62 in atypical fibroxanthoma-immunohistochemical analysis of 41 cases and correlation with clinical, viral and histopathologic features. *Oncol Lett.* 2019;17:1768–76, <http://dx.doi.org/10.3892/ol.2018.9767>.

10. Bitel A, Schönlebe J, Krönert C, Wollina U. Atypical fibroxanthoma: an analysis of 105 tumors. *Dermatol Ther.* 2020, <http://dx.doi.org/10.1111/dth.13962>.
11. Thum C, Husain EA, Mulholland K, Hornick JL, Brenn T. Atypical fibroxanthoma with pseudoangiomatous features: a histological and immunohistochemical mimic of cutaneous angiosarcoma. *Ann Diagn Pathol.* 2013;17:502–7, <http://dx.doi.org/10.1016/j.anndiagpath.2013.08.004>.
12. Hollmig ST, Rieger KE, Henderson MT, West RB, Sundram UN. Reconsidering the diagnostic and prognostic utility of LN-2 for undifferentiated pleomorphic sarcoma and atypical fibroxanthoma. *Am J Dermatopathol.* 2013;35:176–9, <http://dx.doi.org/10.1097/DAD.0b013e318265fb9e>.
13. Gaiser T, Hirsch D, Orouji A, Bach M, Kind P, Helbig D, et al. MYC gene amplification is a rare event in atypical fibroxanthoma and pleomorphic dermal sarcoma. *Oncotarget.* 2018;9:21182–9, <http://dx.doi.org/10.18632/oncotarget.24997>.
14. Koelsche C, Stichel D, Griewank KG, Schrimpf D, Reuss DE, Bewerunge-Hudler M, et al. Genome-wide methylation profiling and copy number analysis in atypical fibroxanthomas and pleomorphic dermal sarcomas indicate a similar molecular phenotype. *Clin Sarcoma Res.* 2019;9, <http://dx.doi.org/10.1186/s13569-019-0113-6>.
15. Nassios A, Wallner S, Haferkamp S, Klingelhöffer C, Brochhausen C, Schreml S. Expression of proton-sensing G-protein-coupled receptors in selected skin tumors. *Exp Dermatol.* 2019;28:66–71, <http://dx.doi.org/10.1111/exd.13809>.
16. Miller TI, Zoumberos NA, Johnson B, Rhodes DR, Tomlins SA, Chan MP, et al. A genomic survey of sarcomas on sun-exposed skin reveals distinctive candidate drivers and potentially targetable mutations. *Hum Pathol.* 2020;102:60–9, <http://dx.doi.org/10.1016/j.humpath.2020.06.003>.
17. Ricci C, De Leo A, Dika E, Lambertini M, Veronesi G, Corti B. Could cathepsin-k be a driver of the myofibroblastic differentiation observed in dermatofibroma, atypical fibroxanthoma and pleomorphic dermal sarcoma? *Acta Histochem.* 2020;122, <http://dx.doi.org/10.1016/j.acthis.2019.151498>.
18. Zschoche C, Hamsch C, Kutzner H, Mentzel T, Werchau S, Enk A, et al. Analysis of the lymphatic vessel architecture of atypical fibroxanthoma and pleomorphic dermal sarcoma. *J Am Acad Dermatol.* 2014;71:842–5, <http://dx.doi.org/10.1016/j.jaad.2014.04.055>.
19. Nonaka D, Bishop PW. Sarcoma-like tumor of head and neck skin. *Am J Surg Pathol.* 2014;38:956–65, <http://dx.doi.org/10.1097/PAS.0000000000000210>.
20. Griewank KG, Schilling B, Murali R, Bielefeld N, Schwamborn M, Sucker A, et al. TERT promoter mutations are frequent in atypical fibroxanthomas and pleomorphic dermal sarcomas. *Mod Pathol.* 2014;27:502–8, <http://dx.doi.org/10.1038/modpathol.2013.168>.
21. Tardío JC, Pinedo F, Aramburu JA, Suárez-Massa D, Pampín A, Requena L, et al. Pleomorphic dermal sarcoma: a more aggressive neoplasm than previously estimated. *J Cutan Pathol.* 2016;43:101–12, <http://dx.doi.org/10.1111/cup.12603>.
22. Helbig D, Ihle MA, Pütz K, Tantcheva-Poor I, Mauch C, Büttner R, et al. Oncogene and therapeutic target analyses in Atypical fibroxanthomas and pleomorphic dermal sarcomas. *Oncotarget.* 2016;7:21763–74, <http://dx.doi.org/10.18632/oncotarget.7845>.
23. Helbig D, Quaas A, Mauch C, Merkelbach-Bruse S, Büttner R, Emberger M, et al. Copy number variations in atypical fibroxanthomas and pleomorphic dermal sarcomas. *Oncotarget.* 2017;8:109457–67, <http://dx.doi.org/10.18632/oncotarget.22691>.
24. Griewank KG, Wiesner T, Murali R, Pischler C, Müller H, Koelsche C, et al. Atypical fibroxanthoma and pleomorphic dermal sarcoma harbor frequent NOTCH1/2 and FAT1 mutations and similar DNA copy number alteration profiles. *Mod Pathol.* 2018;31:418–28, <http://dx.doi.org/10.1038/modpathol.2017.146>.
25. Helbig D, Mauch C, Büttner R, Quaas A. Immunohistochemical expression of melanocytic and myofibroblastic markers and their molecular correlation in atypical fibroxanthomas and pleomorphic dermal sarcomas. *J Cutan Pathol.* 2018;45:880–5, <http://dx.doi.org/10.1111/cup.13346>.
26. Hollmig ST, Rieger KE, Henderson MT, West RB, Sundram UN. Reconsidering the diagnostic and prognostic utility of LN-2 for undifferentiated pleomorphic sarcoma and atypical fibroxanthoma. *Am J Dermatopathol.* 2013;35:176–9, <http://dx.doi.org/10.1097/DAD.0b013e318265fb9e>.
27. Polcz MM, Sebaratnam DF, Fernández-Peñas P. Atypical fibroxanthoma management: recurrence, metastasis and disease-specific death. *Australas J Dermatol.* 2018;59:10–25, <http://dx.doi.org/10.1111/ajd.12646>.
28. Cesinaro AM, Gallo G, Tramontozzi S, Migaldi M. Atypical fibroxanthoma and pleomorphic dermal sarcoma: a reappraisal. *J Cutan Pathol.* 2020, <http://dx.doi.org/10.1111/cup.13787>.
29. Soleymani T, Tyler Hollmig S. Conception and management of a poorly understood spectrum of dermatologic neoplasms: atypical fibroxanthoma, pleomorphic dermal sarcoma, and undifferentiated pleomorphic sarcoma. *Curr Treat Options Oncol.* 2017;18, <http://dx.doi.org/10.1007/s11864-017-0489-6>.
30. Hartel PH, Jackson J, Ducatman BS, Zhang P. CD99 immunoreactivity in atypical fibroxanthoma and pleomorphic malignant fibrous histiocytoma: a useful diagnostic marker. *J Cutan Pathol.* 2006;33 Suppl. 2:24–8, <http://dx.doi.org/10.1111/j.1600-0560.2006.00492.x>.
31. López L, Vélez R. Atypical fibroxanthoma. *Arch Pathol Lab Med.* 2016;140:376–9, <http://dx.doi.org/10.5858/arpa.2014-0495-RS>.
32. Winchester D, Lehman J, Tello T, Chimato N, Hocker T, Kim S, et al. Undifferentiated pleomorphic sarcoma: factors predictive of adverse outcomes. *J Am Acad Dermatol.* 2018;79:853–9, <http://dx.doi.org/10.1016/j.jaad.2018.05.022>.
33. Soleymani T, Aasi SZ, Novoa R, Hollmig ST. Atypical fibroxanthoma and pleomorphic dermal sarcoma: updates on classification and management. *Dermatol Clin.* 2019;37:253–9, <http://dx.doi.org/10.1016/j.det.2019.02.001>.
34. McCalmont TH. Correction and clarification regarding AFX and pleomorphic dermal sarcoma. *J Cutan Pathol.* 2012;39:8, <http://dx.doi.org/10.1111/j.1600-0560.2011.01851.x>.
35. McCalmont TH. AFX: what we now know. *J Cutan Pathol.* 2011;38:853–6, <http://dx.doi.org/10.1111/j.1600-0560.2011.01802.x>.
36. Elder DE, Massi D, Scolyer RA, Willemze R. WHO Classification of Skin Tumours. 4th ed International Agency for Research on Cancer (IARC); 2018.